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DEVELOPMENT AND EVALUATION OF MATRIX MATERIAL FORMULATIONS  
FOR POTENTIAL INTEGRATION INTO IMMUNODIAGNOSTIC BIOSENSORS

by

Payam Aminayi

A dissertation submitted to the Graduate College  
in partial fulfillment of the requirement  
for the degree of Doctor in Philosophy  
Chemical and Paper Engineering  
Western Michigan University  
June 2016

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# DEVELOPMENT AND EVALUATION OF MATRIX MATERIAL FORMULATIONS FOR POTENTIAL INTEGRATION INTO IMMUNODIAGNOSTIC BIOSENSORS

Payam Aminayi, Ph.D.

Western Michigan University, 2016

This study supports the development, characterization and optimization of biosensor material formulations for immunodiagnostic applications based on experimental findings and hypotheses by Wang and Wu [1, 2], and using a test-plate apparatus and thin-film design developed by Young [3].

Certain biosensors working on the basis of conductance/impedance changes have demonstrated their potential to detect various bacteria, enzymes, and biomolecules due to their enhanced electrical properties with incorporated single-walled carbon nanotubes (SWNT). From the work conducted in our laboratory, we are attempting to determine and develop a matrix formulation containing highly dispersed SWNT, antibody and other components that elicit a significantly larger change in matrix conductivity with antigen binding than would occur from only electrochemical impedance effects.

One of the most challenging aspects of using SWNT in biosensors is uniformly dispersing them within the sensing component. The dispersion of the SWNTs in an aqueous solution is particularly challenging. In aqueous solutions, cylindrical SWNTs tend to form aggregates/bundles, appearing as a micro-meshwork or a network of ropes. The resulting semi-solid solutions therefore do not often show the anticipated electrical properties after solution deposition and drying.

Studies included in this dissertation focus on obtaining uniform SWNT dispersions (using a well characterized commercially available conductive SWNT) with either poly(sodium 4-styrenesulfonate) (PSS) or carboxymethylcellulose (CMC), two dispersing agents, and other components in aqueous solutions using various sonication protocols. The high/low values used for the statistical design ranges chosen were based on preliminary work that showed roughly where SWNT dispersion was visually incomplete or dried coated matrix film resistances exceeded the limits of the test equipment used. Once characterized, SWNT dispersions were selected based on resistance level, processing attributes and film characteristics and used in a second phase of the research by combining with antibody (for future use as a capture molecule in biosensing) and glycerin (added as a wetting and perhaps antibody stabilizing agent). These SWNT dispersions together with varying levels of antibody and glycerin using a statistical design protocol were deposited within test sensor channels, dried into a semi-solid/fluid film, and resistance-tested as before.

Formulated solutions were characterized using dynamic light scattering. Dried, semi-solid/fluid composite “matrix” films deposited on test plate templates were characterized using 3D optical microscopy. Visualizing the structural aspects of the matrix film allows for a better understanding of physical factors that may affect the resistivity of the matrices, perhaps allowing for better predictability of resistance with parameter changes. Film characterization with scanning electron microscopy was also attempted, but with very limited success perhaps due to the presence of glycerin within the sample.

Printing technologies consisting of screen printing and material plotting were used to create various elements of the test plate used in these studies. From the results obtained in this study, it



was found that run 15 matrix solution (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr) resulted in dried matrix films with the lowest resistance ( $1.23 \times 10^{-02} \Omega \cdot \text{cm}$ ) and possessed an average particle size of  $\sim 115$  nm. In contrast, the largest particle size (where 81% of the particles have an average particle size of  $\sim 5000$  nm), for the set of formulation experiments in which CMC was used, was observed in Run 3 (SWNT:CMC=0.5, Sonication amplitude=20%, and sonication duration= 2 hr) in which the highest dried film resistivity was obtained. From this it was concluded that a lower SWNT:CMC ratio (lower conductive:insulating components) is contributing to larger SWNT particle size that together are contributing to higher film resistivity. The sonication factor was also found to play an important role in the dispersion and conductivity of SWNTs. The analysis of variance of the particle sizes obtained in the PSS-dispersed matrix solutions shows that the sonication amplitude and its interaction with the concentration of PSS are the only significant parameters among all that parameters were studied ( $\alpha=0.05$ ).

Profilometry of Run 3 (CMC-dispersed coating) at 100% (no dilution) showed an average roughness of  $0.274 \mu\text{m}$  while profilometry of Run 15 at 100% (no dilution) showed an average roughness of  $0.067 \mu\text{m}$ . The same trend was noted for matrix coatings at higher dilutions (70% and 40%). Therefore, increased particle size of the matrix formulations had a strong correlation with increased roughness, and increased resistance of the CMC-dispersed coatings. The thickness of the dried matrix films was found to decrease with the dilution of the matrix solution as would be expected due to reduced solids content. Calculated thickness values based on mass balance results for each prepared solution were compared with thickness measurements by the 3D optical microscope. The calculated values were found to be very close to the experimentally measured thickness measurements.

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# IMMUNODIAGNOSTIC BIOSENSORS BACKGROUND REVIEW

## 1.1 Abstract

This study supports the development, characterization and optimization of biosensor material formulations for immunodiagnostic applications based on experimental findings and hypotheses by Wang and Wu [2, 3], and using a test-plate apparatus and thin-film design developed by Young [1].

Certain biosensors working on the basis of conductance/impedance changes have demonstrated their potential to detect various bacteria, enzymes, and biomolecules due to their enhanced electrical properties with incorporated single-walled carbon nanotubes (SWNT). From the work conducted in our laboratory, we are attempting to determine and develop a matrix formulation containing highly dispersed SWNT, antibody and other components that elicit a significantly larger change in matrix conductivity with antigen binding than would occur from only electrochemical impedance effects.

One of the most challenging aspects of using SWNT in biosensors is uniformly dispersing them within the sensing component. The dispersion of the SWNTs in an aqueous solution is particularly challenging. In aqueous solutions, cylindrical SWNTs tend to form aggregates/bundles, appearing as a micro-meshwork or a network of ropes. The

resulting semi-solid solutions therefore do not often show the anticipated electrical properties after solution deposition and drying.

Studies included in this dissertation focus on obtaining uniform SWNT dispersions (using a well characterized commercially available conductive SWNT) with either poly(sodium 4-styrenesulfonate) (PSS) or carboxymethylcellulose (CMC), two dispersing agents, and other components in aqueous solutions using various sonication protocols. The high/low values used for the statistical design ranges chosen were based on preliminary work that showed roughly where SWNT dispersion was visually incomplete or dried coated matrix film resistances exceeded the limits of the test equipment used. Once characterized, SWNT dispersions were selected based on resistance level, processing attributes and film characteristics and used in a second phase of the research by combining with antibody (for future use as a capture molecule in biosensing) and glycerin (added as a wetting and perhaps antibody stabilizing agent). These SWNT dispersions together with varying levels of antibody and glycerin using a statistical design protocol were deposited within test sensor channels, dried into a semi-solid/fluid film, and resistance-tested as before.

Formulated solutions were characterized using dynamic light scattering. Dried, semi-solid/fluid composite “matrix” films deposited on test plate templates were characterized using 3D optical microscopy. Visualizing the structural aspects of the matrix film allows for a better understanding of physical factors that may affect the resistivity of the matrices, perhaps allowing for better predictability of resistance with parameter changes. Film characterization with scanning electron microscopy was also attempted, but with very limited success perhaps due to the presence of glycerin within the sample.

Printing technologies consisting of screen printing and material plotting were used to create various elements of the test plate used in these studies. From the results obtained in this study, it was found that run 15 matrix solution (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr) resulted in dried matrix films with the lowest resistance ( $1.23 \times 10^{-02} \Omega \cdot \text{cm}$ ) and possessed an average particle size of ~115 nm. In contrast, the largest particle size (where 81% of the particles have an average particle size of ~5000 nm), for the set of formulation experiments in which CMC was used, was observed in Run 3 (SWNT:CMC=0.5, Sonication amplitude=20%, and sonication duration= 2 hr) in which the highest dried film resistivity was obtained. From this it was concluded that a lower SWNT:CMC ratio (lower conductive:insulating components) is contributing to larger SWNT particle size that together are contributing to higher film resistivity. The sonication factor was also found to play an important role in the dispersion and conductivity of SWNTs. The analysis of variance of the particle sizes obtained in the PSS-dispersed matrix solutions shows that the sonication amplitude and its interaction with the concentration of PSS are the only significant parameters among all that parameters were studied ( $\alpha=0.05$ ).

Profilometry of Run 3 (CMC-dispersed coating) at 100% (no dilution) showed an average roughness of 0.274  $\mu\text{m}$  while profilometry of Run 15 at 100% (no dilution) showed an average roughness of 0.067  $\mu\text{m}$ . The same trend was noted for matrix coatings at higher dilutions (70% and 40%). Therefore, increased particle size of the matrix formulations had a strong correlation with increased roughness, and increased resistance of the CMC-dispersed coatings. The thickness of the dried matrix films was found to decrease with the dilution of the matrix solution as would be expected due to reduced solids content. Calculated thickness values based on mass balance results for each prepared solution were

compared with thickness measurements by the 3D optical microscope. The calculated values were found to be very close to the experimentally measured thickness measurements.

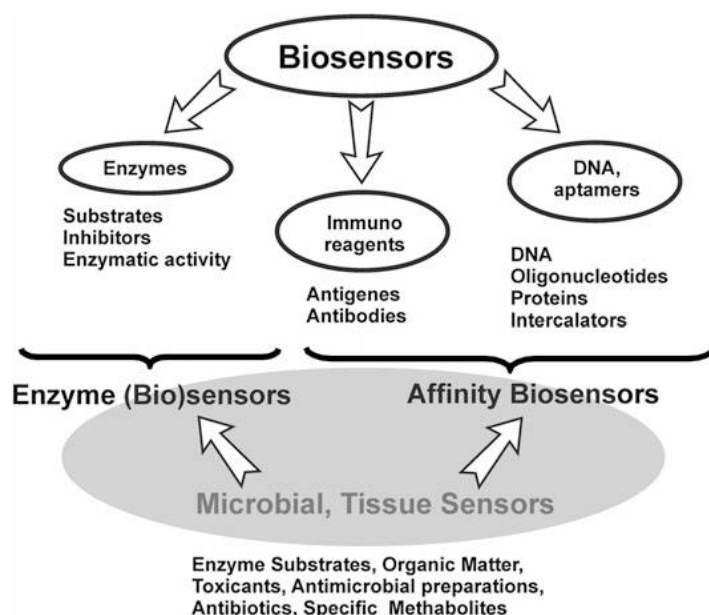
Based on the experimentally-determined higher quality and superior workability of the formulations and resulting dried matrix films, it was concluded that CMC rather than PSS was more consistent and should be used as the dispersant in the matrix formulations developed in the 2<sup>nd</sup> phase (chapter 3) of this dissertation where the addition of antibody and glycerol to the matrix was studied. Matrix formulation protocols from Run 14 (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2), Run 17 (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13), and Run 12 (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2) were selected and labeled as CMC-14, CMC-17, and CMC-12, respectively. The uniformity of the coatings prepared from CMC-12 matrix solutions were found to be the highest among the different matrix solutions tested and the superior uniformity resulted in lower resistance.

Among the three different ratio variations of SWNT:Glycerol studied with CMC-12, the highest 1:1 ratio of the three ratios tested was found to lead to the lowest electrical resistance. These levels of antibody and glycerol together with a base matrix formulation using the CMC dispersant at the ratio of SWNT:CMC=2 and the sonication amplitude of 40% for 2 hours was found to give the best properties. It is hoped that information from this research will be helpful in the design of immunodiagnostic biosensors relying on antibody-SWNT design elements.

## 1.2 Introduction

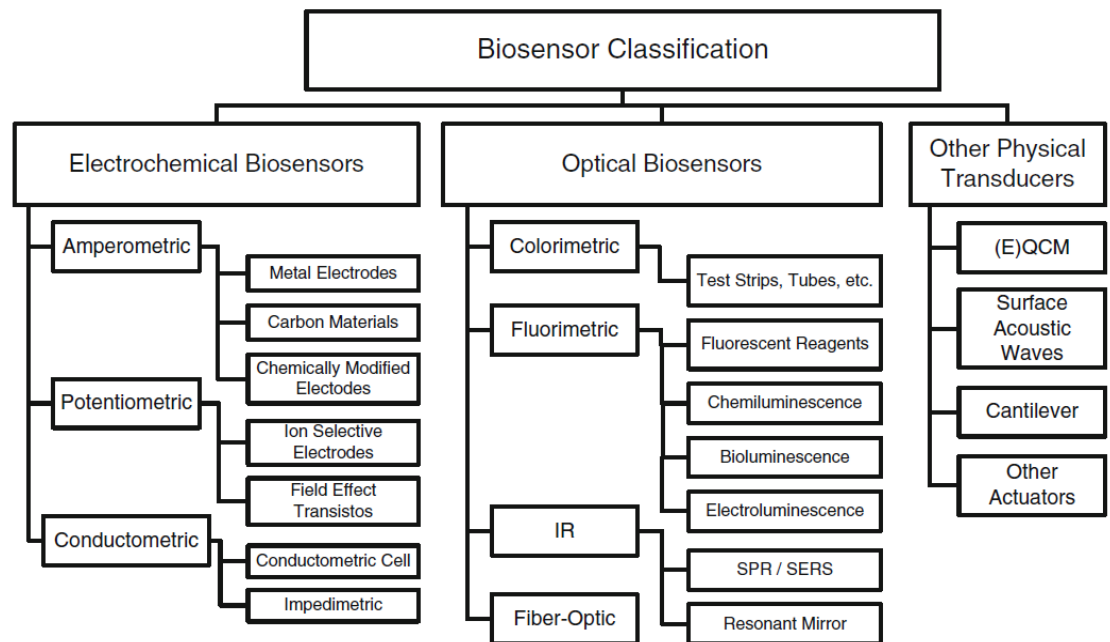
The concept of biomolecular interactions detection using a fast and versatile biosensor has gained attention in the last two decades. By definition, a device intended to detect or quantify a bio-molecule such as a cell or a particular protein is called a biosensor [4]. Biosensors can be classified into two major categories: enzyme and affinity biosensors (Figure 1). The main difference lies in the signal nature: enzyme sensors require kinetic methods of analysis in which the maximum signal intensity occurs in the first few minutes, corresponding to the substrate conversion. In contrast, affinity biosensors work on the basis of reversible biochemical interactions such as antigen–antibody or DNA–protein in which the maximum response corresponds to the equilibrium state. More complicated biochemical components lead to more variable responses, and thus it is difficult to use the term “equilibrium” for biological cells or receptors [5].





**Figure 1. Biochemical components used in the biosensor design and specific analyte determined (Courtesy of [5])**

The principal of most biosensors is based on affinity in which a functionalized and immobilized capture electrode binds the target biomolecule or analyte, selectively. This results in a change at a localized surface [6]. This change can be quantified or detected using numerous methods. One of the methods is to detect a change in impedance, voltage, or current which forms the basis of “electrical biosensors”[7] and this study. Therefore, this study excludes the different methods which employ: optical [8-12], piezoelectric [13, 14], mechanical [15, 16], and impedimetric techniques [17-20] for the detection mechanism and biosensor fabrication. Biosensors can also be classified in terms of their signal generation principles as presented in Figure 2.



**Figure 2. Classification of the biosensors in accordance with the transducer/signal transduction principles (Courtesy of [5])**

Out of the mentioned types of biosensors, the optical approach is believed to be the most accurate. However, the disadvantages of this type of biosensor are its bulkiness and high cost. Also, the optical alignment impediments have limited its functionality as a versatile, portable, and easy-to-use device. On the other hand, biosensors based on the electrochemical method have the advantages of low fabrication cost and low power consumption, which make them suitable for miniaturization. Therefore, an electrochemical biosensor is more suitable for applications where small size and cost are crucial, such as in remote areas where portable diagnostics devices are especially desirable [4]. Table 1 shows a concise list of electrochemical biosensors for cancer analysis.

**Table 1- Examples of biosensors for cancer biomarkers analysis (Courtesy of [21])**

Cancer	Biosensor		
marker	r		
detected	principle	Assay principle	Limit of detection

Table 1 - continued

<b>AFP</b>	Electroch	Protein array with 36 platinum electrodes.	–
	emical	Prussian blue with screen-printed amperometric sensor.	(range 5–500 ng ml <sup>–1</sup> )
<b>AFP and CEA</b>	Electroch	Dual-electrode with amperometric detection.	1 ng ml <sup>–1</sup>
<b>CA15-3</b>	Electroch	Antibody functionalized sol–gel film with potentiometric detection.	5 U ml <sup>–1</sup>
<b>CA125</b>	Electroch	Capillary electrophoretic.	–
	emical	Titania sol–gel on glassy carbon electrode with direct electrochemical detection of HRP.	1.29 U ml <sup>–1</sup> (range 2–14 U ml <sup>–1</sup> )
<b>CA19-9</b>	Electroch	Titania sol–gel on carbon electrode as an amperometric immunosensor.	–
<b>CEA</b>	Electroch	Faradic impedance spectroscopy using gold nanoparticle modified glassy carbon electrode.	–
		Immobilized thionine as a mediator between the electrode and HRP-labelled antibody.	(range 0.6–17 ng ml <sup>–1</sup> <comma> 17–200 ng ml <sup>–1</sup> )
		Direct electrochemical detection of HRP in an immunosensor.	0.4 ng ml <sup>–1</sup> (range 0.5–3.0 ng ml <sup>–1</sup> <comma> 3.0–120 ng ml <sup>–1</sup> )
	Optical	Chemiluminescence immunosensor.	–
<b>Ferritin</b>	Mass	Antibody immobilised on gold chip of a quartz crystal microbalance.	(range 0.1–100 ng ml <sup>–1</sup> )
	Optical	SPR based immunosensor.	–
<b>hCG</b>	Mass	A multi-channel piezoelectric quartz micro-array immunosensor.	–

	Optical	Fluorescence immunosensor.	(range 25–1500 U ml <sup>-1</sup> )
<b>PSA</b>	Electroch	Gold coated microporous membrane.	–
	emical	Amperometric disposable electrode.	0.25 ng l <sup>-1</sup>
		Capacitive immunosensor using lateral flow and impedance detection.	–
	Mass	Antibody immobilized on gold chip of a	–
	sensitive	quartz crystal microbalance.	–
		Microcantilever immunosensor.	–
		Microcantilever immunosensor.	(range 0.2–60 µg ml <sup>-1</sup> )
	Optical	SPR with colloidal gold nanoparticles.	0.15 ng ml <sup>-1</sup>

Electrical biosensors can be categorized by how the electrical measurement is made. These measurements include voltammetric, amperometric/coulometric, and impedance sensors. Impedance biosensors can be further subdivided to non-faradaic and faradaic [22]. A non-faradaic biosensor, also called a capacitive biosensor, works on the basis of change in the capacitance or non-faradaic current as a function of bio-molecular interactions. Factors that affect capacitance properties include changes in the dielectric constant, charge distribution, dimension, and shape which occur as bio-molecular interactions take place. However, the integrity of the conductive layer and the bio-recognition layer is vital for the device functionality, since any defect or pin-hole can cause a short circuit, resulting in device malfunction [23]. In contrast to a capacitance biosensor, a biosensor which works based on the faradaic current method measures the variation in the faradaic current which results from the oxidation/reduction reaction. This variation in the faradaic current is a product of steric hindrance and electrostatic repulsion [4, 24, 25]. Steric hindrance has a direct relationship with the interaction between the biological

element, the substrate and analyte, and as a result, is translated into a change in the faradaic current and impedance. In one approach, known as electrochemical impedance spectroscopy (EIS), an impedance biosensor quantifies the electrical impedance of an interface in AC steady state with constant DC bias conditions. For this purpose, usually, a sinusoidal voltage at a certain frequency is imposed, and the resulting current is measured; this experiment is repeated for a wide range of frequencies. The impedance can then be calculated as the current:voltage ratio [26]. Additionally, an increase in the number of free charges on the bio-molecule will result in an electrostatic repulsion between the bio-molecule and the redox-active species. Consequently, a change in the faradaic current will occur. Therefore, a faradaic biosensor has the ability to virtually detect molecules with any size, including large bio-entities such as cells [27] and small bio-molecules such as DNA [28, 29]. A faradaic biosensor can also measure the concentration of the analyte based on the changes in impedance. In this type of biosensor, a high frequency AC voltage is applied to films consisting of biochemical reagents and auxiliary components, such as immobilization supports, mediators, etc., and the charge transfer resistance and capacitance of the surface layer is monitored. Such measurements are generally performed in the presence of a redox buffer. Therefore a very sensitive biosensor can be developed which measures biomolecular interactions on the electrode interface and the surface layer. The drawback of such biosensors is the weak reproducibility of the results. However, unique information which in some cases cannot be obtained by other electrochemical techniques can be achieved by this method.

With impedance-based biosensors, the attachment of the biomolecule to the substrate material is often achieved by using a linker compound such as thiol or silane to

immobilize and chemically crosslink the biomolecule to the substrate [30, 31]. Other methods have also been used to trap the biomolecules physically on the substrate [32]. In either case, the impedance needs to be within the detectable range of the impedance analyzer to be able to detect the changes resulting from protein interactions. For instance, if the impedance of an immobilization layer is  $1.07\text{ M}\Omega$  in the system, and the applied voltage is  $5\text{ mV}$ , then the resulting current ( $4.67\text{ nA}$ ) will not be distinguishable from the noise in an electrical circuit. Because binding events lead to very low impedance changes, impedance-based biosensors have significant limitations.

Even though traditional methods, such as enzyme-linked immunosorbent assay (ELISA), high-performance liquid chromatography (HPLC), mass spectrometry (MS), and magnetic resonance screening are well established and sensitive for biomolecule detection, they have disadvantages. The ELISA method, for example, is time-consuming and the required reagents are expensive. All of these instruments require trained personnel to operate the tests, a serious disadvantage for use in remote areas. In addition to these impediments, none of the mentioned methods are portable, and they require a full laboratory setup. Therefore there is a pressing need for a portable, disposable, inexpensive, and sensitive device which is capable of detecting and quantifying the target biomolecules.

In a study by Wang et al., a portable and sensitive biosensor was developed by mixing an antibody specific to the microcystin-LR (MC-LR), a poisonous compound found in water, with SWNTs and dip-coating the resulting mixture on paper strips ( $5\text{cm}\times 0.5\text{cm}$ ) and freeze-drying the papers [2]. The prepared paper strips were capable of detecting  $0.6\text{ nmol/L}$  to  $40\text{ nmol/L}$  of MC-LR in the water by merely measuring the change in conductivity of the paper in solutions containing MC-LR at various concentrations. The

detection limit of this approach is comparable to that of the traditional ELISA method of MC-LR detection. The SWNT/Ab ratio used in this study was about 5000:1. The sensing mechanism was hypothesized to be a change in the gaps between carbon nanotubes upon formation of Ab-antigen complex. Since Carbon nanotubes are highly conductive, such change can affect the electron transport, and consequently provide a platform for detecting the presence of the antigen in the system. Similarly, Wu et al. [3] employed a similar method to dip coat SWNTs/antibody mixture onto paper to prepare highly sensitive impedance-based sensors used for the detection neomycin in milk samples.

The work in this dissertation is directed toward designing a conductive antibody-containing material (comprised of SWNT, antibody and other constituents), that, as a semi-solid film micro-structurally flexes upon binding antigen, and imparts a binding-extent-dependent resistivity change that can be measured that is much greater than from purely faradaic impedance. If developed, it is hoped that such a material could be incorporated into a rapid, portable, and inexpensive device for the detection of toxins and chemicals with a level of sensitivity and selectivity on par with immunoassay techniques such as ELISA.

The experiments presented in this dissertation focus on evaluating/analyzing the material properties of matrices by changing the composition of the material components together with surface analysis of the resulting matrix film. The longer term aim resulting from this thesis work is to produce a conductive antibody-containing matrix that microstructurally changes/flexes with specific antibody-antigen binding, causing significant resistivity changes in the material. Thus the prepared matrices in this study may

be eventually tested to attempt to detect antigen binding via electrical resistance measurements, beyond the scope of the work presented in this thesis.

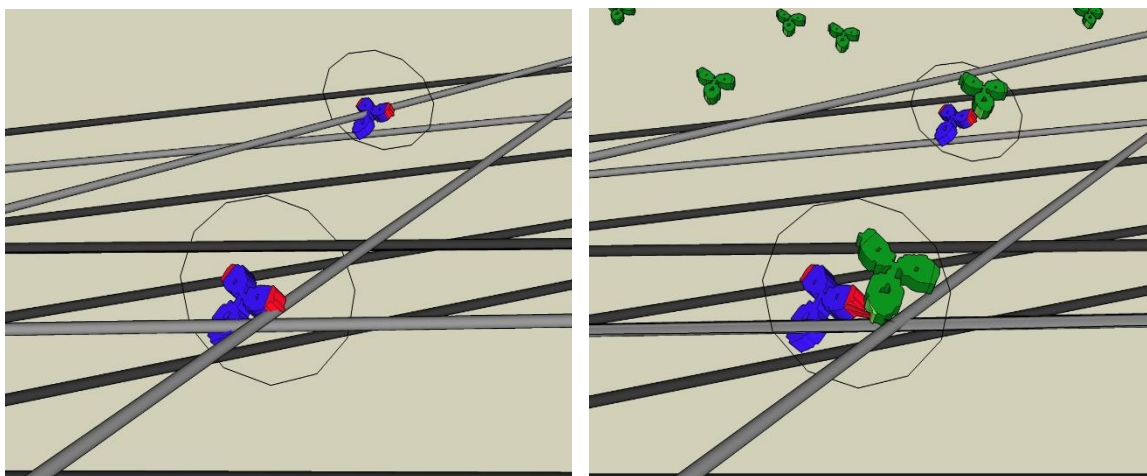
### **1.3 Theory**

The hypothesis behind this work is that under certain conditions, an antibody/SWNT matrix responds to antigen binding by changing the SWNT-to-SWNT contact distance (by expansion or swelling), resulting in a change in resistance of the matrix.

Our long-term goal in this work is to attempt to develop a thin film matrix formulation that elicits a resistance change to an applied microamp current based on the structural changes that occur during the interaction of antibodies and SWNT in the matrix system when exposed to antigen-containing solutions and as antibody-antigen binding occurs.

This conformational change can continue with time as applied antigen continues to bind to the matrix's antibodies. Therefore the current response will continue to change over time until equilibrium (constant resistance) is reached. At this stage, the current response will reach a plateau [6]. Figure 3 shows a conceptual, enlarged schematic of SWNT conformational change before and after antibody-antigen binding interactions [1]. Note SWNT-to-SWNT contacts are changed with binding, (see circled areas before and after antigen binding). Figure 3 only shows several layers of SWNT. In reality, the SWNT and antibody will coexist in many layers, preferably with interstitial spaces large enough to allow antigen to move into the interior of the SWNT:Antibody matrix to allow for a more uniform matrix resistivity change when binding antigen.





**Figure 3. Enlarged schematic of SWNT conformational change before (left) and after (right) antibody-antigen interactions [1]. Blue is matrix bound antibody, green is antigen bound to antibody (antigen depicted here is a different antibody).**

In 2009, paper test strips impregnated with an antibody:nanotube dispersion were successfully used in a standard three electrode chemical cell, with the test strip as one of the electrodes, to correlate chemical potential change of the strip with concentration of a specific toxin (antigen) in water [2]. It was hypothesized that as antigen bound to the antibody, the conductive nanotubes were pushed apart from each other reducing the conductivity of the paper strip. The most significant challenge and time spent to complete the work was in fabricating the paper test strips by dipping them in a nanotube/poly(sodium 4-styrene-sulfonate) dispersion containing antibody (specific for binding microystin-LR) followed by drying. This dipping/drying process was repeated 13 times to achieve an appropriate conductivity for the paper test strip. Although cumbersome to produce the filter paper construct, work by the authors established this as a potentially useful method for environmental testing of harmful chemicals (those chemicals that are antigenic and to which specific antibodies can be created and used in the biosensor design).

The mechanism of resistance change is hypothesized to be due to the physical micro-expansion that occurs due to mechanical forces driving conductive elements, the polymer/antibody-coated nanotubes, further apart upon binding the target molecule. Figure 3 (left) shows a graphical depiction of a thin layer of nanotubes with antibody adsorbed on the nanotube, and intercalated between different nanotubes at many junctions. The hypothesis is that when antibody that is intercalated between two different nanotubes binds antigen, the binding event perturbs or forces the two nanotubes further apart (Figure 3 (right)), thus lowering their electrical charge transfer capability from one nanotube to the next, and thus increasing the resistance of the matrix. Therefore, obtaining a uniform SWNT dispersion for use as a thin film is critical for producing the biosensors in this study and hence is a major focus of this dissertation. Uniform dispersion within an optimized formulation of dispersant agents and surfactants is a key aim.

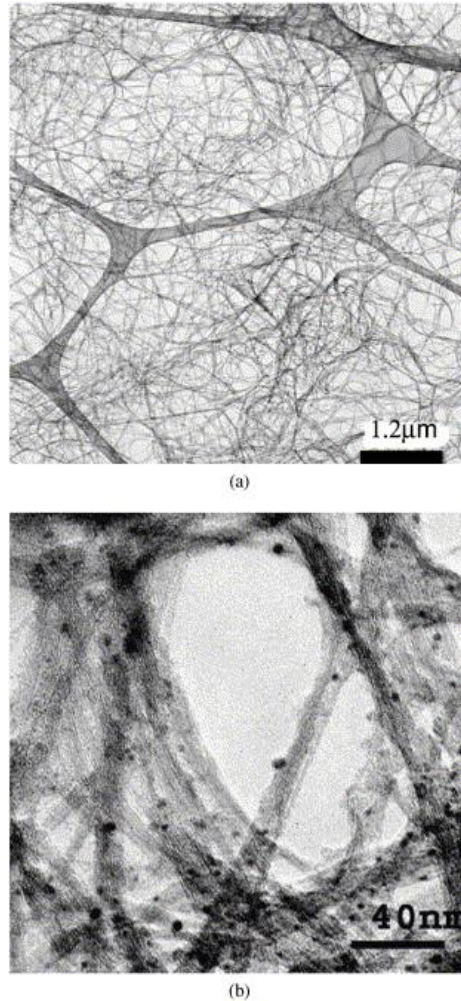
### **1.3.1 Thesis statement**

The objective of this thesis is to characterize and attain stable and uniform aqueous SWNT dispersions using sonication/centrifugation/filtration techniques (1) that when combined with antibody and wetting and surfactant elements is used to produce thin, dispersed semi-solid “matrix” films (2) for characterization and modeling (3). The properties being sought should be suitable for incorporation into a test plate for evaluation of and testing (4) to show microstructural changes due to antigen-antibody binding that can be detected as matrix film resistance changes.

### **1.3.2 SWNTs in biosensor’s matrix formulations**

Carbon nanotubes (CNTs), discovered in 1991 [33], are considered to be a new form of carbon material with unique electrical, mechanical, and chemical properties. CNTs

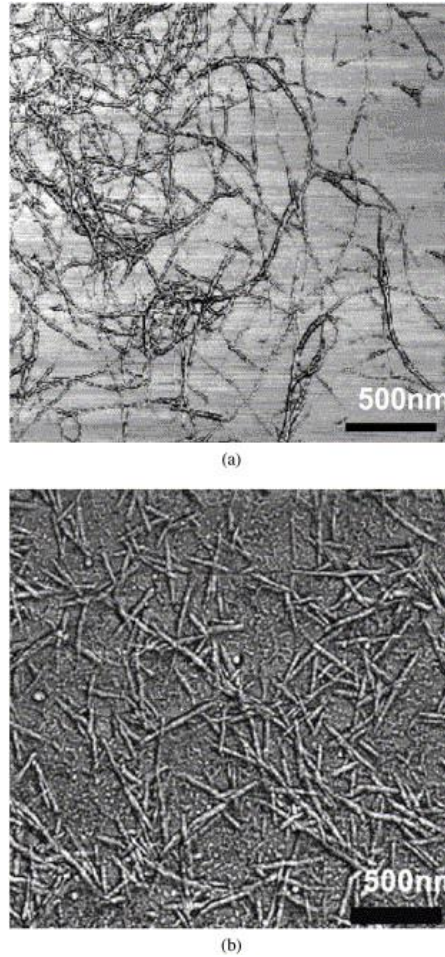
have gained attention because of their potential applications such as additives for high-strength polymer composites, electrode materials for high-capacity batteries, efficient field-emitters as electron sources, and functional components for nanoscale electronic devices [34]. SWNTs have been extensively studied over the past few years [35-38]. In the most commonly used form, atoms of carbon form planar layers of carbon that are rolled to form tubes [39, 40]. Each layer is made up of rings containing six carbon atoms. The rings are linked to each other in a hexagonal structure. Each atom has three sigma bonds with an angle of  $120^\circ$  in between and connects to three neighboring rings. The fourth electron of each atom becomes part of an extensive p bond structure. SWNTs conduct electricity because of the electrons in the p bond structure can move around throughout the graphite structure [41]. Notably, the critical challenge related to achieving the superior electrical properties for application in biosensors lie in uniformly dispersing the nanotubes. Figure 4 shows an example of SWNT aggregate morphology in the dried state.



**Figure 4. TEM image of the as-received HiPCO SWNT dispersion sample under (a) low magnification and (b) higher magnification. (Courtesy of [40])**

Because of the strong van der Waals interaction between the nanotubes, avoiding or controlling the SWNT agglomeration is difficult. It is critical to attain good dispersion to benefit from the electrical properties of the SWNTs in a network structure suitable for the biosensors. Because of the high aspect ratio of the nanotubes (the spaghetti/rope like structure), uniform nanotube dispersion is a particularly challenging problem. In addition to the long aspect ratios, lack of rigidity in nanotubes makes entanglement with complex morphology inevitable. This entanglement is extensive enough to make large and dense nanotube agglomerates in the liquid suspension visible even to the naked eye. TEM and

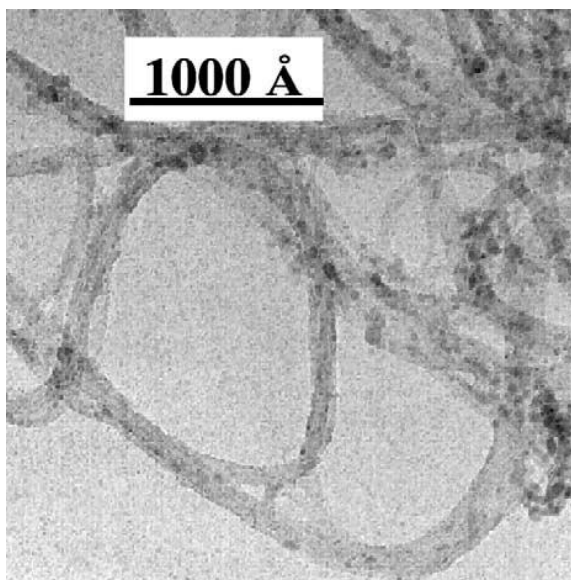
AFM images confirm the complex network structures of the entangled and intertwined nanotubes. The structure of the nanotube networks is strongly affected by the tube dimensions and colloidal conditions. Since SWNTS are insoluble in water and common solvents, sonication with the presence of surfactants or polymers is required for SWNT dispersion in aqueous solutions. Other methods have also been employed for the dispersion of SWNTs such as treating with condensed acid under sonication (Figure 5). Acid treatment results in the functionalization of the SWNT's surfaces which renders them hydrophilic at their interfacial ends. Further chemical modification is also made possible using this approach [40].



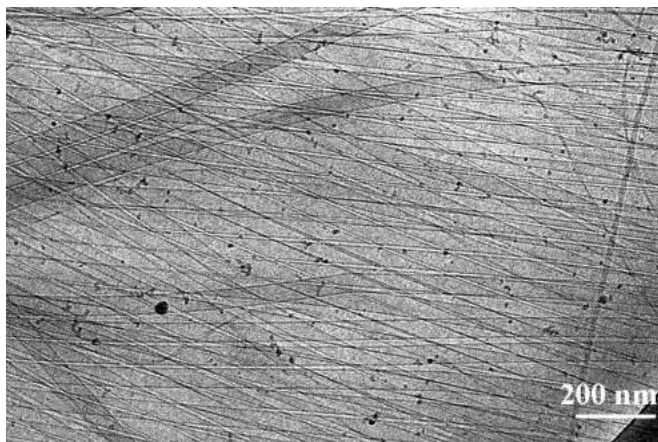
**Figure 5. 2.5×2.5  $\mu\text{m}$  AFM tapping mode scan of (a) as-received and (b) acid-treated HiPCO SWNT (Courtesy of [40])**

Characterization of the nanotube network structure in suspension is also a very challenging problem. Despite the advantages of the electron microscope and scanning probe instruments in observing the nanotubes directly (in small length scales as low as a few nanometers), there are drawbacks for using such techniques for the characterization of the nanotube network structure. The mentioned techniques require samples to be analyzed in a dried state. Therefore, they cannot be used to investigate the network structure in suspension. Additionally, these techniques require strong vacuuming which may affect the network structure, and, as a consequence, affect the statistical analysis of the dispersion.

Therefore, there is a need for non-invasive probing methods that are capable of being applied on liquid suspensions. Electromagnetic scattering has been used to study the particle dispersion. However, because of the limitation in the wavelengths that the instrument can use, the length scales of the particles analyzed are limited. Small angle X-ray scattering (SAXS) and small angle neutron scattering (SANS) are two methods which have been used for the study of nanotube dispersions [42, 43]. These methods benefit from a small probing range varying from Angstroms to few hundred nanometers and require the samples to be suspended in high-density fluids. The drawbacks of such instrumentation are its high cost and restricted availability. The range of static light scattering detection is limited to  $\sim 50$  nm to several microns, which is not suitable for the length scale of single-walled nanotube agglomerates with the size of only a few nanometers [40]. Evidently, further studies are considered necessary to establish light scattering as a reliable methodology for nanotube dispersion. The number of reports on the morphology of the SWNT is very limited, and, therefore, a need for a method to study the SWNT in suspension obviously exists. Figure 6 shows the branched rope morphology of the SWNT in the solution, and Figure 7 shows very long and uniform nanotubes that are successfully dispersed in the solution.



**Figure 6. Transmission electron micrograph of a dried suspension showing the branched rope morphology. (Courtesy of [42])**



**Figure 7. Low-magnification cryo-TEM image of a 0.1% SLC vitrified aqueous dispersion. Very long and uniform tubules form the suspension. (Courtesy of [43])**

Variations of carbon nanomaterials such as carbon nanotubes [44, 45], carbon nanofiber [46, 47], and carbon nanoparticles [48] have been used in biosensors for accurate detection of genotoxic analytes, proteins, and cells. However, due to superior chemical and electrical properties and outstanding mechanical strength, carbon nanotubes (CNTs) have specifically gained special attention [49, 50]. The CNTs can be dispersed in a variety of



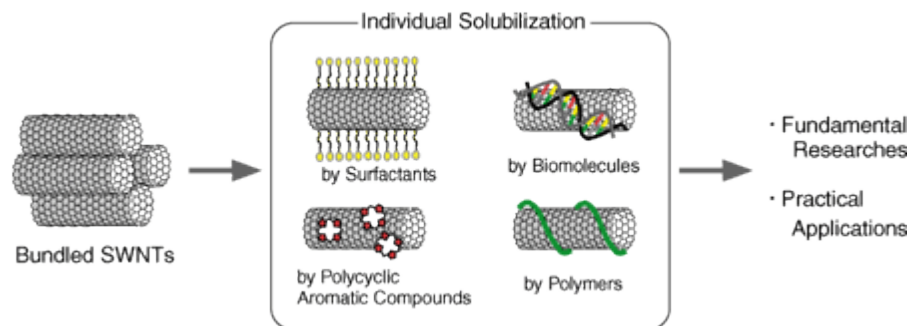
surfactants, and polymers such as the nafion, poly(3-hexylthiophene), poly(sodium 4-styrenesulfonate) (PSS), chitosan and carboxymethylcellulose (CMC) [2] which give it the functionality suitable for biosensor device manufacturing [50].

### **1.3.3 Methods of SWCNT dispersion**

There is ongoing research on single-walled carbon nanotubes (SWNTs or SWCNTs) due to their nano-scale sizes and remarkable electronic, mechanical and thermal properties [51, 52]. These properties have made SWNTs suitable for next generation electronics such as transparent electrodes, solar energy applications, supercapacitors in transistors, and batteries [53-58]. However, SWNTs tend to form bundle or aggregates in aqueous solvents [59] which inhibits the successful implementation of intended promising applications and adversely affect the functional performance properties. As the focus of this dissertation is to produce conductive thin SWCNT films, the formation of SWCNT aggregation can impede conductivity, and therefore, impede the overall performance [60].

A number of methods exist to promote the dispersion of SWNTs in aqueous solutions which can be divided into two main categories: 1- covalent (chemical) [61-63], and 2- non-covalent (physical) surface modifications [64, 65]. Chemical modifications includes nitric acid and ozone treatments [61-63, 66] in which hydrophilic functional groups are covalently bonded to SWNTs [67]. In contrast, physical treatment is accomplished by employing a surfactant which induces non-covalent adsorption of macromolecules to the SWNTs (Figure 8). The mechanism of physical modification is disruption of Van der Waals forces under sonication-induced shears [68]. It is also important to note that exposing SWNT mixtures to high shear for extended duration can play an important role in the agglomeration of SWNTs [60]. Therefore, the sonication

amplitude and the duration of the sonication should be considered when physical modification of SWNTs is being studied.



**Figure 8. Individual solubilization of single-walled carbon nanotubes (SWNTs) in water (Courtesy of [64])**

Chemical functionalization of SWNTs using covalently bonded amine ( $\text{NH}_2$ ), carboxyl ( $\text{COOH}$ ), or polyethylene glycol (PEG) functional groups enhances the ability to disperse SWNTs, but changes many of the intrinsic properties of nanotubes for biological applications [69-72]. As an example, chemical functionalization leads to surface defects which will result in a change in the surface reactivity, such as the rate at which these sites can scavenge electrons [73, 74]. Furthermore, it is believed that under UV-light exposure, chemically functionalized CNTs may lose some of their more polar (e.g., carboxyl) functional groups. Thus, non-covalent association with natural organic matter (NOM) may ultimately be responsible for CNTs dispersion in aqueous solutions [75, 76].

Physical treatment has been shown to preserve the SWNTs' fundamental properties [77, 78] due to the fact that the dispersion can be achieved through the non-covalent association with dispersants (e.g., surfactants, biomolecules, and polymers) with the CNTs. Studies show that the weak CNT-dispersant interactions have a negligible effect on the conjugated carbon arrays [79-81]. Consequently, a number of surfactants (such as Tween

and SDS) and biomolecules (such as bovine serum albumin) have been employed to disperse CNTs for biological purposes [82, 83]. It is expected that nonfunctionalized CNTs in aqueous solutions will interact with NOM, which results in an increase in their colloidal stability and bioavailability [84-86]. However, it is important to note that many conventional surfactants used to disperse SWNTs are not suitable for biological applications as they can be highly toxic [87, 88]. Therefore, biocompatible dispersants should be selected for biological applications [89]. Carboxymethyl cellulose (CMC) and Poly(sodium 4-styrenesulfonate) (PSS) are characterized by their inherent biocompatibility, low toxicity, disposability, and affordability [90, 91]. However, despite the advantages, there are only a few reports available with regards to the application of these dispersants for the dispersion of SWNTs. Therefore in this thesis study, CMC and PSS were used and compared for potential integration into immunoassay biosensors.

The downside of using physical SWNT modification is having to deal with an insulating layer of surfactant/polymer which significantly increases the resistance of thin films made of SWNTs [92, 93]. Therefore the amount of the dispersant added for the dispersion of SWNTs plays an important role in the functionality of the thin film of SWNTs for a sensing device. There is lack of a systematic approach in the literature to identify optimized conditions in which maximum connectivity of SWNTs is obtained with a completely dispersed SWNT suspension when dried in thin films. In this dissertation, experiments have been designed as a reliable method for achieving uniform, stable and highly dispersed SWNTs by employing physical modification suitable for biological applications. This study may be the first to employ a statistical approach to study, compare, model, and optimize the most suitable conditions for SWNTs dispersion.

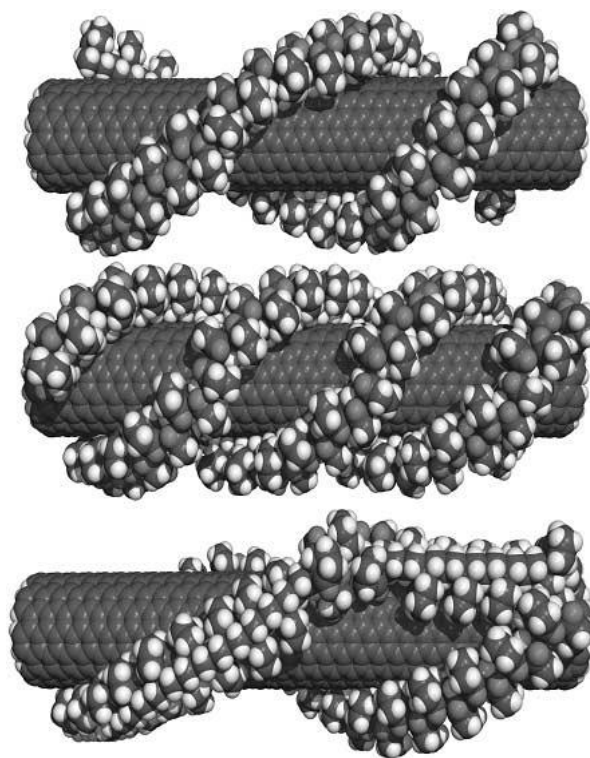
### 1.3.4 Modeling SWNT dispersion

Dispersion of SWNT in an aqueous solution has proven difficult, and the resulting matrix solutions often do not show the anticipated electrical properties. This study seeks to find the statistical relationship between the mass unbundled SWNTs in the solution, the weight ratio of dispersant to SWNT, and the resistance of the matrix film after drying. Dynamic light scattering method was employed to assess the morphology of the SWNTs dispersed in aqueous solutions (shorter and thinner SWNT bundles resulting in greater or lesser conductivity). When SWNT is dispersed in aqueous solutions using chemical processing (acid or dimethylformamide (DMF) immersion) as dispersive aids, micromeshing, or networking of ropes, which are the indicative of nanotube aggregates, are observed [94, 95]. These fractal-like aggregate morphologies can be distinguished by the relationship between the scattering wave vector and the scattered light intensity according to:

$$I(q) \sim q^{-D_f}$$

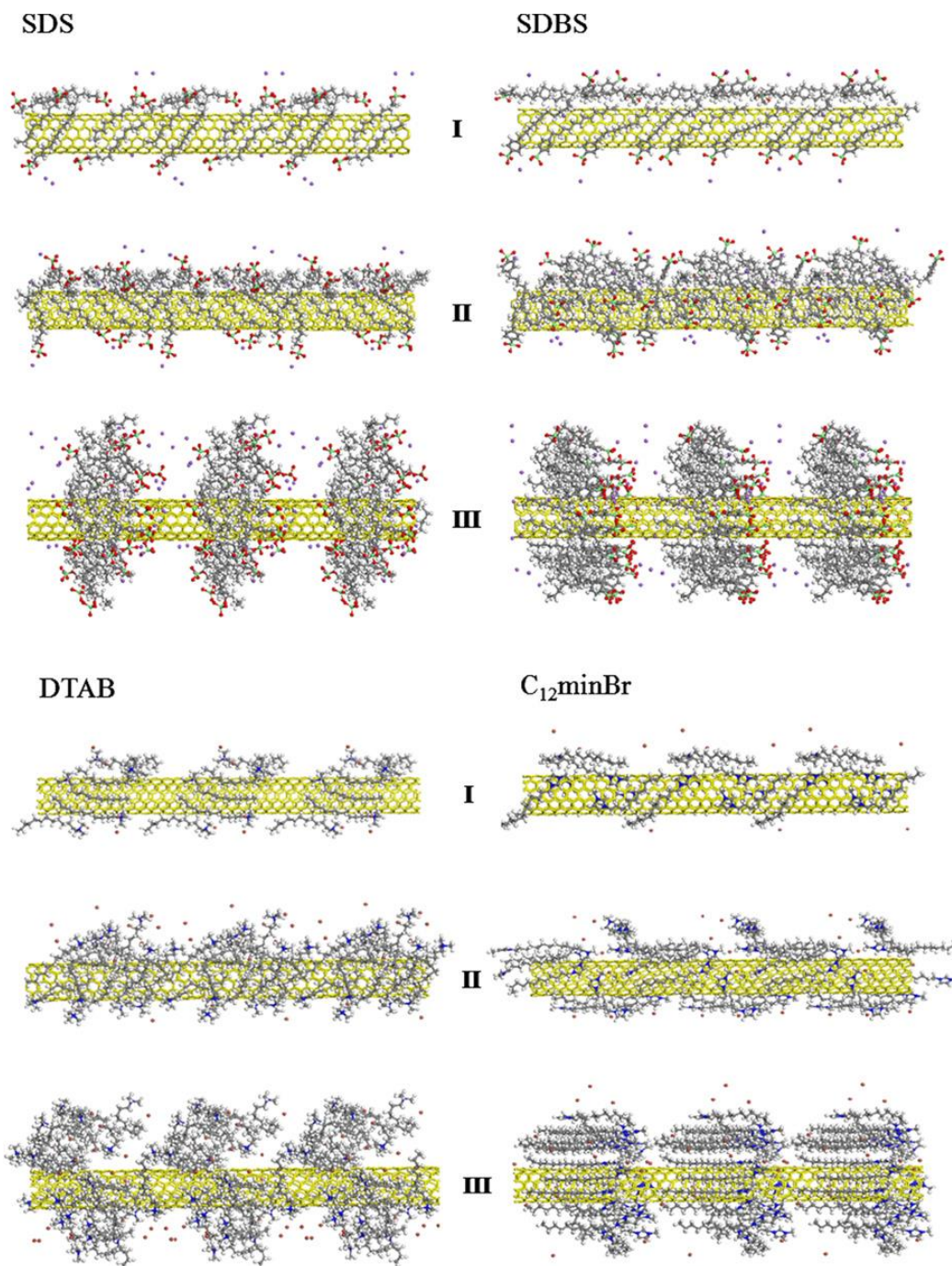
where  $I(q)$  represents the scattered light intensity and  $D_f$  represents the fractal dimension of the SWNT network. By plotting the  $\log[I(q)]$  versus  $\log(q)$ , the fractal-like morphology at the length scale of  $q^{-1}$  can be identify by determining the absolute value of the slope of the straight-line [40]. This value corresponds to the fractal dimension of the SWNT network. Higher the fractal dimension indicates the compact morphological structure of the aggregate. As a rule of thumb,  $D_f$  varies between 2.6 and 1.2, representing a very compact to a very loose, stringy agglomerate structure, respectively.

SWNTs have been dispersed with polyvinyl pyrrolidone (PVP), PVP copolymers, polystyrene sulfonate (PSSO<sub>3</sub>), PSSO<sub>3</sub> copolymers, polyvinyl sulfate, dextran, dextran sulfate, and bovine serum albumin [96]. Anionic surfactants such as sodium dodecyl sulfate (NaDS) have also been used for the nanotube dispersion [97]. In this study carboxy methyl cellulose (CMC) and poly(styrene sulfonate) (PSS) is used for the SWNT dispersion due to their compatibility with the biomolecules. The mechanism of the dispersion using these polyelectrolytes is known to be the adhesion of these long chain polymeric dispersants to the oppositely charged surfaces of nanotubes. Figure 9 shows the hypothesized simulated mechanism of dispersion using PSS and CMC.



**Figure 9. Some possible/hypothesized wrapping arrangements of PSS on a SWNT. A double helix (top) and a triple helix (middle). Backbone bond rotations can induce switch-backs, allowing multiple parallel wrapping strands to come from the same polymer chain (bottom). (Courtesy of [96])**

Molecular dynamics (MD) simulation is a useful tool to investigate the effect of dispersant concentration on the SWNT dispersion in aqueous solutions and the interactions between the SWNTs and the dispersants. It is believed that the conjugated structure in the dispersant forms multi-layers on the surface of SWNT, while the dispersants which do not have conjugation will form semi-micelles on the surface [98]. Moreover, the concentration of the dispersant will affect the dispersant layers around the headgroups. Figure 10 shows an example of different dispersants over the SWNTs surfaces.



**Figure 10.** Snapshots of surfactant/SWNT configurations in systems I, II and III. Atoms C, H, O, N and S are in grey, white, red, blue and green. Counterions are small points in figures. For clarity, water molecules are invisible and SWNT is marked in yellow. (Courtesy of [98])

In literature, one can find computer simulation studies to obtain information about the adsorption, interaction, and morphology of the dispersant over the SWNTs surface.

While SWNTs dispersion in aqueous solutions can be evaluated via experimental

investigations, obtaining the microscopic information due to adsorption and the interaction process is difficult through experimental analysis. Thus, computer simulation can be employed to investigate how dispersants wrap the SWNTs in the solution [98]. Molecular dynamics (MD) simulation is capable of simulating the wrapping of surfactants [98-100], polymers [101-103] and biological macromolecules [104-106] around the nanotubes. Simulations in literature have been carried out in a simulation program (Material Studio) at a fixed temperature (298 K) with the relaxation time of 0.2 ps. A structure optimization has been performed using a Smart Minimizer method. Target systems have been conducted for 10 ns with a time step of 1 fs at a constant volume and temperature (NVT) canonical ensemble. Partial charge of each one of the atoms of the dispersant has been calculated using a Charge Method in the simulation software (i.e Material Studio).

Total potential energy of the system was calculated using the following equation [98, 107]:

$$E_{total} = E_{bonds} + E_{angles} + E_{dihedrals} + E_{cross} + E_{non-bond}$$

The non-bond interaction will also be calculated using:

$$E_{non-bond} = \sum_{ij} \varepsilon_{ij} \left[ 2 \left( \frac{r_{ij}^0}{r_{ij}} \right)^9 - 3 \left( \frac{r_{ij}^0}{r_{ij}} \right)^6 \right] + \sum_{ij} \frac{q_i q_j}{r_{ij}}$$

where  $\varepsilon_{ij}$ ,  $r_{ij}^0$ ,  $r_{ij}$ ,  $q_i$ , and  $q_j$  are energy parameter, dimension parameter, distance between particles i and j, and the charges of i and j, respectively. Therefore, the feasibility of MD simulation in SWNT systems studies has been established and can be employed in conjunction with empirical experiments.



### **1.3.5 Preservation of bioactive elements within biosensors**

A variety of biochemical components has been used for application in biosensors such as enzymes, antibodies, proteins/peptide receptors, nucleic acids, etc. The biochemical element should be immobilized on the surface of a signal transducer to improve performance and stability. Stability of the antibody/proteins in a sensing device toward oxidants, organic solvents, digesting enzymes, etc. plays a crucial role in the point-of-care biosensors. Since, in this dissertation, the incorporation of an antibody within the sensing device is envisaged, it is also critical to preserve the functionality of the antibody in extreme application/storage conditions.

The native conformation of proteins is maintained by a balance of stabilizing and destabilizing forces leading to a stable conformation. Moreover, intermolecular forces comprised of van der Waals interactions and hydrogen bonding stabilizing the structure of the molecule. These forces are sensitive to the temperature: by increase in the temperature these interactions weaken, leading to a change in the protein conformation from the native to a denatured state [109]. Therefore, considering the protein-solvents interactions is crucial for storage of antibodies. It is accepted that numerous organisms are able to survive extreme temperatures or dehydration by producing some sugars or polyalcohols in both their intra- and intercellular fluids [110]. This process, called biopreservation, produces compounds like glycerol and trehalose that are now widely used in pharmaceutical industry [111]. Such stabilizers are added to the antibody containing matrix to reduce the degradation phenomena during processing and storage [112]. However, the mechanism by which protein is stabilized using these substances is not completely understood [109, 113-115].

Two main hypotheses exist that explain the role of stabilizers in stabilizing proteins during drying and storage. 1- “glass dynamics hypothesis” and 2- “water substitute hypothesis”. In the “glass dynamics hypothesis” a stabilizer forms a rigid, inert matrix which engulfs the molecularly dispersed protein. Therefore protein molecules are trapped and separated in the glass matrix of the stabilizer which limits their mobility and bimolecular interactions. The rigid matrix formed by the stabilizer also prohibit or hinder the unfolding and other degradations, such as chemical degradation, which require mobility. Therefore, stabilization is virtually achieved through a kinetic mechanism [116, 117]. The “water substitute hypothesis” states that the formation of hydrogen bonds between stabilizers and specific sites on the surface of the proteins is responsible for providing the thermodynamic stabilization functionality of water that is lost during drying. Therefore, the denaturation of protein, such as unfolding, is hindered thermodynamically during the drying process (for example, hydrogen bonding increases the free energy of unfolding), and increase the shelf life as the result of native structure preservation [118-120].

Regardless of mechanism, bioprotection during drying is needed to preserve the functionality of antibody via preserving the native structure of protein in the dried state [114]. The superiority of bioprotection offered by glycerol to naturally occurring biological systems has been investigated and confirmed [121]. Therefore, in this study, glycerol is used to preserve the functionality of antibody and increase the shelf life of the potential sensors. Various ratios of SWNT to glycerol were studied to investigate the effect of glycerol on the electrical resistance characteristics of the matrix coatings. Different SWNT

to antibody ratio was also examined to explore the effect of antibody on the electrical resistance characteristics.

### **1.3.6 Test plate and test system**

A laboratory test plate has been designed for the coating and “drying” of SWNT matrix films and the measuring of their resistance [1]. The plate design allows for 5 matrix channels to be tested at a time on one test plate. Figure 11, Figure 12, and Figure 13 represent this plate configuration. One formulation or multiple formulations can be tested on one plate. The A and C labels in the figures represent triplicate channels of one formulation and duplicate channels of a second formulation. The test plate design is shown from a top-angled viewing perspective in Figure 11. A side view looking parallel with the matrix channels on the plate showing the inside schematic of the deposited layers is shown in Figure 12 and Figure 13.

The single use 5-channel single-use plates are constructed by (see Figure 11):

- 1) Applying and drying conductive carbon-coated conductive silver ink electrical contacts to IPA-cleaned PET (4 separate small unfilled squares on each side of the plate). (The silver and carbon layers are screen printed, with oven drying at 100°C for 10 minutes between layers). The addition of the conductive carbon layer prevents silver electrolysis and migration from potentially interfering with the response of the device. (the carbon contact is exposed to the interior of each end of each channel and directly contacted by the matrix).

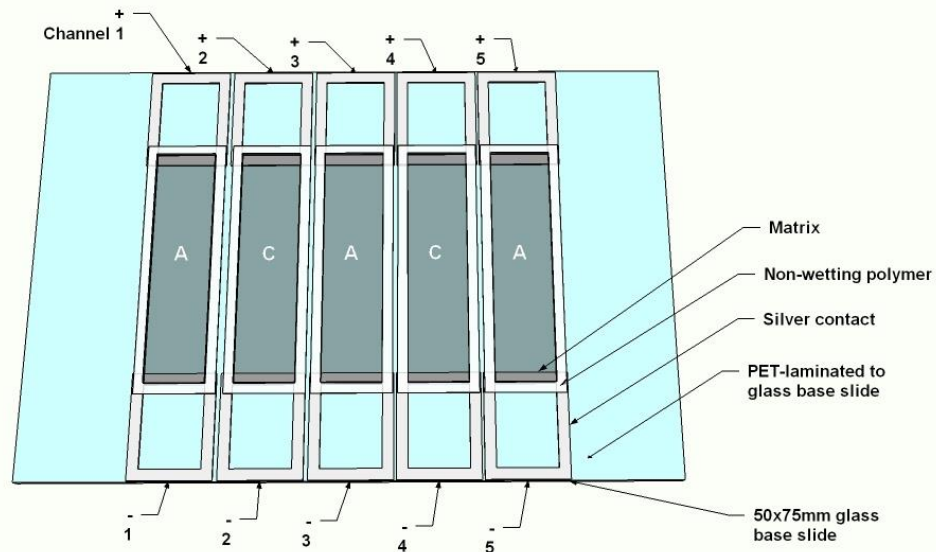
2) Applying and drying non-wetting polymer as unfilled rectangular boxes overlapping each electrical contact at the sides of the glass plate to create a total of 5 channels. This is applied using a Sonoplotter microplotter. PET templates fabricated with steps 1-4 below completed were produced in mass and stored.

3) UV-O<sub>3</sub> treating PET templates

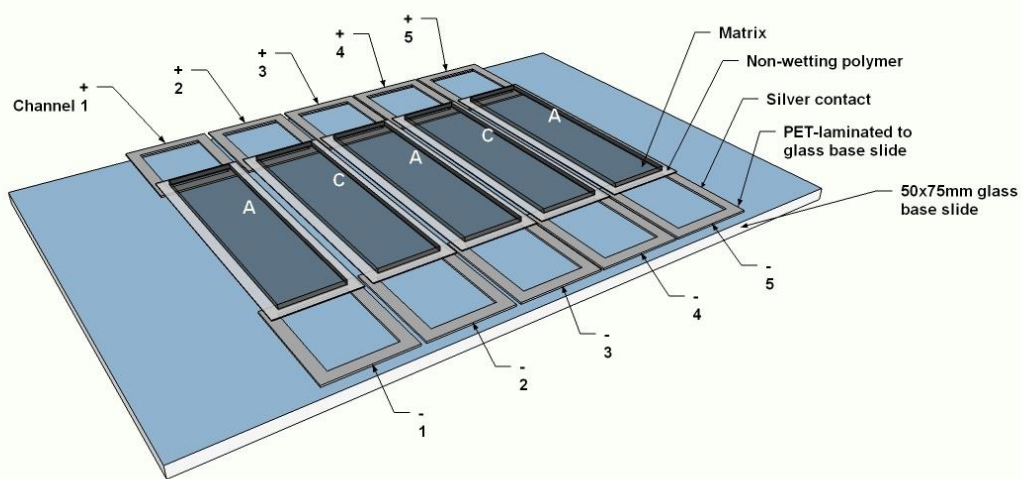
4) Laminating PET film to the ~2"x3" glass base slide.

5) Adding matrix solution to each channel (matrix is contained due to the non-wetting polymer barrier) and then “air drying” the matrix and room temperature to obtain a semi-solid film matrix.

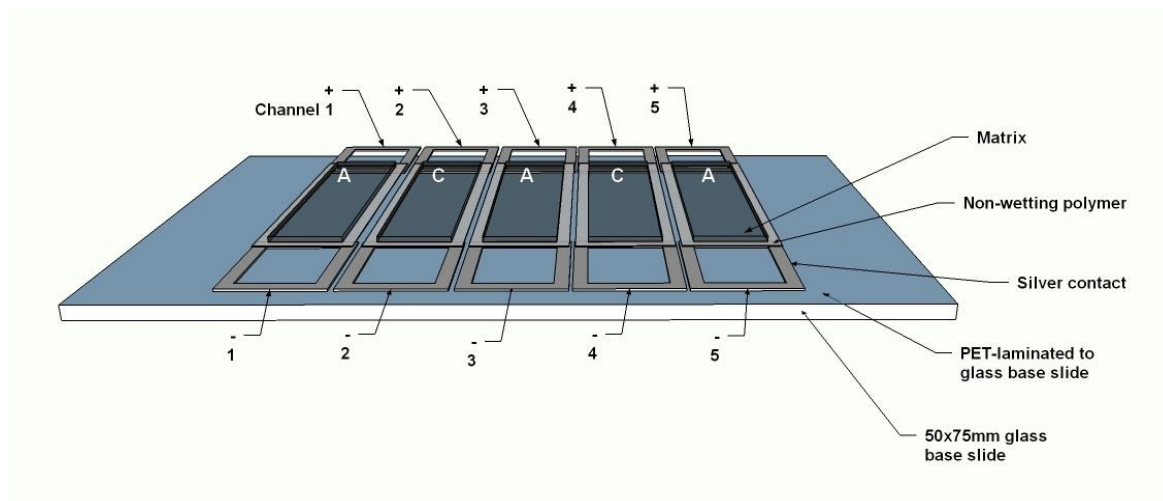
Once dry, positive and negative alligator clips are attached to each channel from a Keithley 2700/7700 Data Acquisition System (20 channel multiplexing capability) for measuring resistance of each channel as a function of time. Once the matrix solution is added to the test plate, the resistance can be measured after drying. The matrix typically dries to a semi-solid thin film within 6 hours following coating at room conditions.



**Figure 11. Schematic of five-channel test plate for testing matrix formulations. Once connected to the data acquisition system, the resistance of each matrix channel can be measured at different time intervals.**



**Figure 12. Angle view of a five-channel test plate.**



**Figure 13. Side view of a five-channel test plate.**

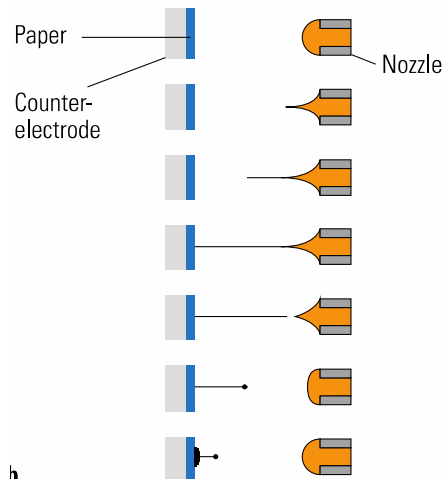
### **1.3.7 Printed sensor components**

One of the critical aspects of biosensor fabrication is designing and patterning conductive, insulating, and functional materials in a way that serves for the ultimate biosensor functionality. Different approaches have been employed to deposit these functional materials on substrates including lithography [122], chemical vapor deposition (CVD) [123], and spin coating [124]. However, these approaches are complicated, time consuming, and expensive. The direct patterning method is a technique that overcomes the drawbacks of the other techniques, consumes less material, and can be run at lower manufacturing temperatures [125, 126]. Masking the substrate (necessary in lithography) is not required, further making direct patterning an economical method.

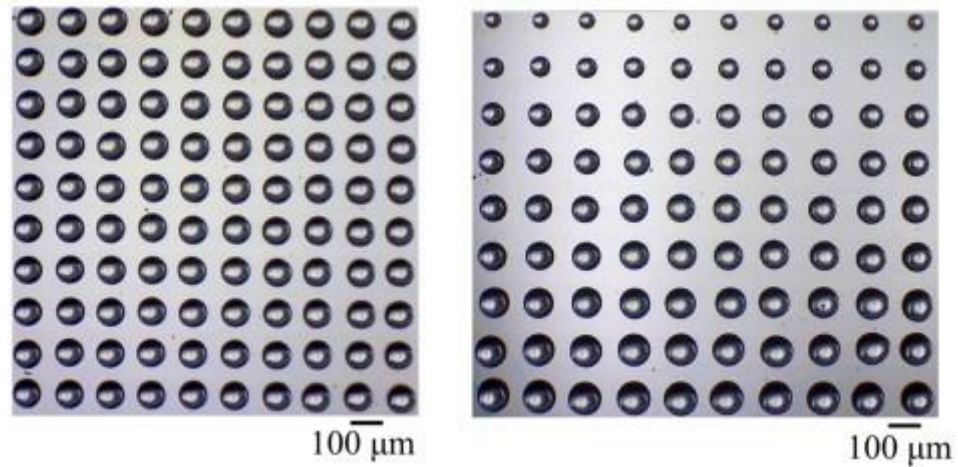
The development of printed electronics on flexible substrates, referred to as “direct printing,” has been the focus of attention in recent years. New electronic ink formulations will allow these printing processes to be cost-effective as compared with the conventional methods of electronics fabrication [127]. The benefits of the direct printing process are high material usage efficiency, elimination of expensive processing steps such as photo-

lithography, etching, and, vacuum deposition, and the ability to rapidly change circuit designs in production lines [127]. Thin and flexible substrate materials may be used, increasing their potential application in areas such as biometric device electronics. Various functional materials can be deposited accurately (printing resolution,  $\pm 5 \mu\text{m}$ ) and precisely (1–100 picoliter) where small feature patterning is desired [128].

Numerous printing methods exist such as flexography, gravure, screen printing, and inkjet printing. Out of the mentioned methods, screen printing and inkjet printing methods have gained attention for printed electronics in the past decade. Inkjet printing (Figure 14) is a relatively new technology for the fabrication of microelectronic devices. Inkjet printers use piezoelectric motion to deposit droplets of liquid materials (referred to as inks) onto the substrate [129]. Figure 15 shows a representation of droplets deposited using an inkjet printer.



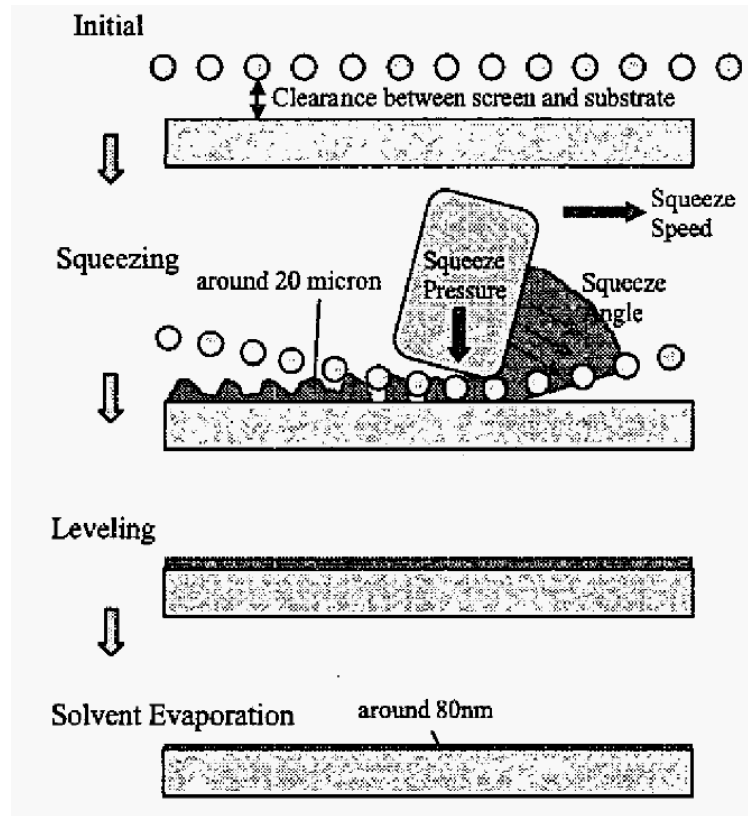
**Figure 14. DOD droplet generator. (Courtesy of [130])**



**Figure 15. Droplets deposited by DOD droplet generator. (Courtesy of [131])**

The screen printing method is also a very simple printing method which can deposit large area patterns with very high precision. The screen printing method has been also widely used for the fabrication of different types of electronic devices such as solar cells [132] and surface enhanced Raman spectroscopy (SERS) based sensors [133, 134]. Figure 16 shows a schematic of a screen printing process.





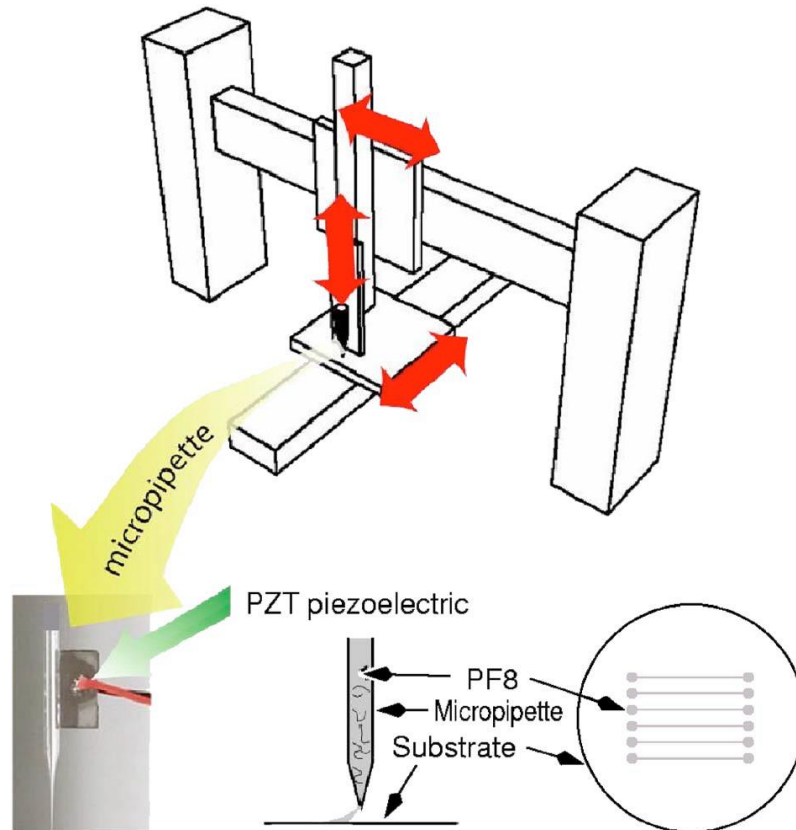
**Figure 16. Schematic of screen print deposition. (Courtesy of [132])**

In this study, a couple of printing methods were employed and for the depositing sensor materials onto surfaces for testing.

### **1.3.7.1 Microplotter printing**

Microplotters are similar in nature to the inkjet printers. However there is a significant difference between the dispensing mechanism of the Microplotter systems and the existing inkjet technology. In the Microplotter systems (Figure 17), the dispenser acts like a pen plotter, unlike the inkjet printers which eject droplets over a distance to a surface. The Microplotter directly dispenses the liquids as continuous features over the surface. The dispenser employs an ultrasonic pumping action to force the liquid out from the nozzle and

form a meniscus on the surface, from where the surface tension force takes action and dispenses the liquid where programmed.



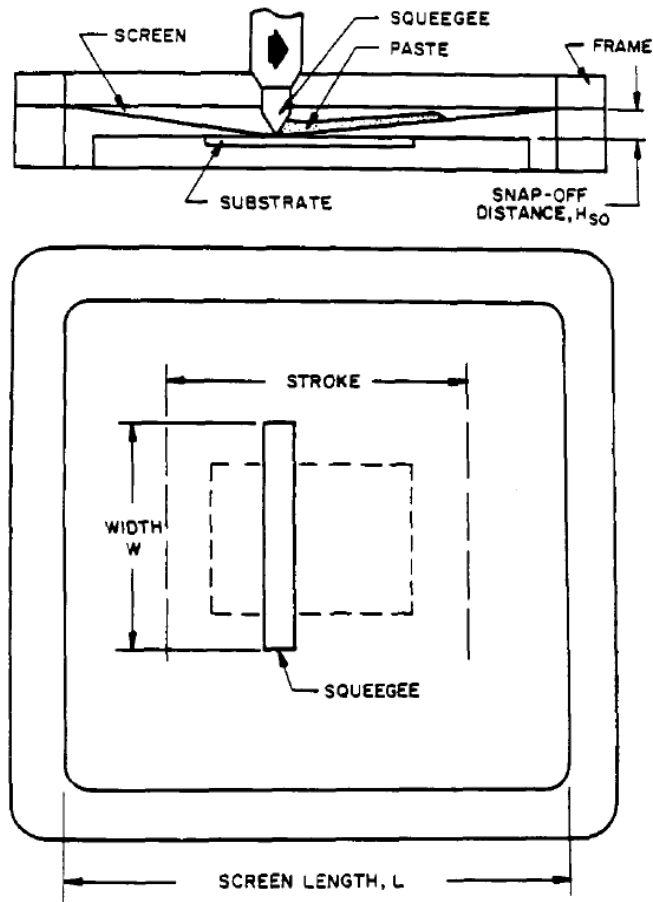
**Figure 17. Top: schematic layout of the XYZ microplotter system that provides micron positional precision. Bottom: a photograph of the micropipette/piezoelectric crystal assembly and a typical test pattern of 1 cm long lines on the sapphire substrate (Courtesy of [135])**

One of the most important characteristics of Microplotters which distinguish them from the inkjet printers is the ability to work with a much wider variety of solutions. Solutions with viscosities as high as up to 450 centipoise can be dispensed which would be impossible to print with inkjet printers. Microplotters also have the advantage of printing solutions that would normally present clogging problems to inkjet printers, like suspensions with high solid particle content such as carbon nanotube solutions. This advantage enables the Microplotters to print a variety of inks with minimum tuning as opposed to the modifications and refinement required for an inkjet printer. Another

important advantage of Microplotters is the capability to print continuous features such as lines. The objects deposited by Microplotters possess smooth edges, in contrast to objects printed by overlapping inkjet droplets. Various features with line widths as narrow as 1 micron and as wide as ~2 mm can be deposited in this method [136]. Although a microplotter was used to deposit anti-wetting polymer in this study, numerous disadvantages such as slow speed and inability to deposit certain solutions or dispersions due to clogging limits the application of this method. In this study, a GIX Microplotter II microplotter was employed to print the non-wetting features in the test template design which will be discussed later.

#### **1.3.7.2 Screen printing**

The screen printing method is an ancient technique which was introduced thousands of years ago for applications in art, textiles, and advertising. Screen printers use a woven mesh with the desired pattern and a roller or squeegee which moves across the woven screen and forces the ink past the mesh deposit over the surface of the target substrate (see Figure 19 and Figure 18). The screen mesh design (Figure 18) defines the different parts of the electrode of the biosensors. After the ink deposition, a thermal treatment is required to set the electrodes.

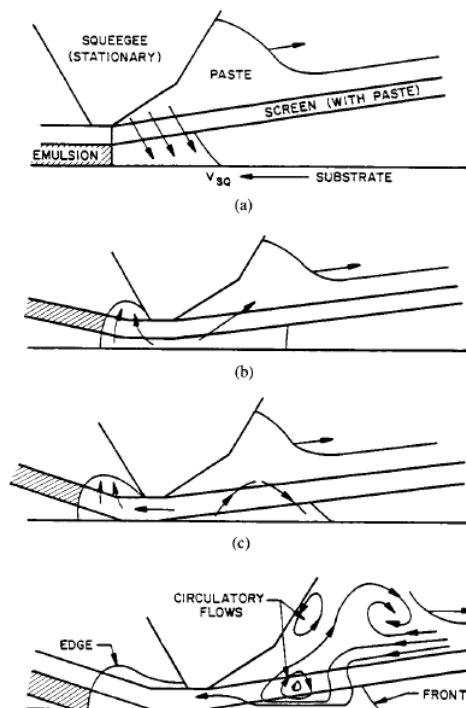


**Figure 18. Schematic of the screen print machine. (Courtesy of [137])**

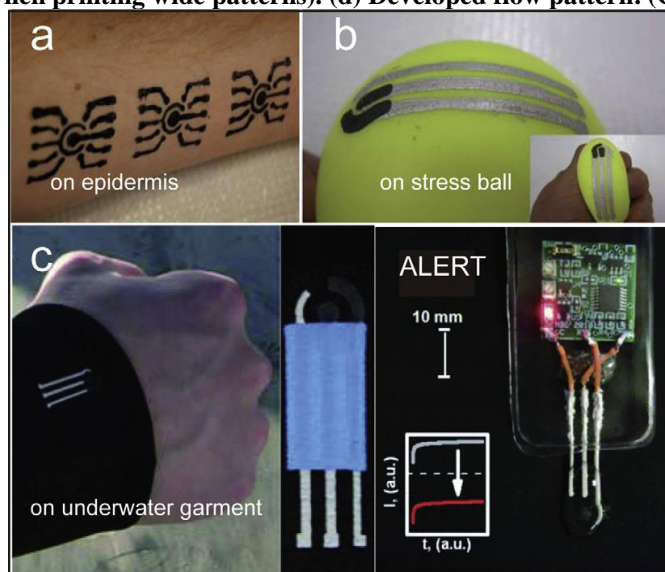
Two of the most common inks/pastes for developing devices using screen print technology are silver and carbon. Silver inks are very conductive and therefore work as the electrodes, whereas the carbon inks (less conductive) are printed over the silver inks to prevent the silver electrons from migration. Carbon inks have inert chemical characteristics and are relatively inexpensive. Gold inks are also available, however because of the high cost of such inks their application has been limited to electronics where a very high conductivity is needed, such as in enzymatic, immuno-, or genosensors [138-141].

The limitation of screen printing technology is the incompatibility with non-planar substrates. Therefore, special attention must be focused on surface treatments if a non-planar substrate is used.

The development of a flow pattern in a screen printing process is presented in Figure 19. The advancements in functional electronic inks are making conventional printing methods useful for manufacturing electronic devices. Figure 20 shows examples of how diversified and creative screen printed sensors can be. Highly sophisticated printers are now capable of miniaturizing and developing electronic devices such as highly sensitive and selective electrochemical sensors. Miniaturized sensors are very useful where the sample volume being tested is limited. Highly sensitive electrochemical sensors are capable of detecting the target compound in sample volumes as low as a few microliters. This capability allows sensors to be miniaturized while increasing the number of possible target compounds. The screen printing method is one of the powerful methods which make the fabrication of microelectrodes possible and has been the focus of attention for numerous studies in the field of sensing technology [142, 143]. The accuracy and sensitivity of the fabricated sensors using printing methods have shown to be competitive with that of the conventional microelectromechanical systems (MEMS) sensors.



**Figure 19.** Development of flow pattern in a screen printing process viewed in reference frame of the squeegee. (a) Squeegee at the edge of emulsion. (b) Squeegee begins to deflect the screen. (c) Screen touches the substrate (when printing wide patterns). (d) Developed flow pattern. (Courtesy of [137])



**Figure 20.** Photos of (a) three eight-electrode carbon ink-based arrays stamped on the epidermis and (b) three-electrode contingent on a stress ball. (c) Photo of SPE on an underwater garment (left) and a red indicative LED (right) when pollutant content in seawater exceeds a certain level. (Courtesy of [144])

Screen printing technology has also been successfully used for the fabrication of point-of-care biosensors for environmental analyses. Screen-printed electrodes (SPEs)

have been commonly developed using screen printing technology [144]. It is believed that SPEs are the most appropriate electrochemical sensors for point-of-care for electrochemical biosensors because of their linear output, low power consumption, and rapid response with high sensitivity [145, 146]. Screen-printing machines are easy to use and fast for research scales. Since the above mentioned impediments of the inkjet printing inks do not exist for the screen printers, they are also convenient to use. As a consequence, SPEs can be used in the fabrication of biosensors requiring a low detection limit.

In this study dissertation, screen printing and microplotting were employed in the fabrication of the test system/sensors in this dissertation as inkjet printing was found to have significant limitations with the materials used. Preliminary work using inkjet for a different test system was conducted and difficulty in printing SWNT-containing materials reliably required designing an entirely new laboratory biosensor test system and approach.

# SINGLE-WALLED CARBON NANOTUBES DISPERSION AND ITS EFFECT ON THE ELECTRONIC TRANSPORT PROPERTIES

## 2.1 Objective

Obtaining a uniformly dispersed SWNT within an aqueous system is a challenge for biosensor coating. Moreover, after solution deposition and drying, the resulting semi-solid materials often do not show the anticipated electrical properties. In this section, obtaining uniform SWNT dispersions in aqueous solution using various sonication protocols along with poly(sodium 4-styrenesulfonate) (PSS) and carboxymethylcellulose (CMC), two dispersing agents, and other additives comprised of phosphate buffer and sodium azide is studied. An experiment using designed using Response Surface Methodology (RSM) in order to analyze, model and predict the resistance of dried thin films of matrix solution produced under various conditions. Dispersed SWNT and aggregated/bundled SWNT present after various sonication protocols were separated by centrifugation and microfiltration, and percent dispersed was determined by weight difference of dried samples. This information was augmented with particle size distribution data using the dynamic light scattering technique. In addition, SEM was attempted and 3D optical microscopy was performed on the dried matrix films to investigate the morphology and draw correlations and trends in dispersion conditions.

Once characterized, selected SWNT dispersions were combined with antibody and other constituents (such as wetting agent glycerin) and deposited within test sensor



channels, dried into a semi-solid/fluid film, and resistance tested using a constant microamp current (see Section 3.2.1.2).

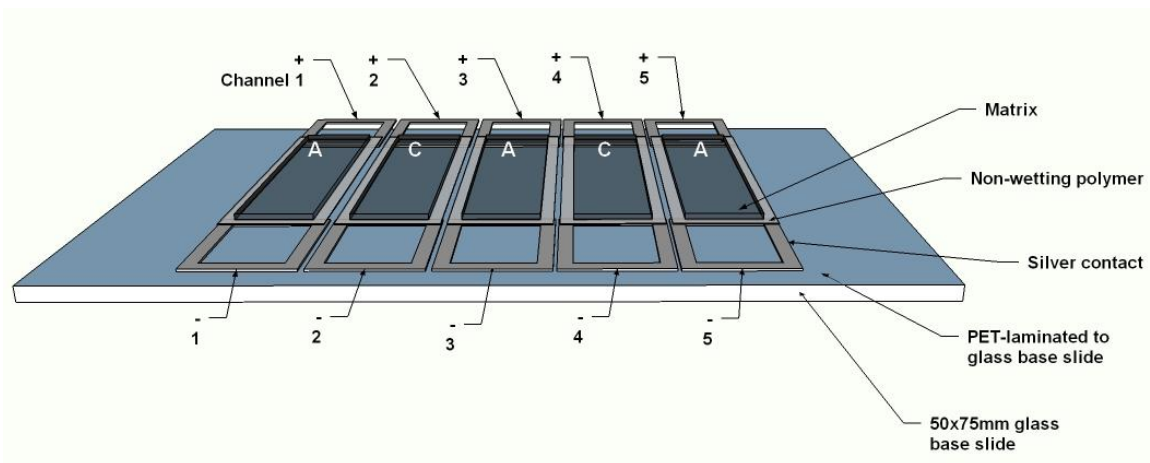
## **2.2 Methodology**

### **2.2.1 Template production procedure**

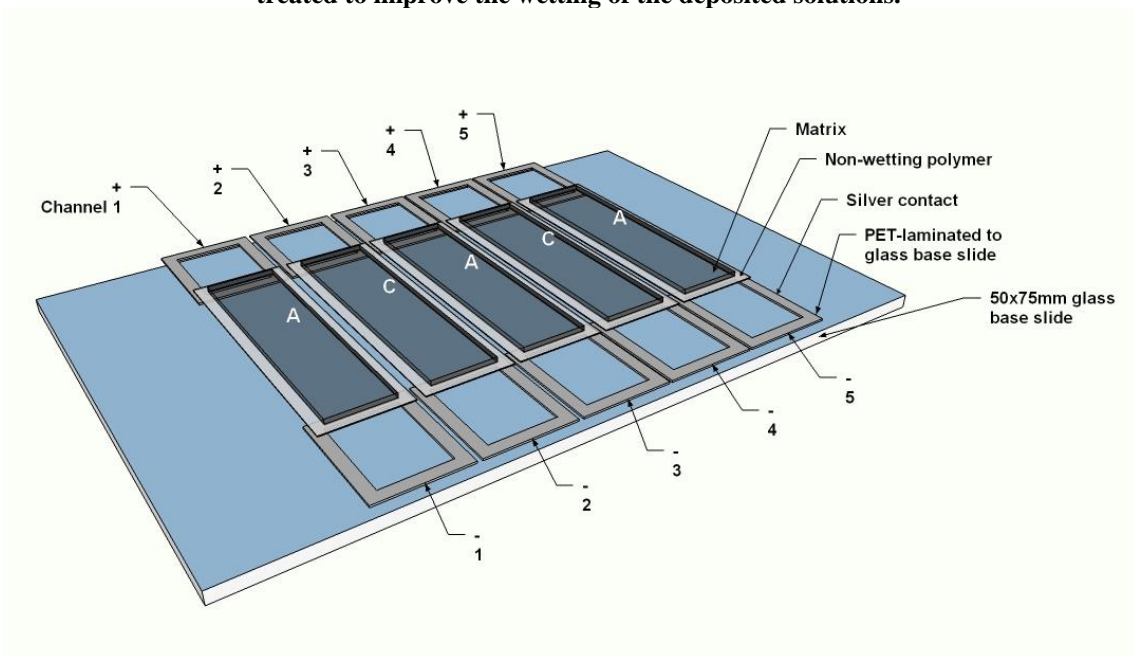
The schematic picture of a fabricated laboratory test biosensor plate is depicted in Figure 21, Figure 22 and Figure 23. The testing template was fabricated as follows (slightly modified from [1]): Conductive Ag silver ink (Electrodag 7019 from Henkel) was deposited as a 175  $\mu\text{m}$  thick electrode on DuPont Teijin Films™ (Melinex® ST505) polyester films using Western Michigan University's (WMU) MSP-485 screen printer (from Affiliated Manufacturers Inc.). The resulting Ag electrode prints were cured for 10 min at 107°C in a VWR 1320 oven. Carbon ink (Electrodag 440B) was screen-printed on the silver layer as an over-layer to protect the silver layer from electron migration. The patterns on the screen used for this layer were deliberately slightly larger than that of the silver pattern to make sure that the carbon layer completely covered the silver. The carbon layer print was then cured for 10 min at 107°C. A SonoPlot® GIX Microplotter II was used to fabricate the matrix containment channels as pictured in Figure 21. 3M™ Novec™ 2702 Electronic Grade Coating fluorochemical acrylic polymer carried in a hydrofluoroether solvent as a non-wetting polymer was used for printing the containment well. This polymer has been characterized as a low viscosity and low surface tension solution and comes in clear aqueous solution. The deposited channels were then cure at 100 °C for 1 hr for maximum attachment.

In order to confirm that a matrix solution's surface tension was appropriate for wetting the PET, a dynamic contact angle analyzer FTA200 from First Ten Angstroms

Inc., equipped with image analysis software, was used to investigate the surface energy of the PET substrate using the sessile drop method. PET was subjected to various UV-O<sub>3</sub> treatment times (UV-O<sub>3</sub>-CLEANER Model 144AX) and the optimum amount of time for UV-O<sub>3</sub> treatment (5 min) was selected. The PET sheets were then UV-O<sub>3</sub> treated for 5 minutes and attached to 75×50 mm microscope slides. 0.15 ml of matrix solution (Figure 21) was deposited in the channels (Figure 21) using a pipette and allowed to dry. Each channel was then resistance tested (Figure 23).



**Figure 21.** Side view of a five-channel test plate (adapted from [1]). The test template is UV-O<sub>3</sub> treated to improve the wetting of the deposited solutions.



**Figure 22.** Schematic of five-channel test plate (adapted from [1]) for testing matrix formulations. Once connected to data acquisition system, the resistance of each matrix channel was measured.

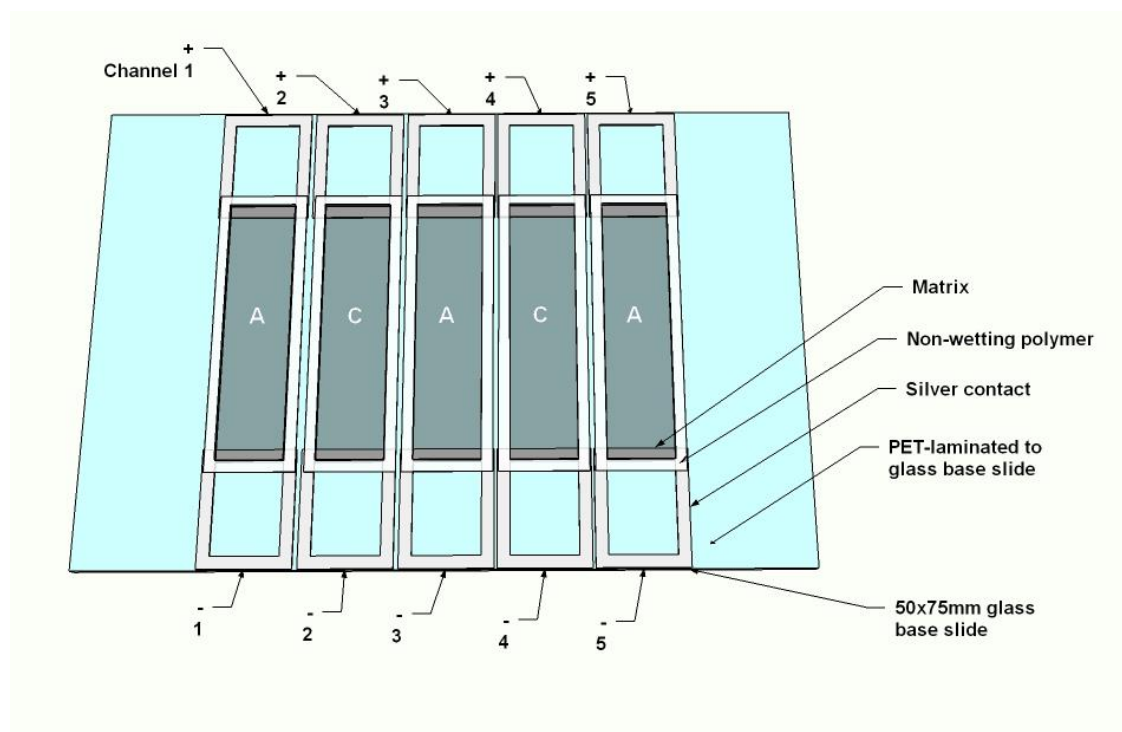


Figure 23. Top view of the various layers that are affixed to the glass base slide of a test plate (adapted from [1]).

## 2.2.2 Matrix Formulation

### 2.2.2.1 Single-walled nano-tube (SWNT) dispersion

SWNTs were dispersed with different concentrations of PSS and CMC. The formulations include SWNT and additives (phosphate buffer pH 8, and azide (at fixed concentrations of 0.07% and 0.003%, respectively)) in water. These additives are necessary to preserve the protein functionality in the future studies, which will be discussed later in this dissertation. The following parameters are investigated in this study at a constant SWNT concentration of 0.21%.

- 1) 3 SWNT to PSS or CMC weight ratios (0.5:1, 1.25:1, 2:1)
- 2) 3 Sonication amplitudes (20%, 30%, 40%)
- 3) 3 time durations (2, 13, 24 hrs)

- 4) Centrifuge at 1000 G for 1 minutes, followed by filtration through 10  $\mu\text{m}$  filter (Versapor 25 mm syringe filters).

The sensor and formulation materials used in this study are presented in Table 2.

**Table 2- The key materials for investigating the resistance response of different matrix formulations.**

<b>Materials</b>	<b>Source</b>
<b>Single Wall Nanotube (SWNT):</b>	SWeNT SG 76 (High Conductivity, 1,000 aspect ratio to tube diameter $0.92 \pm 0.27 \text{ nm}$ ), Specific Surface Area 800, Raman G/D Ratio $>16$ , Bulk Density (Tapped) $0.1 \text{ g/cm}^3$ , has both metal and semiconductor chirality
<b>Nanotube dispersant:</b>	Carboxymethylcellulose (CMC) (MW 90,000) Poly(sodium 4-styrenesulfonate) (PSS) (MW 70,000)
<b>Silver Ink:</b>	Henkel Electrodag 479SS
<b>Conductive Carbon Ink:</b>	Henkel Electrodag 965SS
<b>Fluoropolymer:</b>	3M™ Novec™ 2702 Electronic Grade Coating
<b>PET Film:</b>	Polyethylene terephthalate, Dupont Teijin Films Melinex® ST505

The sonicator used was a 130 Watt Ultrasonic Processor (Sonics & Materials, Inc.) employing a standard titanium alloy probe with tip diameter of 1/8" (3 mm) and length of 57/16" (138 mm). Each 5ml matrix solution was sonicated in a 15 ml polypropylene vial (VWR® centrifuge tubes with flat caps Cat. No. 89004-368) with the sonicator tip extended down to the vial's 2ml graduation line. The obtained suspended filtrate particles size distribution was evaluated by dynamic light scattering method (Particle Size Analyzer, Nicomp 370 and Accusizer 770) immediately after the sample preparation.

#### **2.2.2.2 Matrix formulation materials (with SWNT, PSS or CMC)**

The base composite matrix material consists of conductive nanotube, polymer, phosphate buffer, and sodium azide. The mixture is referred to as the "matrix" formulation. This matrix is coated onto a polyethylene terephthalate (PET) (DuPont Teijin Films™ Melinex® ST505) film test channels at different component ratios and film thicknesses. The purpose of this part of the study is to evaluate the coating uniformity properties and resistance of coated matrix formulations (with SWNT and PSS or CMC). A 3D optical microscope (White light interferometer ContourGT-I) was used as a profilometer to visualize surface topography, and to provide a complete 3D surface profilometric image. The morphology of the dried matrix coating was also assessed by SEM for select formulations. The purpose of the above experiments is to find matrix film formulations that have a low dried resistance (less than 2000 ohms) and to correlate resistance with film properties. Several promising matrix formulations produced from stable SWNT dispersions in this experiment were selected, and aliquots of the SWNT dispersion were mixed with solutions of antibody, and glycerol and reported in the next chapter (section 3.3).

Dispersed nanotubes in solution (0.15 ml) was deposited on test channels (see Figure 21 and Figure 22 and Figure 23) and let dry in a desiccant chamber. The test channel on which the matrix material is coated is 2.5 cm long. Three different matrix film thicknesses based on the theoretically calculated densities of the varying components in the formulations for the remaining solids (SWNT and PSS or CMC) was achieved by preparing diluted aliquots of the dispersions (as noted in section 2.2.2.1 at 100% (no

dilution with water), 70% (addition of 30% purified water), and 40 % (addition of 60% purified water) before coating.

The research test template uses a 2-wire constant current resistance measurement with a Keithley 2700/7700 multichannel data acquisition system to record matrix resistance. All matrix-coated plates were tested following drying in a desiccant chamber to reduce moisture. Test matrix resistance was assessed immediately after complete drying in the desiccant chamber.

### **2.2.3 RSM**

Response surface methodology (RSM) is an empirical statistical approach in which quantitative data is obtained from appropriately designed experiments in order to identify the best regression model and operating conditions [147, 148]. This technique employs a factorial design in order to construct mathematical models which describe the effects of several factors on the response. This statistical approach is a suitable technique for multi-factor experiments and has the advantage of identifying the most favorable conditions of the processes by determining the common relationship between various factors. Central composite design (CCD), which is a popular response surface method for the experimental design, was employed to optimize the dispersion conditions [149]. The objective of employing CCD is to optimize the effects of the variables to get the best response. This design has the following components: 1- full or fractional factorial points, 2- an additional design (a star design) at a distance of  $\alpha$  ( $\alpha=2^{(k-p)/4}$ ) calculated from the center, and 3- central points. The total number of experiments can be calculated by  $N=k^2+2k+c_p$  in which  $k$  is the factor number, and  $c_p$  is the number of replications of the experiment at the central point

[150]. For statistical calculations, the actual variables can be converted to coded variables using the relation below:

$$X_i = \frac{x_i - x_0}{\partial x} \quad (1)$$

where  $X_i$  is a coded value of the variable,  $x_i$  is the actual value of the variable,  $x_0$  is the actual value of  $X_i$  at the center point, and  $\partial x$  is the step change of the variable.

In this experiment, the design was composed of three factors (SWNT:CMC ratio, Sonication amplitude (%), and Sonication durations (hr)), each with two levels (low, high), and a total of 20 runs were carried out to optimize the chosen variables. For the purpose of statistical computations, the three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , respectively. The range of the values used for the levels of each factor investigated in the experiments was selected based on the preliminary experiments and are listed in Table 3. The quadratic equation can be used for this methodology to fit the response variables:

$$Y = \beta_0 + \sum_{i=1}^k \beta_i x_i + \sum_{i=1}^k \beta_{ii} x_i^2 + \sum_{i < j} \beta_{ij} x_i x_j \quad (2)$$

where  $Y$ ,  $k$ ,  $\beta_0$ ,  $\beta_i$ ,  $\beta_{ii}$ , and  $\beta_{ij}$  are the predicted response variable, number of variables, a constant term, coefficients of the linear parameters, coefficients of the quadratic parameters, and coefficients of the interaction parameters, respectively. The coefficient of determination ( $R^2$ ) can be estimated to test the certainty of the above polynomial model [151]. In this dissertation, RSM was employed as a useful method to investigate the effect of SWNT:CMC ratio, Sonication amplitude (%), and Sonication durations (hr) on the resistance and particle size of the prepared matrix solutions.



**Table 3- Factors and levels used in this experimental design (DOE)**

<b>Factor</b>	<b>Low level (-1)</b>	<b>High level (+1)</b>
<b>SWNT:CMC (X<sub>1</sub>)</b>	0.5	2
<b>Sonication amp (%) (X<sub>2</sub>)</b>	20	40
<b>Sonication durations (h) (X<sub>3</sub>)</b>	2	24

Different mixtures using SWNT:PSS/CMC ratios were prepared as shown in Table 4 through Table 9 and sonicated for durations varying from 0.9 hr to 25.1 hr (see Table 4 through Table 9). The amplitude of the sonicator was selected according to the design of experiment (up to 40% as presented in Table 4 through Table 9). The particle size analysis was performed immediately following the matrix solution preparation. The matrix preparation parameters were set according to the response surface methodology (RSM) experimental design, with the parameters tabulated in Table 4 through Table 9. Dilutions of the prepared solution (100%, 70%, and 40%) were prepared as explained earlier and 0.15 ml of the resulting solutions was deposited into the designated channels in duplicates. The prepared templates were then dried in a desiccant chamber for 24 hr. Subsequently, the surface profile of the select dried matrix was obtained and analyzed. The predicted resistance values using the statistical model (discussed in section 2.2.3) were also estimated and tabulated in Table 4 through Table 9. The predicted resistance values of the outlier data points are also reported and will be discussed in detail in the results section.

**Table 4- The design of experiment based on RSM showing the factor levels and their responses. The three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , which represent SWNT:CMC, Sonication amplitude (%), and Sonication durations (h), respectively for the CMC matrix solution at 100% (no dilution)**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:CMC)	<i>X<sub>2</sub></i> (Sonication amp (%))	<i>X<sub>3</sub></i> (Sonication duration (h))	<i>Resistance</i> (ohm)	<i>Predicted</i> <i>Resistance</i> ohm(eq. 5)	<i>particle</i> <i>size</i> (nm)
<i>1</i>	1.250	30	13.0	134	164	110
<i>2</i>	1.250	30	13.0	149	164	110
<i>3</i>	0.500	20	2.0	3868395	157*	4131
<i>4</i>	2.075	30	13.0	555	492	2230
<i>5</i>	1.250	30	13.0	130	164	66
<i>6</i>	0.500	20	24.0	24231	157	112
<i>7</i>	1.250	30	25.1	199	465	270
<i>8</i>	2.000	20	24.0	58878	1211*	193
<i>9</i>	0.500	40	24.0	1317	1212	932
<i>10</i>	1.250	19	13.0	149	164	313
<i>11</i>	2.000	20	2.0	10914	1211*	675
<i>12</i>	2.000	40	2.0	148	157	554
<i>13</i>	1.250	30	13.0	166	164	235
<i>14</i>	0.500	40	2.0	1069	1212	2731
<i>15</i>	2.000	40	24.0	128	157	115
<i>16</i>	1.250	30	13.0	165	164	348
<i>17</i>	1.250	30	13.0	147	164	247
<i>18</i>	0.425	30	13.0	6256	493*	213
<i>19</i>	1.250	30	0.9	794	465	409
<i>20</i>	1.250	41	13.0	225	164	180

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 5- The design of experiment based on RSM showing the factor levels and their responses. The three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , which represent SWNT:CMC, Sonication amplitude (%), and Sonication durations (h), respectively for the CMC matrix solution diluted to 70% (70%)**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:CMC)	<i>X<sub>2</sub></i> (Sonication amp (%))	<i>X<sub>3</sub></i> (Sonication durations (h))	<i>Resistance</i> (ohm)	<i>Predicted</i> <i>Resistance</i> ohm(eq. 6)	<i>particle</i> <i>size</i> (nm)
<i>1</i>	1.250	30	13.0	190	235	110
<i>2</i>	1.250	30	13.0	214	235	110
<i>3</i>	0.500	20	2.0	2812535	238	4131
<i>4</i>	2.075	30	13.0	866	744	2230
<i>5</i>	1.250	30	13.0	174	235	66
<i>6</i>	0.500	20	24.0	28272	238	112
<i>7</i>	1.250	30	25.1	254	608	270

Table 5 - continued

<b>8</b>	2.000	20	24.0	71517	<i>1690*</i>	193
<b>9</b>	0.500	40	24.0	1812	1692	932
<b>10</b>	1.250	19	13.0	210	235	313
<b>11</b>	2.000	20	2.0	15109	<i>1690*</i>	675
<b>12</b>	2.000	40	2.0	202	238	554
<b>13</b>	1.250	30	13.0	244	235	235
<b>14</b>	0.500	40	2.0	1498	1692	2731
<b>15</b>	2.000	40	24.0	200	238	115
<b>16</b>	1.250	30	13.0	223	235	348
<b>17</b>	1.250	30	13.0	195	235	247
<b>18</b>	0.425	30	13.0	8141	<i>745*</i>	213
<b>19</b>	1.250	30	0.9	1085	608	409
<b>20</b>	1.250	41	13.0	331	235	180

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 6- The design of experiment based on RSM showing the factor levels and their responses. The three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , which represent SWNT:CMC, Sonication amplitude (%), and Sonication durations (h), respectively for the CMC matrix solution diluted to 40% (40%)**

<b><i>Runs</i></b>	<b><i>X<sub>1</sub></i></b> <b><i>(SWNT:CMC)</i></b>	<b><i>X<sub>2</sub></i></b> <b><i>(Sonication amp (%))</i></b>	<b><i>X<sub>3</sub></i></b> <b><i>(Sonication durations (h))</i></b>	<b><i>Resistanc</i></b> <b><i>e (ohm)</i></b>	<b><i>Predicted</i></b> <b><i>Resistance</i></b> <b><i>ohm(eq. 7)</i></b>	<b><i>particle</i></b> <b><i>size</i></b> <b><i>(nm)</i></b>
<b>1</b>	1.250	30	13.0	362	448	110
<b>2</b>	1.250	30	13.0	410	448	110
<b>3</b>	0.500	20	2.0	4405986	<i>469*</i>	4131
<b>4</b>	2.075	30	13.0	1420	1150	2230
<b>5</b>	1.250	30	13.0	333	448	66
<b>6</b>	0.500	20	24.0	36426	<i>469*</i>	112
<b>7</b>	1.250	30	25.1	463	1028	270
<b>8</b>	2.000	20	24.0	122910	<i>2547*</i>	193
<b>9</b>	0.500	40	24.0	2838	2548	932
<b>10</b>	1.250	19	13.0	342	448	313
<b>11</b>	2.000	20	2.0	20437	<i>2547*</i>	675
<b>12</b>	2.000	40	2.0	372	468	554
<b>13</b>	1.250	30	13.0	481	448	235
<b>14</b>	0.500	40	2.0	2095	2548	2731
<b>15</b>	2.000	40	24.0	401	468	115
<b>16</b>	1.250	30	13.0	423	448	348
<b>17</b>	1.250	30	13.0	423	448	247
<b>18</b>	0.425	30	13.0	10685	<i>1152*</i>	213
<b>19</b>	1.250	30	0.9	1862	1028	409

Table 6 - continued

20	1.250	41	13.0	595	448	180
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\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 7- The design of experiment based on RSM showing the factor levels and their responses. The three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , which represent SWNT:PSS, Sonication amplitude (%), and Sonication durations (h), respectively for the PSS matrix solution at 100% (no dilution)**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:PSS)	<i>X<sub>2</sub></i> (Sonicatio n amp (%))	<i>X<sub>3</sub></i> (Sonication durations (h))	<i>Resistance</i> (ohm)	<i>Predicted</i> <i>Resistance</i> ohm(eq. 8)	<i>particle</i> <i>size</i> (nm)
1	1.250	30	13.0	459	643	206
2	1.250	30	13.0	749	643	107
3	0.500	20	2.0	6,453,796	1232*	152
4	2.075	30	13.0	304	515	15
5	1.250	30	13.0	393	643	18
6	0.500	20	24.0	16,316,438	1232*	133
7	1.250	30	25.1	163	643	155
8	2.000	20	24.0	701,788	550	53
9	0.500	40	24.0	277	286	110
10	1.250	19	13.0	2,174,028	916*	169
11	2.000	20	2.0	332,381	550	211
12	2.000	40	2.0	220	503	914
13	1.250	30	13.0	1,211	643	915
14	0.500	40	2.0	431	286	292
15	2.000	40	24.0	921	503	2880
16	1.250	30	13.0	365	643	122
17	1.250	30	13.0	1,557	643	127
18	0.425	30	13.0	560	770	110
19	1.250	30	0.9	17,817	643*	161
20	1.250	41	13.0	125	370	110

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 8- The design of experiment based on RSM showing the factor levels and their responses. The three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , which represent SWNT:PSS, Sonication amplitude (%), and Sonication durations (h), respectively for the PSS matrix solution diluted to 70%**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:PSS)	<i>X<sub>2</sub></i> (Sonicatio n amp (%))	<i>X<sub>3</sub></i> (Sonication durations (h))	<i>Resistance</i> (ohm)	<i>Predicted</i> <i>Resistance</i> ohm(eq. 9)	<i>particle</i> <i>size</i> (nm)
1	1.250	30	13.0	650	922	206
2	1.250	30	13.0	1,299	922	107
3	0.500	20	2.0	6,855,038	1932*	152

Table 8 - continued

<b>4</b>	2.075	30	13.0	390	707	15
<b>5</b>	1.250	30	13.0	544	922	18
<b>6</b>	0.500	20	24.0	16,505,225	<i>1932*</i>	133
<b>7</b>	1.250	30	25.1	241	922	155
<b>8</b>	2.000	20	24.0	1,141,095	202	53
<b>9</b>	0.500	40	24.0	366	301	110
<b>10</b>	1.250	19	13.0	4,977,047	<i>1081*</i>	169
<b>11</b>	2.000	20	2.0	509,977	<i>202*</i>	211
<b>12</b>	2.000	40	2.0	1,339	1252	914
<b>13</b>	1.250	30	13.0	1,698	922	915
<b>14</b>	0.500	40	2.0	555	301	292
<b>15</b>	2.000	40	24.0	1,484	1252	2880
<b>16</b>	1.250	30	13.0	502	922	122
<b>17</b>	1.250	30	13.0	2,094	922	127
<b>18</b>	0.425	30	13.0	819	1136	110
<b>19</b>	1.250	30	0.9	21,716	<i>922*</i>	161
<b>20</b>	1.250	41	13.0	183	762	110

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 9- The design of experiment based on RSM showing the factor levels and their responses. The three independent variables were denoted as  $x_1$ ,  $x_2$ , and  $x_3$ , which represent SWNT:PSS, Sonication amplitude (%), and Sonication durations (h), respectively for the PSS matrix solution diluted to 40% (40%)**

<b><i>Runs</i></b>	<b><i>X<sub>1</sub></i></b> <b><i>(SWNT:PSS)</i></b>	<b><i>X<sub>2</sub></i></b> <b><i>(Sonication amp (%))</i></b>	<b><i>X<sub>3</sub></i></b> <b><i>(Sonication durations (h))</i></b>	<b><i>Resistance (ohm)</i></b>	<b><i>Predicted Resistance ohm(eq. 10)</i></b>	<b><i>particle size (nm)</i></b>
<b>1</b>	1.250	30	13.0	1,464	2078	206
<b>2</b>	1.250	30	13.0	2,176	2078	107
<b>3</b>	0.500	20	2.0	14,250,756	<i>4465*</i>	152
<b>4</b>	2.075	30	13.0	770	1532	15
<b>5</b>	1.250	30	13.0	1,202	2078	18
<b>6</b>	0.500	20	24.0	25,362,920	<i>4465</i>	133
<b>7</b>	1.250	30	25.1	575	2078	155
<b>8</b>	2.000	20	24.0	7,231,889	<i>1002*</i>	53
<b>9</b>	0.500	40	24.0	801	685	110
<b>10</b>	1.250	19	13.0	12,746,253	<i>2799*</i>	169
<b>11</b>	2.000	20	2.0	1,099,619	<i>1002*</i>	211
<b>12</b>	2.000	40	2.0	2,529	2162	914
<b>13</b>	1.250	30	13.0	4,164	2078	915
<b>14</b>	0.500	40	2.0	1,100	685	292
<b>15</b>	2.000	40	24.0	2,326	2162	2880

Table 9 - continued

<b>16</b>	1.250	30	13.0	1,099	2078	122
<b>17</b>	1.250	30	13.0	5,297	2078	127
<b>18</b>	0.425	30	13.0	1,863	2625	110
<b>19</b>	1.250	30	0.9	52,246	2078*	161
<b>20</b>	1.250	41	13.0	391	1358	110

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

#### 2.2.4 Particle size analysis

Dynamic light scattering system, Nicomp, was used to measure the particle size of the prepared formulations after the filtration. For this study, the angle of detection was selected to be 90° which is recommended by the manufacturer. The purpose of using a particle size analyzer was to monitor agglomeration or entanglement caused by long nanotubes. As discussed earlier, the effect of SWNT:CMC/PSS ratio, sonication amplitude, and sonication duration on the particle size was investigated. Volume-based PSD is most sensitive to the appearance of large particle size. Therefore, in this study, volume-based size distribution was reported and analyzed.

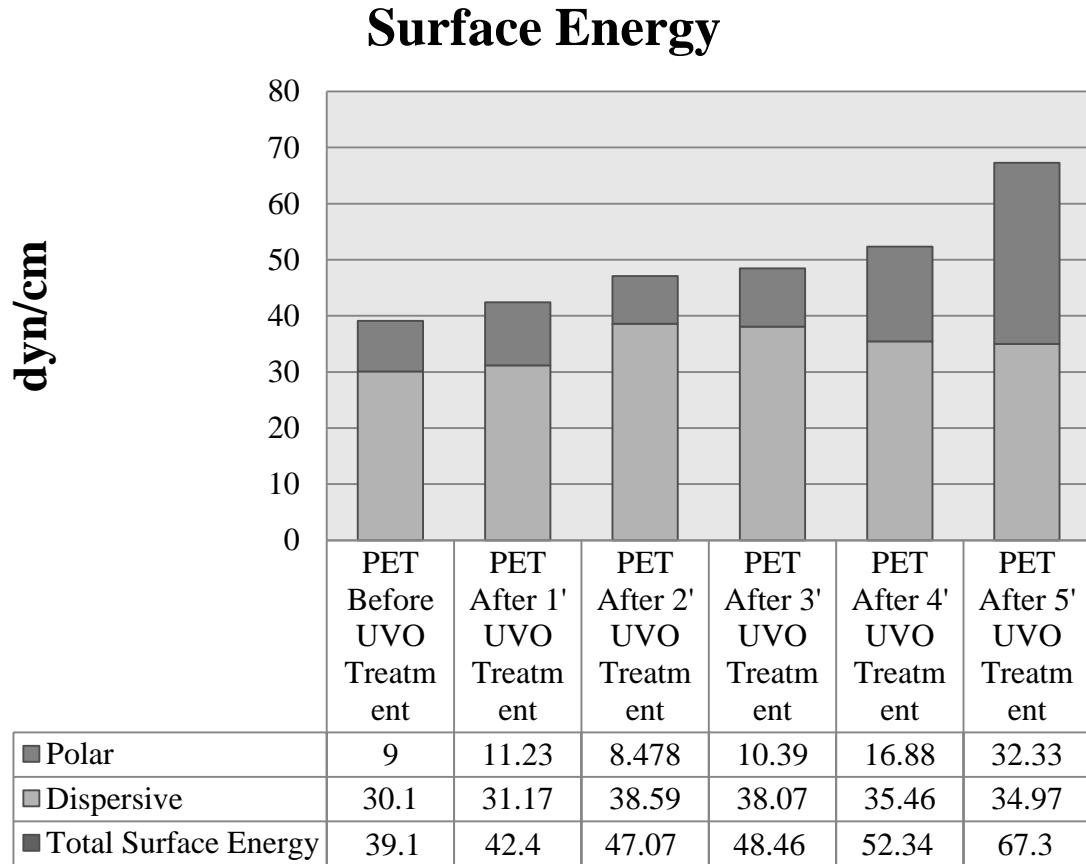
### 2.3 Results and Discussion

#### 2.3.1 Substrate modification

In order to confirm that a matrix solution surface tension was appropriate for wetting the PET, the surface energy of the PET was measured using a First Ten Angstrom, FTA200, contact angle instrument. The surface tension of the solutions was around 72 dynes/cm because the solutions were mostly constituted of water. As a rule of thumb, the surface energy of a substrate needs to be higher than (or similar to) the surface energy of the solution, a larger difference will improve the wetting [152].

Figure 24 shows the effect of a 1 through 5 minute UV-O<sub>3</sub> treatment on the surface energy of the IPA-cleaned PET. With the 5 minute UV-O<sub>3</sub> treatment, the surface energy

increased by ~25 dynes/cm for the PET by making the surfaces significantly more polar and more similar to the surface tension of the matrix solutions to allow some wetting. This showed that UV-O<sub>3</sub> treatment improved surface wetting. Five minutes of UVO exposure was used to treat all PET templates prior to coating matrix.



**Figure 24. Bar graph showing the effect of UV-O<sub>3</sub> treatment for up to 5 minutes on the surface energy of cleaned PET. The sum of the polar and dispersive components is equal to the total surface energy of the substrate.**

### 2.3.2 Resistance response analysis of variance

The dimensions of the test templates are identical for all of the experiments and therefore the resistance of a coated and dried matrix will be indicative of the physical/electrical properties of a particular matrix formulation once its final thickness is taken into account. Table 10 and Table 11 show the summary of the resistance test results

sorted by dilution percentage (100% (no dilution), 70%, and 40%), SWNT:CMC/PSS ratio (2 to 0.5), Sonication amplitude (40% to 20%), and sonication duration (24 to 2 hr). They are also color coded according to resistance (green (low) to red (high)).



Table 10. Summary of resistance test results for CMC matrix, color coded by resistance (green (low), red (high))

<i>Runs</i>	$X_1$ (SWNT:CMC)	$X_2$ (Sonication amp (%))	$X_3$ (Sonication durations (h))	<i>diluted to</i>	<i>particle size (nm)</i>	<i>Resistance (ohm)</i>	<i>Predicted Resistance (ohm)</i>	<i>Calculated coatings thickness (<math>\mu\text{m}</math>)</i>	<i>Measured coatings thickness (<math>\mu\text{m}</math>)</i>	<i>Coating resistivity (ohm.cm)</i>
15	2	40	24	100%	115	128	157	3.84	3.86	0.0123
5	1.25	30	13	100%	66	130	164	4.04	-	0.0131
1	1.25	30	13	100%	110	134	164	4.23	-	0.0141
17	1.25	30	13	100%	247	147	164	4.19	-	0.0154
12	2	40	2	100%	554	148	157	3.56	-	0.0131
2	1.25	30	13	100%	110	149	164	4.13	-	0.0153
10	1.25	19	13	100%	313	149	164	3.95	-	0.0147
16	1.25	30	13	100%	348	165	164	4.13	-	0.0170
13	1.25	30	13	100%	235	166	164	4.26	-	0.0177
7	1.25	30	25.1	100%	270	199	465	4.26	-	0.0212
20	1.25	41	13	100%	180	225	164	4.15	-	0.0234
4	2.075	30	13	100%	2230	555	492	2.26	-	0.0313
19	1.25	30	0.9	100%	409	794	465	0.85	-	0.0169
14	0.5	40	2	100%	2731	1069	1212	4.60	-	0.1230
9	0.5	40	24	100%	932	1317	1212	4.81	-	0.1580
18	0.425	30	13	100%	213	6256	493*	4.78	-	0.7470
11	2	20	2	100%	675	10914	1211*	2.55	-	0.697
6	0.5	20	24	100%	112	24231	157*	4.27	-	2.580
8	2	20	24	100%	193	58878	1211*	2.53	-	3.730
3	0.5	20	2	100%	4131	3868395	157*	3.70	3.81	357.00
5	1.25	30	13	70%	66	174	235	2.83	-	0.0123
1	1.25	30	13	70%	110	190	235	2.96	-	0.0140

Table 10 - continued

17	1.25	30	13	70%	247	195	235	2.93	-	0.0143
15	2	40	24	70%	115	200	238	2.69	2.81	0.0134
12	2	40	2	70%	554	202	238	2.49	-	0.0126
10	1.25	19	13	70%	313	210	235	2.76	-	0.0145
2	1.25	30	13	70%	110	214	235	2.89	-	0.0155
16	1.25	30	13	70%	348	223	235	2.89	-	0.0161
13	1.25	30	13	70%	235	244	235	2.98	-	0.0182
7	1.25	30	25.1	70%	270	254	608	2.98	-	0.0189
20	1.25	41	13	70%	180	331	235	2.91	-	0.0241
4	2.075	30	13	70%	2230	866	744	1.58	-	0.0342
19	1.25	30	0.9	70%	409	1085	608	0.6	-	0.0161
14	0.5	40	2	70%	2731	1498	1692	3.22	-	0.1210
9	0.5	40	24	70%	932	1812	1692	3.36	-	0.1520
18	0.425	30	13	70%	213	8141	745*	3.34	-	0.6810
11	2	20	2	70%	675	15109	1690*	1.79	-	0.6750
6	0.5	20	24	70%	112	28272	238*	2.99	-	2.1100
8	2	20	24	70%	193	71517	1690*	1.77	-	3.1700
3	0.5	20	2	70%	4131	2812535	238*	2.59	2.42	285.00
5	1.25	30	13	40%	66	333	448	1.62	-	0.0135
10	1.25	19	13	40%	313	342	448	1.58	-	0.0135
1	1.25	30	13	40%	110	362	448	1.69	-	0.0153
12	2	40	2	40%	554	372	468	1.42	-	0.0132
15	2	40	24	40%	115	401	468	1.54	1.63	0.0154
2	1.25	30	13	40%	110	410	448	1.65	-	0.0169
16	1.25	30	13	40%	348	423	448	1.65	-	0.0175
17	1.25	30	13	40%	247	423	448	1.68	-	0.0177
7	1.25	30	25.1	40%	270	463	1028	1.7	-	0.0197

Table 10 - continued

<i>13</i>	1.25	30	13	40%	235	481	448	1.7	-	0.0205
<i>20</i>	1.25	41	13	40%	180	595	448	1.66	-	0.0247
<i>4</i>	2.075	30	13	40%	2230	1420	1150	0.9	-	0.032
<i>19</i>	1.25	30	0.9	40%	409	1862	1028	0.34	-	0.0158
<i>14</i>	0.5	40	2	40%	2731	2095	2548	1.84	-	0.0963
<i>9</i>	0.5	40	24	40%	932	2838	2548	1.92	-	0.1360
<i>18</i>	0.425	30	13	40%	213	10685*	<i>1152</i>	1.91	-	0.5110
<i>11</i>	2	20	2	40%	675	20437*	<i>2547</i>	1.02	-	0.5220
<i>6</i>	0.5	20	24	40%	112	36426*	<i>469</i>	1.71	-	1.550
<i>8</i>	2	20	24	40%	193	122910*	<i>2547</i>	1.01	-	3.110
<i>3</i>	0.5	20	2	40%	4131	4405986*	<i>469</i>	1.48	1.18	104.00

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

Table 11- Summary of resistance test results for PSS matrix, color coded by resistance (green (low), red (high))

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:PSS)	<i>X<sub>2</sub></i> (Sonication amp (%))	<i>X<sub>3</sub></i> (Sonication durations (h))	<i>diluted to</i>	<i>particle size (nm)</i>	<i>Resistance (ohm)</i>	<i>Predicted Resistance (ohm)</i>	<i>Calculated coatings thickness (μm)</i>	<i>Measured coatings thickness (μm)</i>	<i>Coating resistivity (ohm.cm)</i>
<i>20</i>	1.25	41	13	100%	110	125	370	6.23	5.66	0.0194
<i>7</i>	1.25	30	25.1	100%	155	163	643	5.04	-	0.0206
<i>12</i>	2	40	2	100%	914	220	503	0.47	-	0.0026
<i>9</i>	0.5	40	24	100%	110	277	286	8.11	-	0.0561
<i>4</i>	2.075	30	13	100%	15	304	515	3.82	-	0.0291
<i>16</i>	1.25	30	13	100%	122	365	643	4.87	-	0.0444
<i>5</i>	1.25	30	13	100%	18	393	643	4.29	-	0.0422
<i>14</i>	0.5	40	2	100%	292	431	286	8.30	-	0.0894
<i>1</i>	1.25	30	13	100%	206	459	643	4.78	-	0.0549
<i>18</i>	0.425	30	13	100%	110	560	770	8.51	-	0.1190

Table 11 - continued

<b>2</b>	1.25	30	13	100%	107	749	643	4.23	-	0.0793
<b>15</b>	2	40	24	100%	2880	921	503	3.19	-	0.0735
<b>13</b>	1.25	30	13	100%	915	1211	643	4.61	-	0.1400
<b>17</b>	1.25	30	13	100%	127	1557	643	4.00	-	0.1560
<b>19</b>	1.25	30	0.9	100%	161	17817	643*	0.66	-	0.2950
<b>11</b>	2	20	2	100%	211	332381	550*	3.57	-	29.700
<b>8</b>	2	20	24	100%	53	701788	550*	3.44	-	60.400
<b>10</b>	1.25	19	13	100%	169	2174028	916*	3.71	-	202.00
<b>3</b>	0.5	20	2	100%	152	6453796	1232*	6.27	-	1010.00
<b>6</b>	0.5	20	24	100%	133	16316438	1232*	6.57	6.15	2680.00
<b>20</b>	1.25	41	13	70%	110	183	762	4.36	4.23	0.0199
<b>7</b>	1.25	30	25.1	70%	155	241	922	3.53	-	0.0212
<b>9</b>	0.5	40	24	70%	110	366	301	5.68	-	0.0519
<b>4</b>	2.075	30	13	70%	15	390	707	2.67	-	0.0261
<b>16</b>	1.25	30	13	70%	122	502	922	3.41	-	0.0428
<b>5</b>	1.25	30	13	70%	18	544	922	3.00	-	0.0409
<b>14</b>	0.5	40	2	70%	292	555	301	5.81	-	0.0807
<b>1</b>	1.25	30	13	70%	206	650	922	3.35	-	0.0544
<b>18</b>	0.425	30	13	70%	110	819	1136	5.96	-	0.1220
<b>2</b>	1.25	30	13	70%	107	1299	922	2.96	-	0.0962
<b>12</b>	2	40	2	70%	914	1339	1252	0.33	-	0.0109
<b>15</b>	2	40	24	70%	2880	1484	1252	2.23	-	0.0829
<b>13</b>	1.25	30	13	70%	915	1698	922	3.23	-	0.1370
<b>17</b>	1.25	30	13	70%	127	2094	922	2.80	-	0.1470
<b>19</b>	1.25	30	0.9	70%	161	21716	922*	0.46	-	0.2520
<b>11</b>	2	20	2	70%	211	509977	202*	2.50	-	31.900
<b>8</b>	2	20	24	70%	53	1141095	202*	2.41	-	68.800

Table 11 - continued

<b>10</b>	1.25	19	13	70%	169	4977047	<i>1081*</i>	2.60	-	323.00
<b>3</b>	0.5	20	2	70%	152	6855038	<i>1932*</i>	4.39	-	753.00
<b>6</b>	0.5	20	24	70%	133	16505225	<i>1932*</i>	4.60	4.29	1900.00
<b>20</b>	1.25	41	13	40%	110	391	1358	2.49	2.54	0.0244
<b>7</b>	1.25	30	25.1	40%	155	575	2078	2.02	-	0.0290
<b>4</b>	2.075	30	13	40%	15	770	1532	1.53	-	0.0294
<b>9</b>	0.5	40	24	40%	110	801	685	3.24	-	0.0650
<b>16</b>	1.25	30	13	40%	122	1099	2078	1.95	-	0.0535
<b>14</b>	0.5	40	2	40%	292	1100	685	3.32	-	0.0913
<b>5</b>	1.25	30	13	40%	18	1202	2078	1.72	-	0.0516
<b>1</b>	1.25	30	13	40%	206	1464	2078	1.91	-	0.0700
<b>18</b>	0.425	30	13	40%	110	1863	2625	3.41	-	0.1590
<b>2</b>	1.25	30	13	40%	107	2176	2078	1.69	-	0.0921
<b>15</b>	2	40	24	40%	2880	2326	2162	1.28	-	0.0742
<b>12</b>	2	40	2	40%	914	2529	2162	0.19	-	0.0118
<b>13</b>	1.25	30	13	40%	915	4164	2078	1.84	-	0.1920
<b>17</b>	1.25	30	13	40%	127	5297	2078	1.60	-	0.2120
<b>19</b>	1.25	30	0.9	40%	161	52246	<i>2078*</i>	0.27	-	0.3470
<b>11</b>	2	20	2	40%	211	1099619	<i>1002*</i>	1.43	-	39.300
<b>8</b>	2	20	24	40%	53	7231889	<i>1002*</i>	1.38	-	249.00
<b>10</b>	1.25	19	13	40%	169	12746253	<i>2799*</i>	1.48	-	473.00
<b>3</b>	0.5	20	2	40%	152	14250756	<i>4465*</i>	2.51	-	894.00
<b>6</b>	0.5	20	24	40%	133	25362920	<i>4465*</i>	2.63	2.46	1670.00

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

For CMC as the dispersant, the minimum resistance (average of 2 duplicates) of  $128\ \Omega$  was observed in Run 14 (see Table 4). In this run, CMC was used at 100% (no dilution). The SWNT:CMC ratio (2), sonication amplitude (40%), and sonication duration (24 hr), were all maximal for their parameter for the study. The maximum resistance of  $3.9\text{M}\Omega$  was observed when matrix was used at no dilution was noted in Run 3 (see Table 4) This corresponded to the minimum values of each parameters at: SWNT:CMC ratio of 0.5, sonication amplitude of 20%, and sonication duration of 2 hr. Thus for CMC, resistance decreases with more sonication processing (higher amplitude, longer processing) and higher SWNT concentration and ratios of SWNT (conductive):CMC (low/non-conductive), as one might logically hypothesize. The relationship is a bit more complex in that the response is the culmination of the interplay between all three parameters.

For PSS as the dispersant, the minimum resistance of  $125\ \Omega$  was noted in Run 20 (see Table 7). This corresponds to the maximum for sonication amplitude (41%), but only intermediate values of the other parameters, SWNT:PSS ratio of 1.25, and sonication duration of 13 hr, unlike what was observed for CMC. The maximum resistance of  $25\ \text{K}\Omega$  was used observed at 60% dilution was noted in Run 6 (see Table 9). This corresponds to the minimum values SWNT:PSS ratio of 0.5 and sonication amplitude of 20%, but maximum value of sonication duration of 24 hours, again unlike CMC. For PSS, the maximum and minimum resistance process conditions do not occur as one might deduce from logic.

The analysis of variance, presented in Table 12 through Table 17, shows that the resistance of the matrix solutions containing CMC as the dispersant are significantly dependent on the SWNT:CMC ratio, sonication amplitude, and sonication duration as well

as the interaction terms. Considering the t-statistics for each model term and the provisions of the RSM, the following models can be used to fit the resistance responses where  $x_1$ ,  $x_2$ , and  $x_3$ , which represent coded values of SWNT:PSS, Sonication amplitude (%), and Sonication durations (h), respectively :

CMC at 100% (no dilution):

$$\text{Resistance (ohm)} = 164.1 + 271.3 X_1 * X_1 + 249.0 X_3 * X_3 - 527.7 X_1 * X_2 \quad (5)$$

CMC diluted to 70%

$$\text{Resistance (ohm)} = 234.7 + 421 X_1 * X_1 + 309 X_3 * X_3 - 727 X_1 * X_2 \quad (6)$$

CMC at 40%

$$\text{Resistance (ohm)} = 448 + 581 X_1 * X_1 + 480 X_3 * X_3 - 1040 X_1 * X_2 \quad (7)$$

PSS at 100% (no dilution)

$$\text{Resistance (ohm)} = 2704 - 1053 X_1 - 62.3 X_2 + 30.0 X_1 * X_2 \quad (8)$$

PSS diluted to 70%

$$\text{Resistance (ohm)} = 5033 - 2941 X_1 - 126.2 X_2 + 89.4 X_1 * X_2 \quad (9)$$

PSS diluted to 40%

$$\text{Resistance (ohm)} = 11047 - 5602 X_1 - 271 X_2 + 165 X_1 * X_2 \quad (10)$$

**Table 12- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC matrix solution at 100% (no dilution)**

Source	Resistance (ohm)			Particle size (nm)	
	DF <sup>†</sup>	F-Value <sup>†</sup>	P-Value <sup>†</sup>	F-Value	P-Value
<b>Model</b>	9	3.6	0.029	2.24	0.112
<b>Linear</b>	3	4.28	0.035	2.9	0.088
<b>SWNT:CMC</b>	1	4.21	0.067	2.29	0.161
<b>Sonication amp (%)</b>	1	4.5	0.06	0.11	0.744
<b>Sonication durations</b>	1	4.13	0.069	6.3	0.031
<b>Square</b>	3	1.01	0.428	2.14	0.159
<b>SWNT:CMC*SWNT:CMC</b>	1	0.37	0.555	3.78	0.08

Table 12 - continued

Sonication amp (%)*Sonication amp (%)	1	0.36	0.56	0.01	0.935
Sonication durations*Sonication durations	1	0.36	0.56	0.07	0.799
<b>2-Way Interaction</b>	3	5.5	0.017	1.68	0.233
<b>SWNT:CMC*Sonication amp (%)</b>	1	5.45	0.042	0.02	0.878
<b>SWNT:CMC*Sonication durations</b>	1	5.66	0.039	4.14	0.069
<b>Sonication amp (%)*Sonication durations</b>	1	5.38	0.043	0.89	0.369

†DF: degree of freedom; F-value: the ratio of the variance between groups to the variance within groups; P-value: probability value

Table 13- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC matrix solution diluted to 70%

Source	DF	Resistance (ohm)		Particle size (nm)	
		F-Value	P-Value	F-Value	P-Value
<b>Model</b>	9	3.59	0.029	2.24	0.112
<b>Linear</b>	3	4.28	0.035	2.9	0.088
<b>SWNT:CMC</b>	1	4.2	0.068	2.29	0.161
<b>Sonication amp (%)</b>	1	4.51	0.06	0.11	0.744
<b>Sonication durations</b>	1	4.13	0.07	6.3	0.031
<b>Square</b>	3	1.01	0.426	2.14	0.159
<b>SWNT:CMC*SWNT:CMC</b>	1	0.38	0.553	3.78	0.08
<b>Sonication amp (%)*Sonication amp (%)</b>	1	0.36	0.56	0.01	0.935
<b>Sonication durations*Sonication durations</b>	1	0.36	0.559	0.07	0.799
<b>2-Way Interaction</b>	3	5.49	0.017	1.68	0.233
<b>SWNT:CMC*Sonication amp (%)</b>	1	5.43	0.042	0.02	0.878
<b>SWNT:CMC*Sonication durations</b>	1	5.66	0.039	4.14	0.069
<b>Sonication amp (%)*Sonication durations</b>	1	5.38	0.043	0.89	0.369

Table 14- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC matrix solution diluted to 40%

Source	DF	Resistance (ohm)		Particle size (nm)	
		F-Value	P-Value	F-Value	P-Value
<b>Model</b>	9	3.5	0.032	2.24	0.112
<b>Linear</b>	3	4.17	0.037	2.9	0.088
<b>SWNT:CMC</b>	1	3.94	0.075	2.29	0.161
<b>Sonication amp (%)</b>	1	4.76	0.054	0.11	0.744
<b>Sonication durations</b>	1	3.81	0.079	6.3	0.031
<b>Square</b>	3	1.07	0.404	2.14	0.159
<b>SWNT:CMC*SWNT:CMC</b>	1	0.41	0.538	3.78	0.08
<b>Sonication amp (%)*Sonication amp (%)</b>	1	0.38	0.552	0.01	0.935



Table 14 - continued

<b>Sonication durations*Sonication durations</b>	1	0.38	0.55	0.07	0.799
<b>2-Way Interaction</b>	3	5.26	0.02	1.68	0.233
<b>SWNT:CMC*Sonication amp (%)</b>	1	5.07	0.048	0.02	0.878
<b>SWNT:CMC*Sonication durations</b>	1	5.75	0.037	4.14	0.069
<b>Sonication amp (%)*Sonication durations</b>	1	4.97	0.05	0.89	0.369

Table 15- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the PSS matrix solution at 100% (no dilution)

Source	DF	Resistance (ohm)		Particle size (nm)	
		F-Value	P-Value	F-Value	P-Value
<b>Model</b>	9	7.01	0.003	2.55	0.081
<b>Linear</b>	3	10.57	0.002	3.47	0.059
<b>SWNT:PSS</b>	1	11.86	0.006	4.26	0.066
<b>Sonication amp (%)</b>	1	17.22	0.002	5.12	0.047
<b>Sonication durations</b>	1	2.62	0.137	1.02	0.336
<b>Square</b>	3	3.19	0.071	0.79	0.526
<b>SWNT:PSS*SWNT:PSS</b>	1	0.56	0.471	0.12	0.741
<b>Sonication amp (%)*Sonication amp (%)</b>	1	2.73	0.13	0.35	0.565
<b>Sonication durations*Sonication durations</b>	1	0.57	0.467	0.43	0.526
<b>2-Way Interaction</b>	3	7.27	0.007	3.39	0.062
<b>SWNT:PSS*Sonication amp (%)</b>	1	15.45	0.003	6.06	0.034
<b>SWNT:PSS*Sonication durations</b>	1	2.95	0.117	2.1	0.178
<b>Sonication amp (%)*Sonication durations</b>	1	3.42	0.094	2	0.187

Table 16- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the PSS matrix solution diluted to 70%

Source	DF	Resistance (ohm)		Particle size (nm)	
		F-Value	P-Value	F-Value	P-Value
<b>Model</b>	9	10.37	0.001	2.55	0.081
<b>Linear</b>	3	16.56	0.001	3.47	0.059
<b>SWNT:PSS</b>	1	15.54	0.003	4.26	0.066
<b>Sonication amp (%)</b>	1	30.65	0.001	5.12	0.047
<b>Sonication durations</b>	1	3.47	0.092	1.02	0.336
<b>Square</b>	3	5.13	0.021	0.79	0.526
<b>SWNT:PSS*SWNT:PSS</b>	1	0.19	0.669	0.12	0.741
<b>Sonication amp (%)*Sonication amp (%)</b>	1	7.89	0.019	0.35	0.565
<b>Sonication durations*Sonication durations</b>	1	0.2	0.662	0.43	0.526

Table 16 - continued

<b>2-Way Interaction</b>	3	9.43	0.003	3.39	0.062
<b>SWNT:PSS*Sonication amp (%)</b>	1	20.25	0.001	6.06	0.034
<b>SWNT:PSS*Sonication durations</b>	1	3.49	0.091	2.1	0.178
<b>Sonication amp (%)*Sonication durations</b>	1	4.54	0.059	2	0.187

Table 17- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the PSS matrix solution diluted to 40%

Source	DF	Resistance (ohm)		Particle size (nm)	
		F-Value	P-Value	F-Value	P-Value
<b>Model</b>	9	22.84	0.001	2.55	0.081
<b>Linear</b>	3	40.16	0.001	3.47	0.059
<b>SWNT:PSS</b>	1	23.06	0.001	4.26	0.066
<b>Sonication amp (%)</b>	1	90.47	0.001	5.12	0.047
<b>Sonication durations</b>	1	6.96	0.025	1.02	0.336
<b>Square</b>	3	15.04	0.001	0.79	0.526
<b>SWNT:PSS*SWNT:PSS</b>	1	0.06	0.81	0.12	0.741
<b>Sonication amp (%)*Sonication amp (%)</b>	1	28.89	0.001	0.35	0.565
<b>Sonication durations*Sonication durations</b>	1	0.07	0.795	0.43	0.526
<b>2-Way Interaction</b>	3	13.31	0.001	3.39	0.062
<b>SWNT:PSS*Sonication amp (%)</b>	1	30.04	0.001	6.06	0.034
<b>SWNT:PSS*Sonication durations</b>	1	0.76	0.403	2.1	0.178
<b>Sonication amp (%)*Sonication durations</b>	1	9.13	0.013	2	0.187

The predicted resistance from the models obtained from the prior analysis of variance, are shown in column 8 in Table 10 and Table 11. The predicted resistance values are in close agreement with the experimental results for resistance values less than 2000Ω (30 and 40 sonication amplitude data points). The surface and contour plots of the most significant process parameters (SWNT:CMC ratio, sonication amplitude, and sonication duration) are presented in Figure 25 to Figure 30. These figures show the effect of each factor on the resistance of the matrix solution. In general, intermediate to high values of

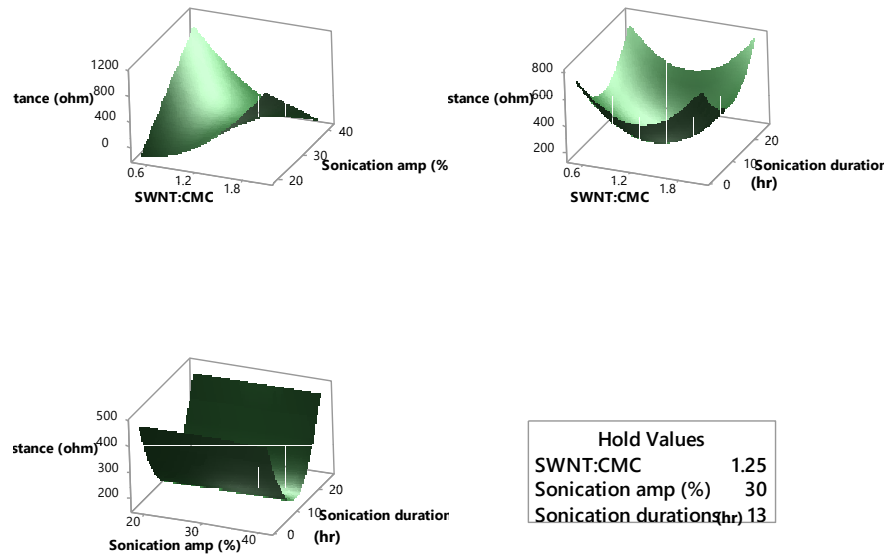
SWNT:CMC/PSS ratio and intermediate values of sonication amplitude and sonication duration provides the lowest resistance for the corresponding matrix solutions.

The effect of formulation dilution on dried matrix film resistance at low resistances appears to follow dilution closely, inversely proportional to the mass of conductor deposited per unit length. Resistance is inversely proportional to the conductor cross-sectional area. For instance the inverse of the ratio of the RSM predicted resistance values from run 5 in Table 18 for 70% dilution and 100% dilution ( $164\Omega/235\Omega$ ) yields a value of 0.698. Since the mass concentration is diluted 70%, the expected value would be 0.700, suggesting that the dried matrix at low resistances is acting like a normal conductor with regard to mass dilution. The inverse of the ratio of the RSM predicted resistance values from run 5 in Table 18 for 40% dilution and 100% dilution ( $165\Omega/333\Omega$ ) yields a value of 0.495, slight higher than the expect value 0.400.

The RSM model was not able to adequately predict dried matrix resistances at higher resistances above 2000  $\Omega$  and show these as outlier data points (even though we included them in the predicted results of the various tabulated tables). The difficulty in modeling this data is two-fold, the first and most significant is that this corresponds to all formulations performed at 20% sonication amplitude (the low level of the model for sonication). For instance, in Table 10, the four formulations (11, 6, 8, and 3) at 100% strength (dilution) in the section highlighted in red are the only 20% amplitude sonications run for this group and show dramatic effects (very large resistant changes) and in several cases opposite trends than what was observed in the low resistance data. Data sets 11 and 8 show a decrease in resistance with increasing sonication duration and data sets 6 and 3 show an increase in resistance with increasing sonication duration. These are opposing

trends just in this limited set. As a result it is difficult to model this data in the absence of a much larger experimental design (the second issue). Such a problem did not occur for modeling of resistances in the second phase of this research that is discussed in Chapter 3 where 20% sonication amplitudes were not used and where the model predicts resistance much better over a very large range of resistances. For this first research phase (Chapter 2), the models will focus on the lower resistance range due to the phenomena encountered with the lowest sonication amplitude that apparently is unpredictably affecting how the SWNT is being wrapped/not wrapped and dispersed/not dispersed by the CMC/PSS.

### Three-dimensional response surface plots



### Contour plots of the resistance responses

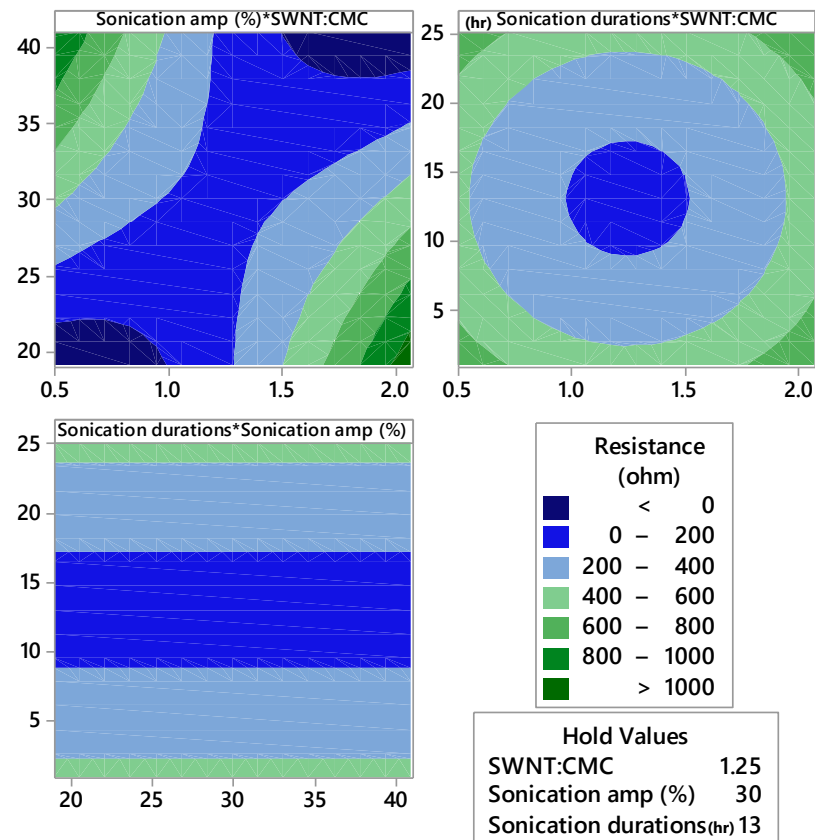
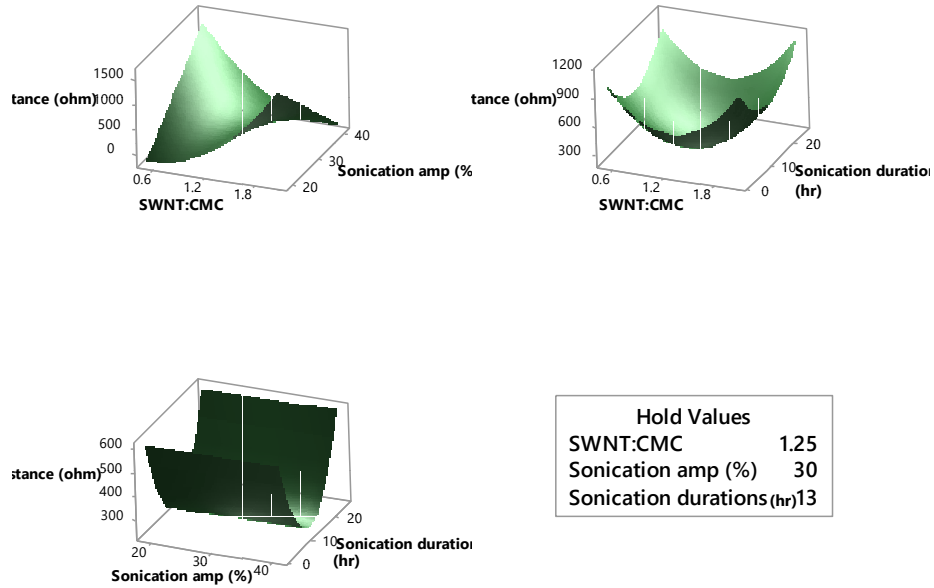


Figure 25. Contour plots and three-dimensional response surface of the resistance response for CMC at 100% (no dilution) corresponding to equation 5.

## Three-dimensional response surface plots



## Contour plots of the resistance response

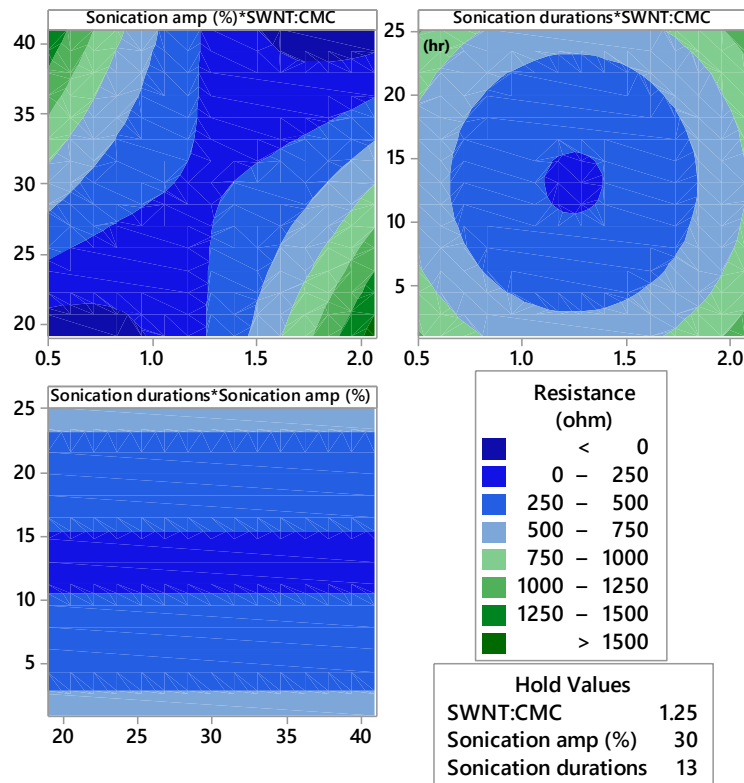
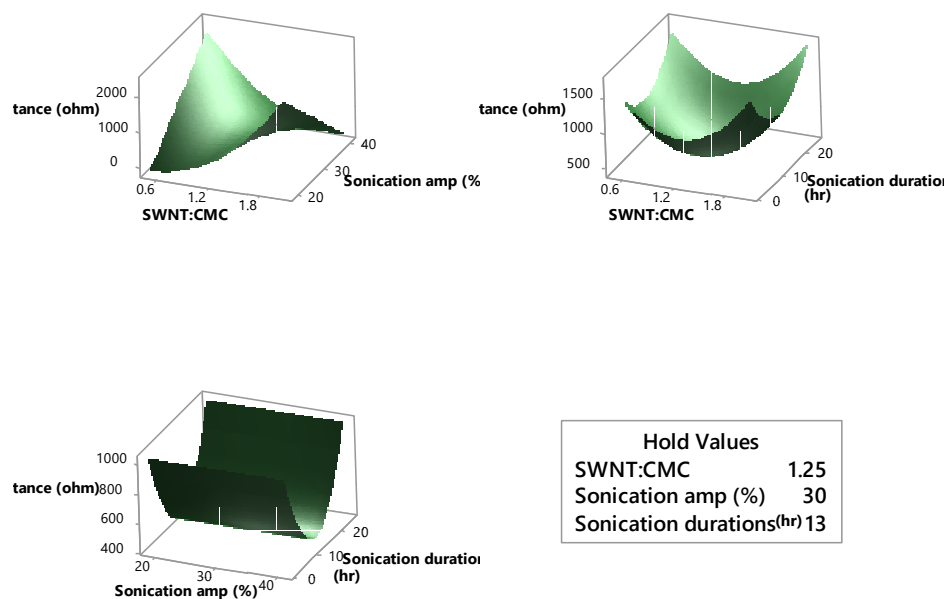


Figure 26. Contour plots and three-dimensional response surface of the resistance response for CMC diluted to 70%, corresponding to equation 6.

## Three dimensional response surface plots



## Contour plots of the resistance response

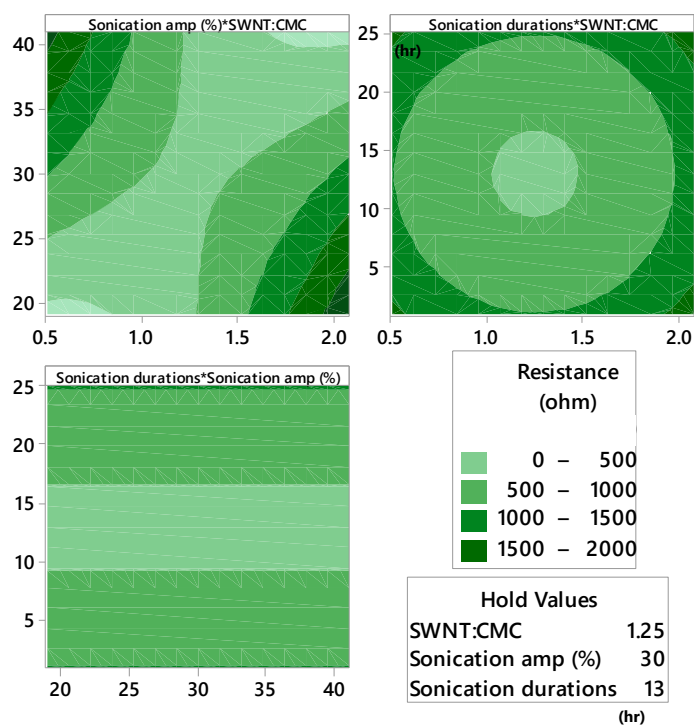
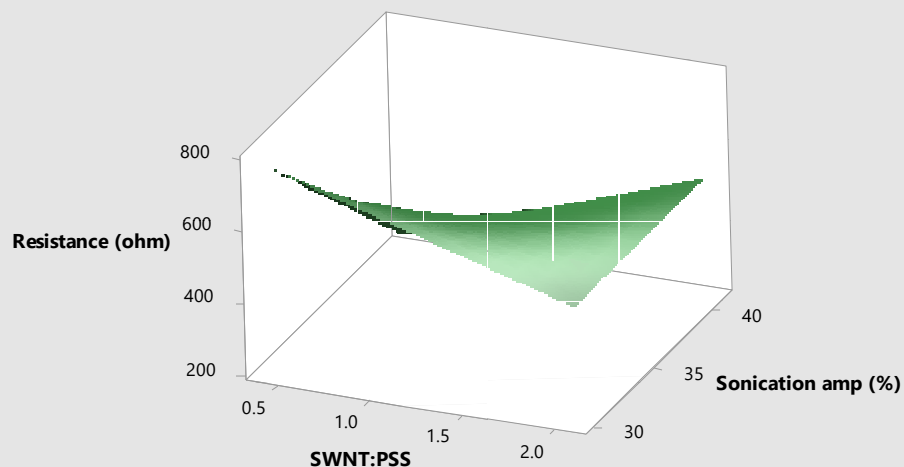


Figure 27. Contour plots and three-dimensional response surface of the resistance response for CMC diluted to 40%, corresponding to equation 7.

Surface Plot of Resistance (ohm) vs Sonication amp (%), SWNT:PSS



Contour Plot of Resistance (ohm) vs Sonication amp (%), SWNT:PSS

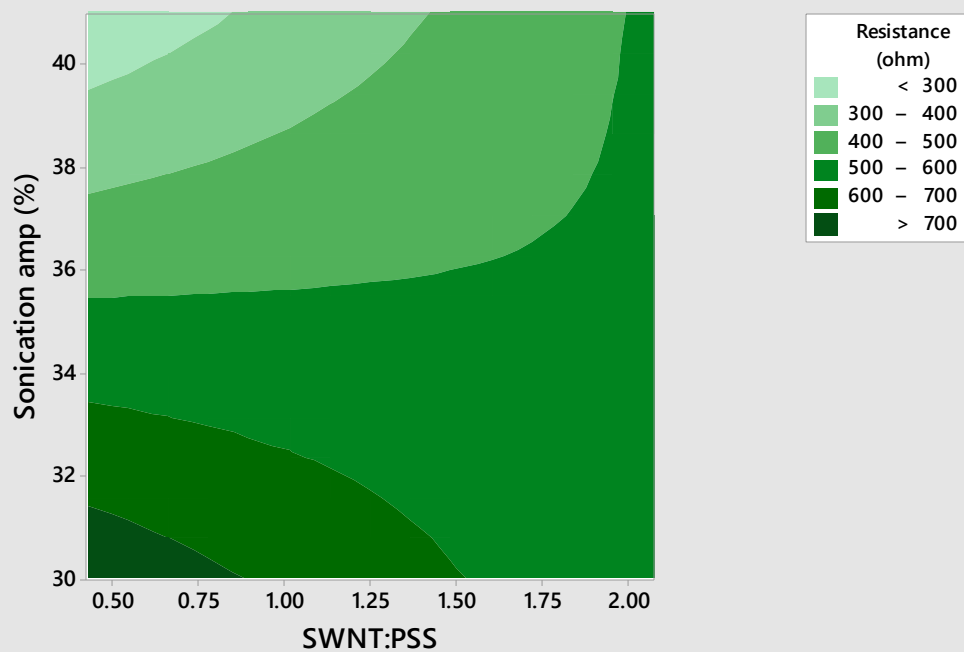
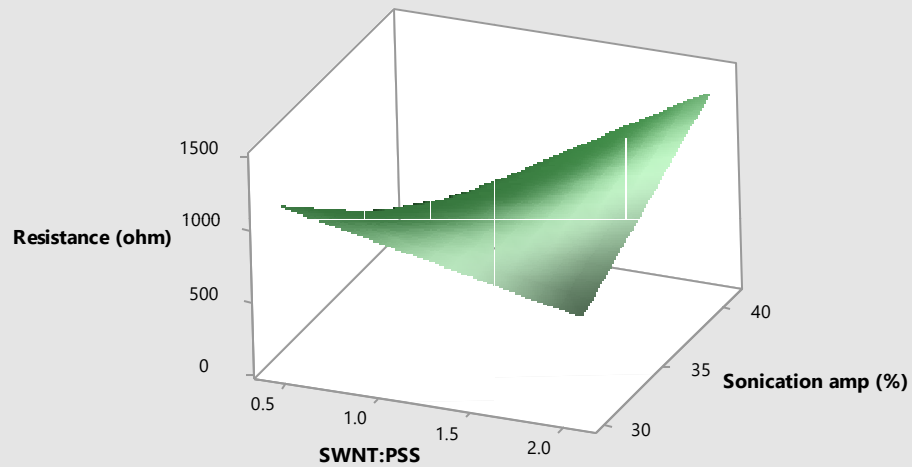


Figure 28. Contour plots and three-dimensional response surface of the resistance response for PSS at 100% (no dilution), corresponding to equation 8.



Surface Plot of Resistance (ohm) vs Sonication amp (%), SWNT:PSS



Contour Plot of Resistance (ohm) vs Sonication amp (%), SWNT:PSS

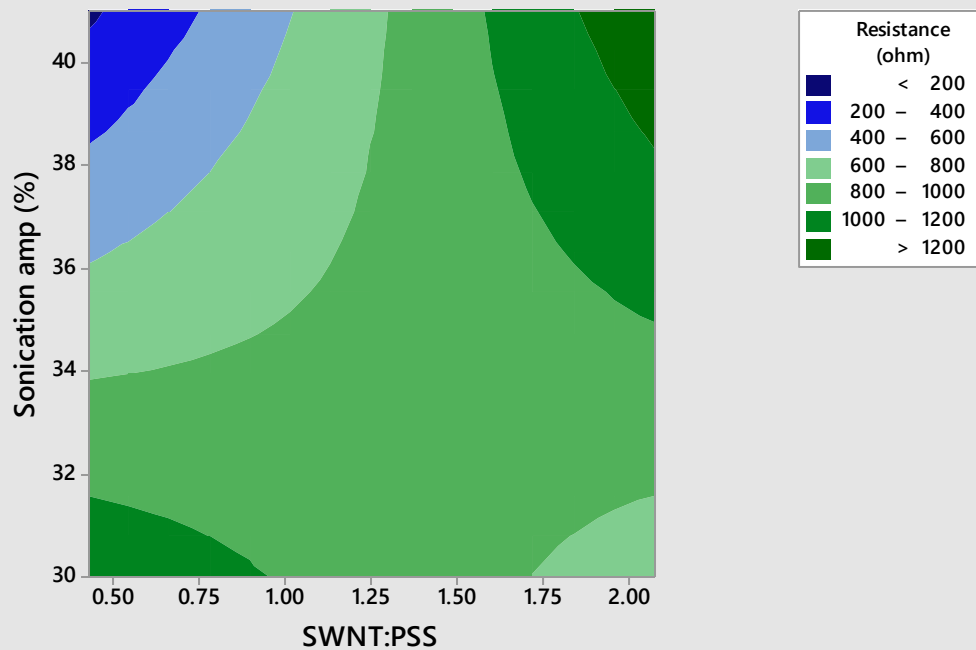
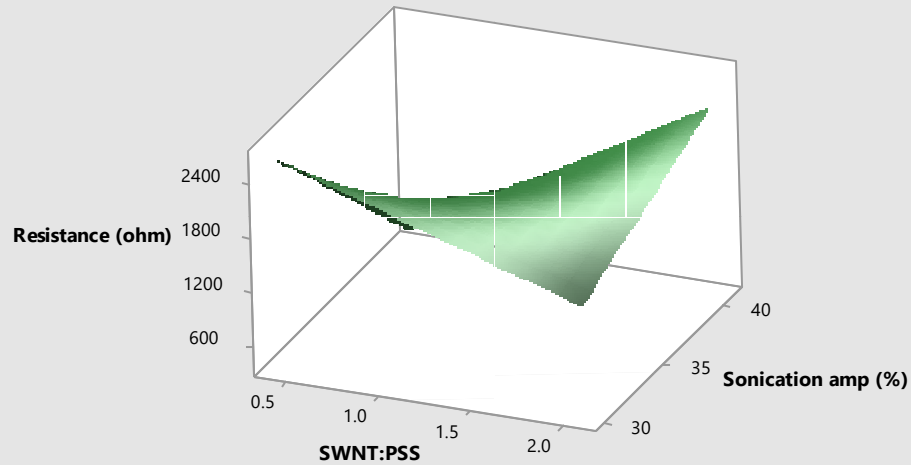


Figure 29. Contour plots and three-dimensional response surface of the resistance response for PSS diluted to 70%, corresponding to equation 9

Surface Plot of Resistance (ohm) vs Sonication amp (%), SWNT:PSS



Contour Plot of Resistance (ohm) vs Sonication amp (%), SWNT:PSS

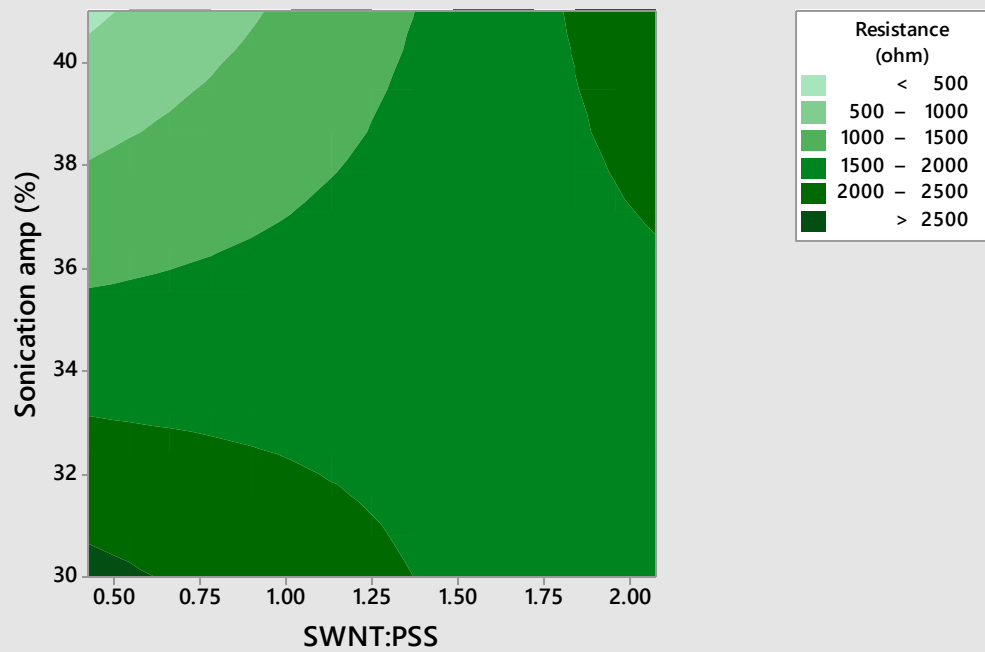


Figure 30. Contour plots and three-dimensional response surface of the resistance response for PSS diluted to 40%, corresponding to equation 10

### **2.3.3 Particle size analysis**

Table 18 and Table 19 show the summary of the particle size analysis test results sorted from small to large for each of the matrix formulations (dilution percentages (100% (no dilution), 70%, and 40%), SWNT:CMC/PSS ratio (2 to 0.5), Sonication amplitude (40% to 20%), sonication duration (24 to 2 hr)), and color coded by particle size (green (low) to red (high)).

Table 18- Summary of particle size analysis for CMC matrix, color coded by particle size (green (low) to red (high)).

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:CMC)	<i>X<sub>2</sub></i> (Sonication amp (%))	<i>X<sub>3</sub></i> (Sonication durations (h))	<i>diluted to</i>	<i>particle size (nm)</i>	<i>Resistance (ohm)</i>	<i>Predicted Resistance (ohm)</i>	<i>Calculated coatings thickness (μm)</i>	<i>Measured coatings thickness (μm)</i>	<i>Coating resistivity (ohm.cm)</i>
5	1.25	30	13	100%	66	130	164	4.04	-	0.0131
1	1.25	30	13	100%	110	134	164	4.23	-	0.0141
2	1.25	30	13	100%	110	149	164	4.13	-	0.0153
6	0.5	20	24	100%	112	24231	157*	4.27	-	2.580
15	2	40	24	100%	115	128	157	3.84	3.86	0.0123
20	1.25	41	13	100%	180	225	164	4.15	-	0.0234
8	2	20	24	100%	193	58878	1211*	2.53	-	3.730
18	0.425	30	13	100%	213	6256	493*	4.78	-	0.7470
13	1.25	30	13	100%	235	166	164	4.26	-	0.0177
17	1.25	30	13	100%	247	147	164	4.19	-	0.0154
7	1.25	30	25.1	100%	270	199	465	4.26	-	0.0212
10	1.25	19	13	100%	313	149	164	3.95	-	0.0147
16	1.25	30	13	100%	348	165	164	4.13	-	0.0170
19	1.25	30	0.9	100%	409	794	465	0.85	-	0.0169
12	2	40	2	100%	554	148	157	3.56	-	0.0131
11	2	20	2	100%	675	10914	1211*	2.55	-	0.697
9	0.5	40	24	100%	932	1317	1212	4.81	-	0.1580
4	2.075	30	13	100%	2230	555	492	2.26	-	0.0313
14	0.5	40	2	100%	2731	1069	1212	4.60	-	0.1230
3	0.5	20	2	100%	4131	3868395	157*	3.70	3.81	357.00
5	1.25	30	13	70%	66	174	235	2.83	-	0.0123
1	1.25	30	13	70%	110	190	235	2.96	-	0.014

Table 18 - continued

2	1.25	30	13	70%	110	214	235	2.89	-	0.0155
6	0.5	20	24	70%	112	28272	238*	2.99	-	2.110
15	2	40	24	70%	115	200	238	2.69	2.81	0.0134
20	1.25	41	13	70%	180	331	235	2.91	-	0.0241
8	2	20	24	70%	193	71517	1690*	1.77	-	3.170
18	0.425	30	13	70%	213	8141	745*	3.34	-	0.6810
13	1.25	30	13	70%	235	244	235	2.98	-	0.0182
17	1.25	30	13	70%	247	195	235	2.93	-	0.0143
7	1.25	30	25.1	70%	270	254	608	2.98	-	0.0189
10	1.25	19	13	70%	313	210	235	2.76	-	0.0145
16	1.25	30	13	70%	348	223	235	2.89	-	0.0161
19	1.25	30	0.9	70%	409	1085	608	0.6	-	0.0161
12	2	40	2	70%	554	202	238	2.49	-	0.0126
11	2	20	2	70%	675	15109	1690*	1.79	-	0.675
9	0.5	40	24	70%	932	1812	1692	3.36	-	0.1520
4	2.075	30	13	70%	2230	866	744	1.58	-	0.0342
14	0.5	40	2	70%	2731	1498	1692	3.22	-	0.1210
3	0.5	20	2	70%	4131	2812535	238*	2.59	2.42	285.00
5	1.25	30	13	40%	66	333	448	1.62	-	0.0135
1	1.25	30	13	40%	110	362	448	1.69	-	0.0153
2	1.25	30	13	40%	110	410	448	1.65	-	0.0169
6	0.5	20	24	40%	112	36426	469*	1.71	-	1.550
15	2	40	24	40%	115	401	468	1.54	1.63	0.0154
20	1.25	41	13	40%	180	595	448	1.66	-	0.0247
8	2	20	24	40%	193	122910	2547*	1.01	-	3.110
18	0.425	30	13	40%	213	10685	1152*	1.91	-	0.5110
13	1.25	30	13	40%	235	481	448	1.7	-	0.0205

Table 18 - continued

<i>17</i>	1.25	30	13	40%	247	423	448	1.68	-	0.0177
<i>7</i>	1.25	30	25.1	40%	270	463	1028	1.7	-	0.0197
<i>10</i>	1.25	19	13	40%	313	342	448	1.58	-	0.0135
<i>16</i>	1.25	30	13	40%	348	423	448	1.65	-	0.0175
<i>19</i>	1.25	30	0.9	40%	409	1862	1028	0.34	-	0.0158
<i>12</i>	2	40	2	40%	554	372	468	1.42	-	0.0132
<i>11</i>	2	20	2	40%	675	20437	2547*	1.02	-	0.522
<i>9</i>	0.5	40	24	40%	932	2838	2548	1.92	-	0.1360
<i>4</i>	2.075	30	13	40%	2230	1420	1150	0.9	-	0.032
<i>14</i>	0.5	40	2	40%	2731	2095	2548	1.84	-	0.0963
<i>3</i>	0.5	20	2	40%	4131	4405986	469	1.48	1.18	104.00

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

Dynamic light scattering system, Nicomp, was used to measure the particle size of the prepared formulations after the filtration. Figure 36 shows the particle size distribution of the matrix solution obtained for the Run 3 formulation using CMC dispersant. The largest particle size for the set of experiments in which CMC was used was observed in Run 3 (see Table 18) in which the maximum resistance was obtained. About 81% of the particles have an average particle size of ~5000 nm which is indicative of SWNT particle agglomeration.

Run 5 and 15 which resulted in matrix solutions with the minimum resistance (130  $\Omega$  and 128  $\Omega$ , respectively) possessed the minimum particle size (Figure 31 and Figure 32). The particle size distribution of the matrix solution obtained using Run 15 shows a monodisperse particle size distribution of about 115 nm (Figure 32). The matrix solution obtained using Run 5 has a bimodal particle size distribution with a small fraction of the particles (~15 %) having average particle size of ~210 nm and the larger fraction of the particles having average particle size of ~40 nm. These particles sizes are much greater than 20 times smaller than in Run 3 that demonstrated the highest resistance. In general, it can be seen from Table 18 that smaller particle sizes result in lower resistance. However, there are three runs with small particle size and very high resistance, runs 6, 8 and 18 (Table 18). These runs have the lowest sonication amplitude or the lowest SWNT:CMC ratio of the group thus suggesting potentially other factors other than particle size may be influencing resistivity. Regardless, the data in Table 18 suggests that agglomeration has an adverse effect on the conductivity of the nanotubes.

The topography of the deposited matrix solutions in the channels was investigated using an optical profiler.

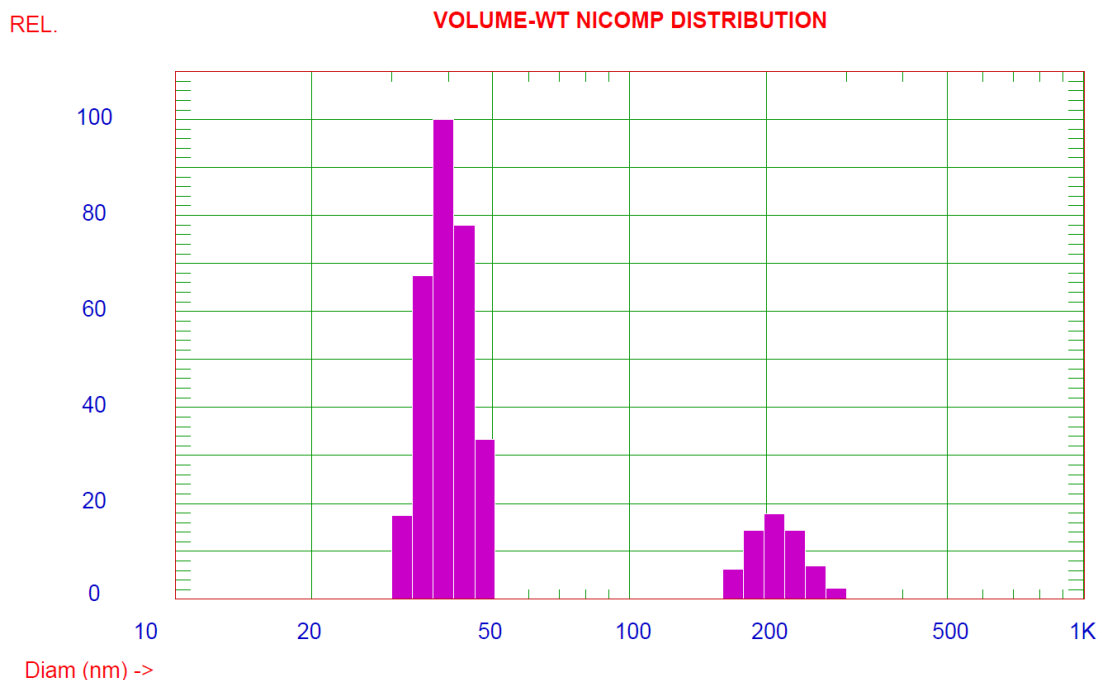
The surface profile of Run 15 of the experiments in which CMC was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution is presented in Figure 33 through Figure 35. In this run the minimum resistance for all formulations of  $128\ \Omega$  (see Table 4) was achieved with a monodisperse particle size distribution and small average particle size of 115 nm (see Figure 32). As can be observed in Figure 33, a uniform coating with an average roughness of 67 nm was obtained for the matrix coating at 100% (no dilution). It can also be observed that the smoothness of the coating increased with dilution. The same formulation that was diluted down to 70% showed a roughness of 50 nm (Figure 34), and the coating that was diluted down to 40% showed a roughness of 34 nm (Figure 35). Note that as the thickness of the layer is reduced (with dilution) that some larger particles become more exposed and protrude from the smoother surrounding layer.

On the other hand, Run 3 (see Table 4), which was characterized by having the largest particle size (4131 nm) (Figure 36) and the maximum resistance  $3.9\ M\Omega$  (see Table 6) among the set of experiments in which CMC was used, exhibits a rough surface. Figure 37 shows the surface morphology of the film obtained for Run 3 at 100% (no dilution) has an average roughness of 274 nm. The same formulation that was diluted down to 70% showed a slightly higher average roughness of 307 nm (one would expected it to be slightly lower) (Figure 38), and the coating that was diluted down to 40% showed an average roughness of 196 nm (Figure 39). This roughness is about 5 times larger than what run 15 (low resistance) showed.

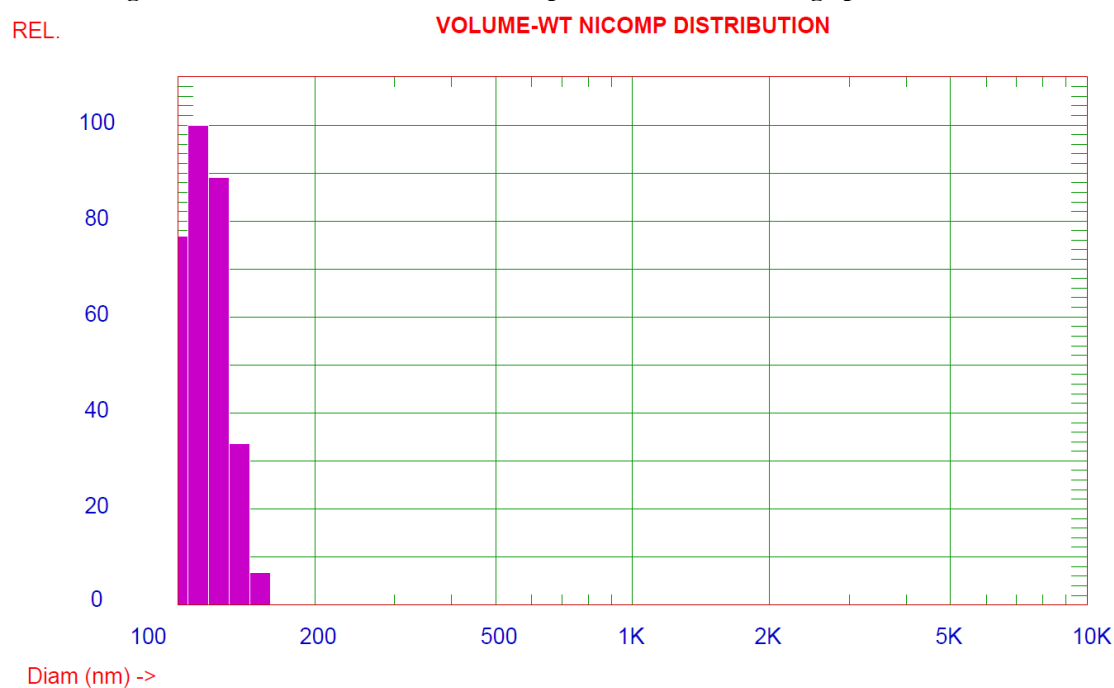
Thus for matrix formulations with CMC, the surface topography of the dried matrix films shows some correlation with the resistance response of the matrix. Uniform and smooth surfaces can be translated to low SWNT agglomeration and low resistance of the



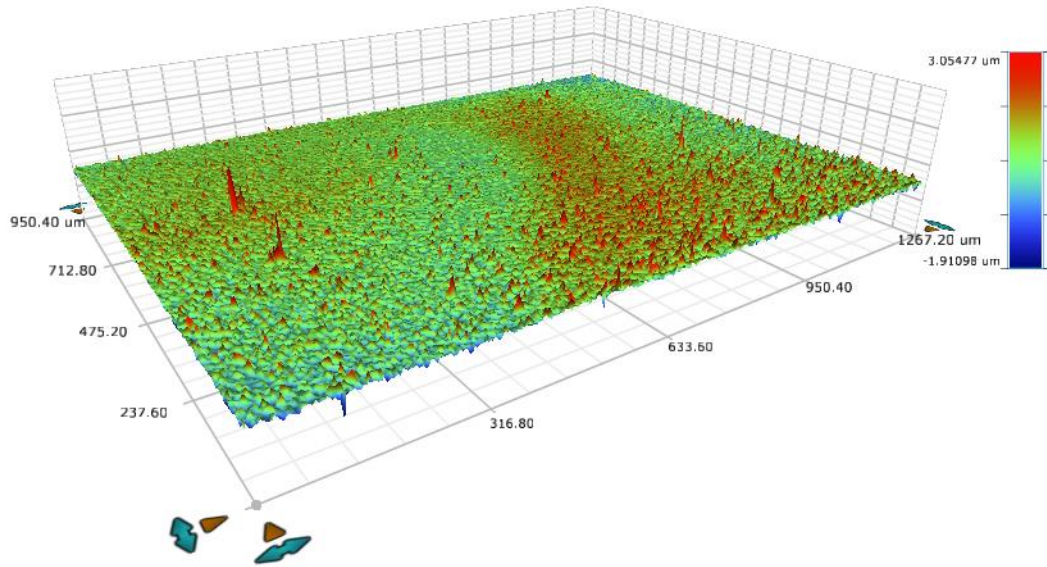
dried matrix solutions. Therefore it is desirable to obtain a uniform and smooth coating to lower resistivity of the matrix film.



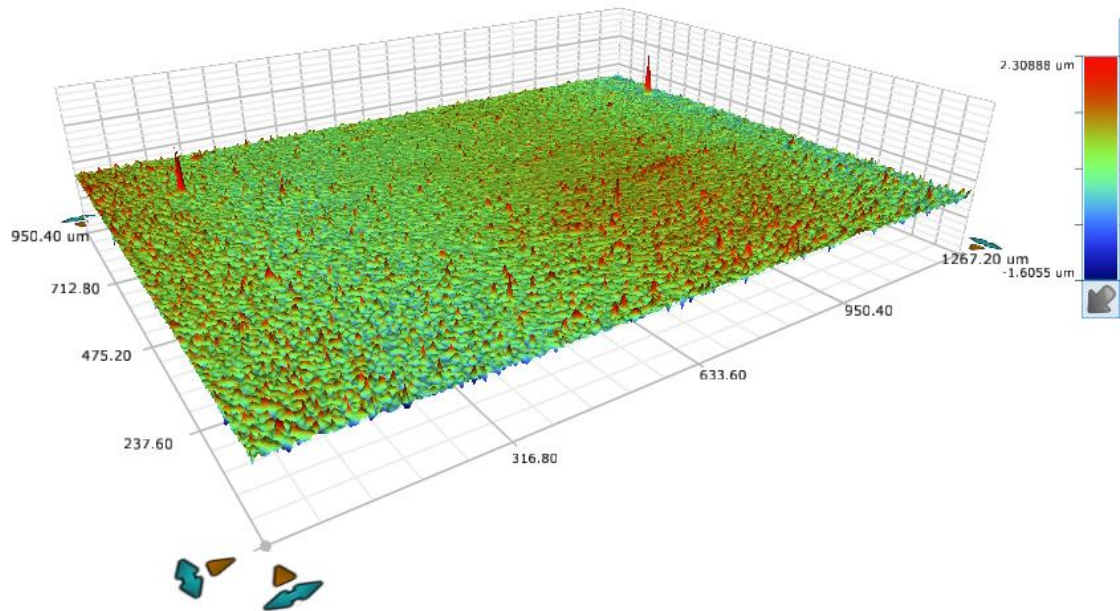
**Figure 31. Particle size distribution of the matrix solution obtained through Run 5 (SWNT:CMC=1.25, sonication amplitude=30%, sonication duration=13 hr) using CMC dispersant, showing a low resistance. About 85% of the particles have an average particle size of ~40 nm.**



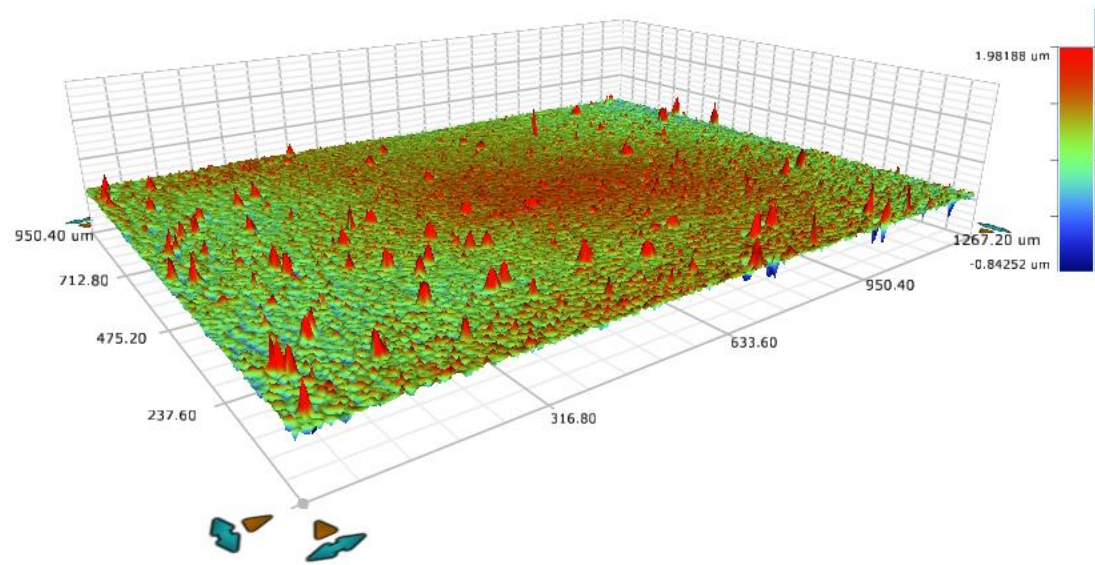
**Figure 32. Particle size distribution of the matrix solution obtained through Run 15 (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr) using CMC dispersant, showing a low resistance. A monodisperse average particle size of ~115 nm was obtained for this matrix solution.**



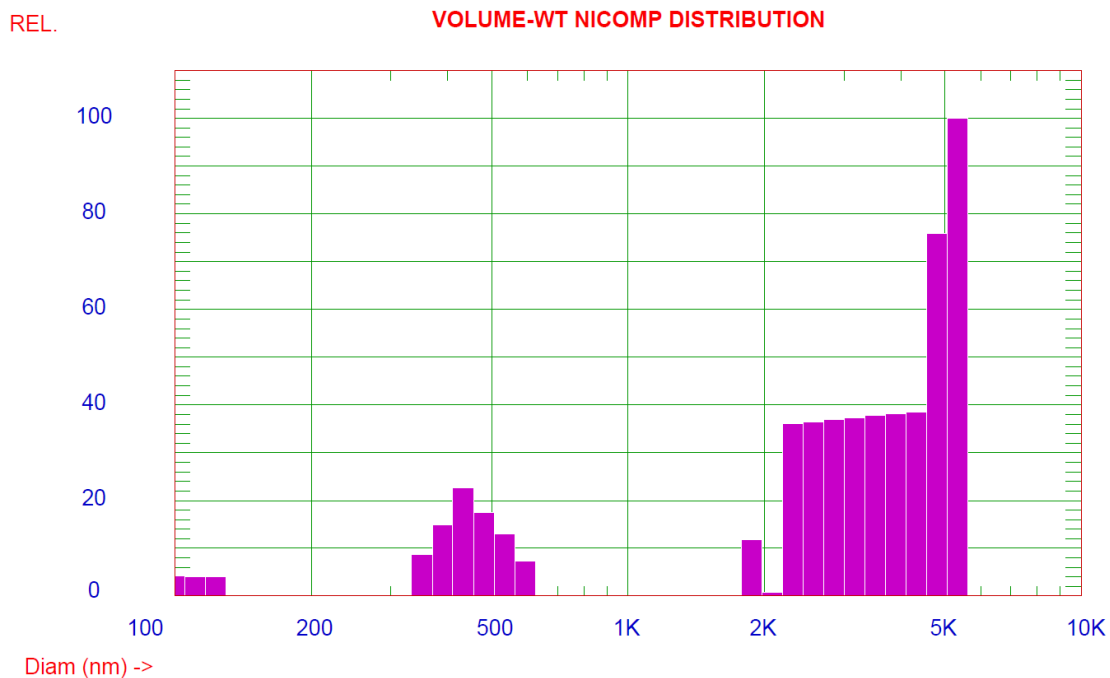
**Figure 33. Profilometry of Run 15 (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr), showing a uniform CMC-dispersed coating at 100% (no dilution) with an average roughness of 0.067  $\mu\text{m}$ .**



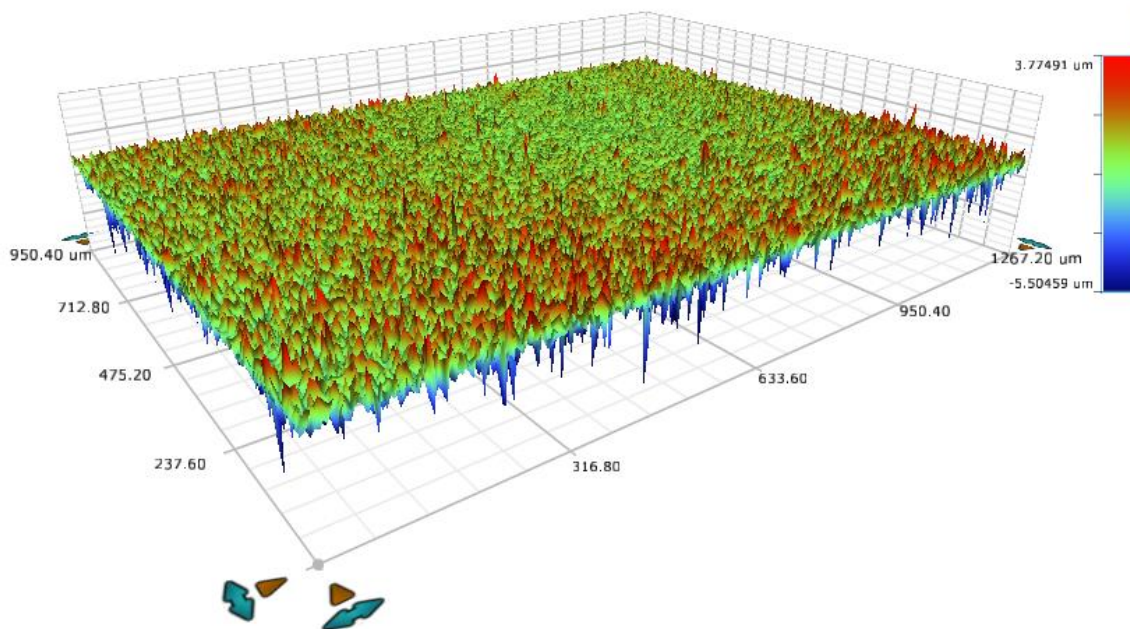
**Figure 34. Profilometry of Run 15 (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr), showing a uniform CMC-dispersed coating diluted to 70% with an average roughness of 0.05  $\mu\text{m}$ .**



**Figure 35. Profilometry of Run 15 (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr), showing a uniform CMC-dispersed coating diluted to 40% with an average roughness of 0.034 μm.**

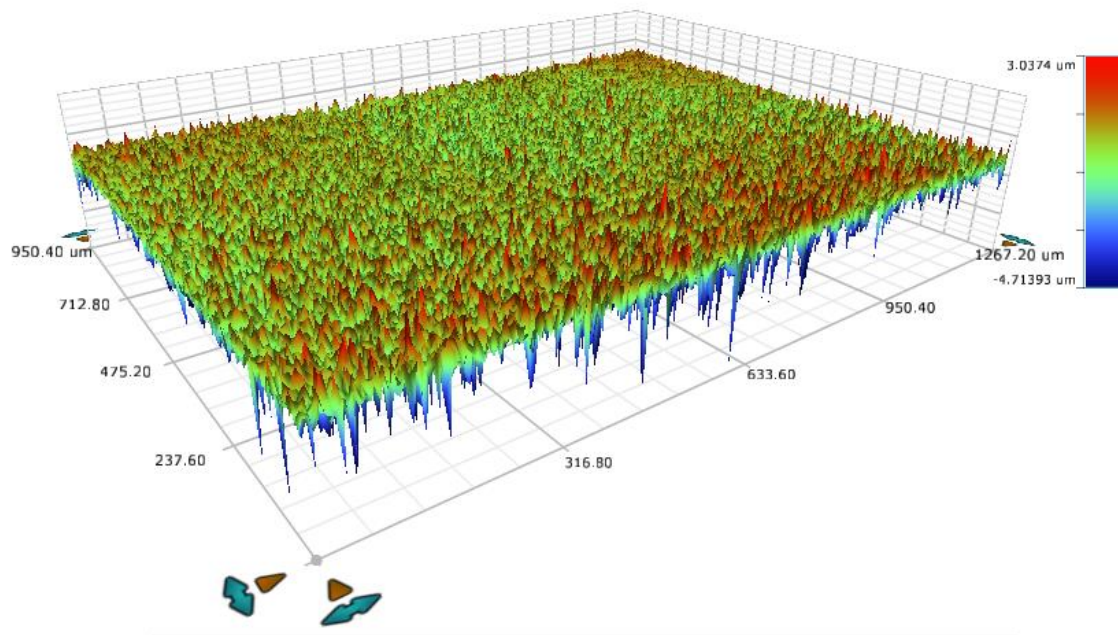


**Figure 36. Particle size distribution of the matrix solution obtained through Run 3 (SWNT:CMC=0.5, sonication amplitude=20%, sonication duration=2 hr) using CMC dispersant, showing agglomeration. About 81% of the particles have an average particle size of ~5000 nm.**

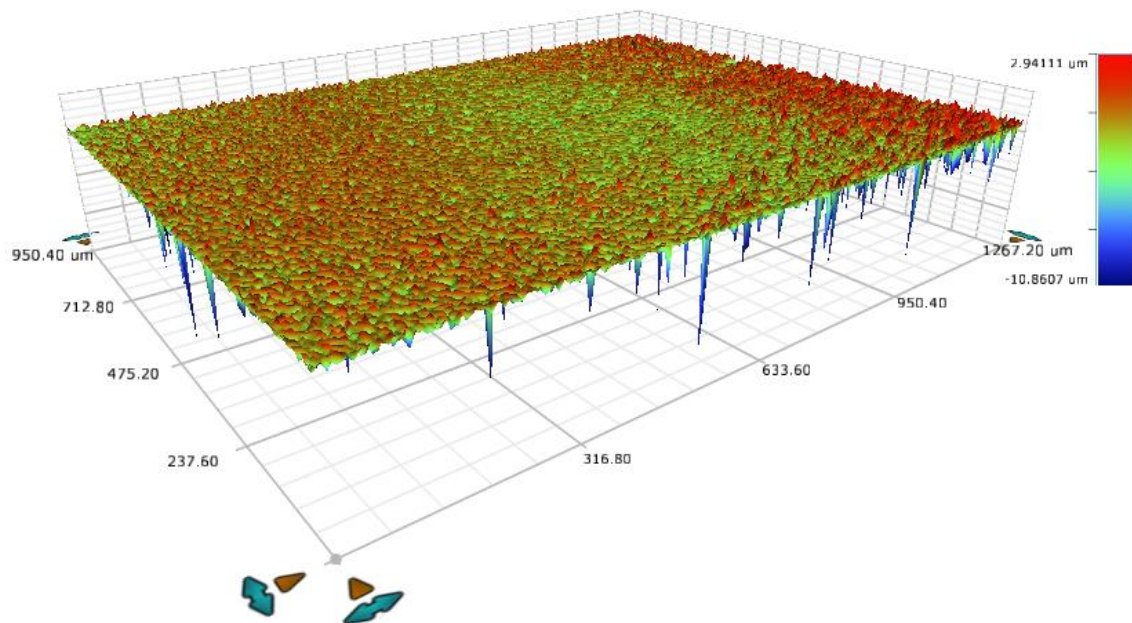


**Figure 37. Profilometry of Run 3 (SWNT:CMC=0.5, sonication amplitude=20%, sonication duration=2 hr), showing a uniform CMC-dispersed coating at 100% (no dilution) with an average roughness of 0.274  $\mu\text{m}$ .**





**Figure 38. Profilometry of Run 3 (SWNT:CMC=0.5, sonication amplitude=20%, sonication duration=2 hr), showing a uniform CMC-dispersed coating diluted to 70% with an average roughness of 0.307  $\mu\text{m}$ .**



**Figure 39. Profilometry of Run 3 (SWNT:CMC=0.5, sonication amplitude=20%, sonication duration=2 hr), showing a uniform CMC-dispersed coating diluted to 40% with an average roughness of 0.196  $\mu\text{m}$ .**

The behavior of the matrix solution in which PSS was used as the dispersant was found to be less predictable when compared to CMC-dispersed matrix solutions. Table 19 shows the summary of the particle size analysis test results sorted by particle size for all formulations within a given dilution percentage (100% (no dilution), 70%, and 40%), and color coded by particle size (green (low) to red (high)). The minimum particle size was observed for Run 4 with an average particle size of 15 nm. Run 5 had the next lowest average particle size, with average particle size of 18 nm (Figure 40) and the maximum particle size was observed in Run 15 with an average particle size of ~2880 nm (Figure 41).

**Table 19. Summary of particle size analysis for PSS matrix, color coded by particle size (green (low) to red (high)).**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:PSS)	<i>X<sub>2</sub></i> (Sonication amp (%))	<i>X<sub>3</sub></i> (Sonication durations (h))	<i>diluted to</i>	<i>particle size (nm)</i>	<i>Resistance (ohm)</i>	<i>Predicted Resistance (ohm)</i>	<i>Calculated coatings thickness (μm)</i>	<i>Measured coatings thickness (μm)</i>	<i>Coating resistivity (ohm.cm)</i>
4	2.075	30	13	100%	15	304	515	3.82	-	0.0291
5	1.25	30	13	100%	18	393	643	4.29	-	0.0422
8	2	20	24	100%	53	701788	550*	3.44	-	60.400
2	1.25	30	13	100%	107	749	643	4.23	-	0.0793
20	1.25	41	13	100%	110	125	370	6.23	5.66	0.0194
9	0.5	40	24	100%	110	277	286	8.11	-	0.0561
18	0.425	30	13	100%	110	560	770	8.51	-	0.1190
16	1.25	30	13	100%	122	365	643	4.87	-	0.0444
17	1.25	30	13	100%	127	1557	643	4.00	-	0.1560
6	0.5	20	24	100%	133	16316438	1232*	6.57	6.15	2680.00
3	0.5	20	2	100%	152	6453796	1232*	6.27	-	1010.00
7	1.25	30	25.1	100%	155	163	643	5.04	-	0.0206
19	1.25	30	0.9	100%	161	17817	643*	0.66	-	0.2950
10	1.25	19	13	100%	169	2174028	916*	3.71	-	202.00
1	1.25	30	13	100%	206	459	643	4.78	-	0.0549
11	2	20	2	100%	211	332381	550*	3.57	-	29.700
14	0.5	40	2	100%	292	431	286	8.30	-	0.0894
12	2	40	2	100%	914	220	503	0.47	-	0.0026
13	1.25	30	13	100%	915	1211	643	4.61	-	0.1400
15	2	40	24	100%	2880	921	503	3.19	-	0.07
4	2.075	30	13	70%	15	390	707	2.67	-	0.0261
5	1.25	30	13	70%	18	544	922	3.00	-	0.0409



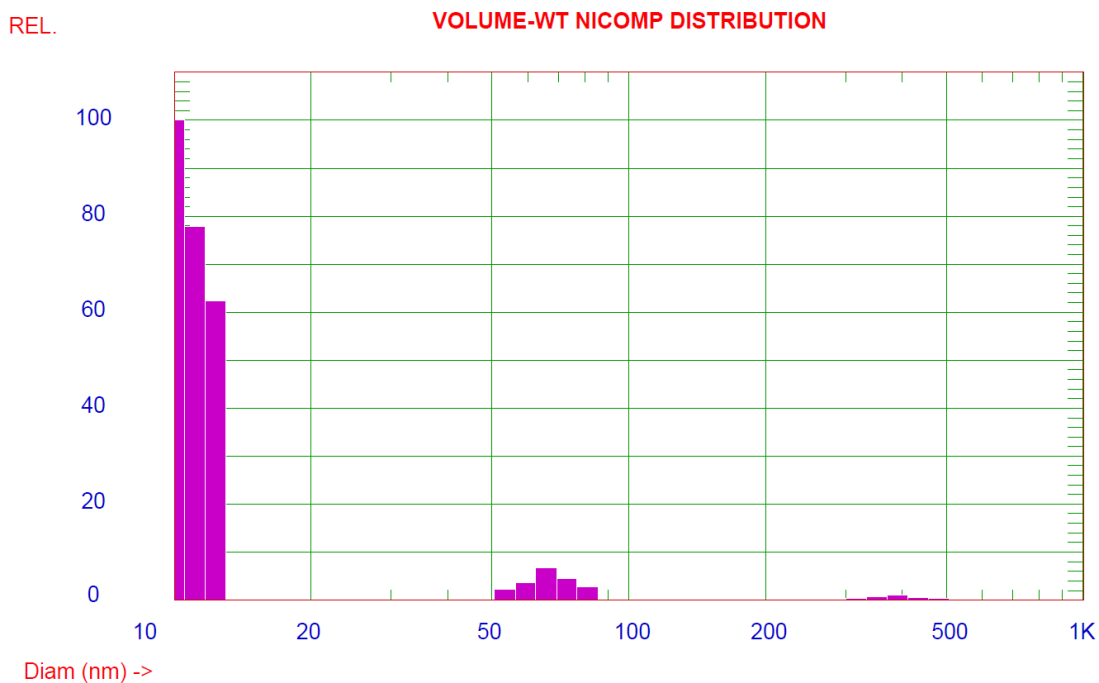
Table 19 - continued

8	2	20	24	70%	53	1141095	202*	2.41	-	68.800
2	1.25	30	13	70%	107	1299	922	2.96	-	0.0962
20	1.25	41	13	70%	110	183	762	4.36	4.23	0.0199
9	0.5	40	24	70%	110	366	301	5.68	-	0.0519
18	0.425	30	13	70%	110	819	1136	5.96	-	0.1220
16	1.25	30	13	70%	122	502	922	3.41	-	0.0428
17	1.25	30	13	70%	127	2094	922	2.80	-	0.1470
6	0.5	20	24	70%	133	16505225	1932*	4.60	4.29	1900.00
3	0.5	20	2	70%	152	6855038	1932*	4.39	-	753.00
7	1.25	30	25.1	70%	155	241	922	3.53	-	0.0212
19	1.25	30	0.9	70%	161	21716	922	0.46	-	0.2520
10	1.25	19	13	70%	169	4977047	1081*	2.60	-	323.00
1	1.25	30	13	70%	206	650	922	3.35	-	0.0544
11	2	20	2	70%	211	509977	202*	2.50	-	31.900
14	0.5	40	2	70%	292	555	301	5.81	-	0.0807
12	2	40	2	70%	914	1339	1252	0.33	-	0.0109
13	1.25	30	13	70%	915	1698	922	3.23	-	0.1370
15	2	40	24	70%	2880	1484	1252	2.23	-	0.08
4	2.075	30	13	40%	15	770	1532	1.53	-	0.0294
5	1.25	30	13	40%	18	1202	2078	1.72	-	0.0516
8	2	20	24	40%	53	7231889	1002*	1.38	-	249.000
2	1.25	30	13	40%	107	2176	2078	1.69	-	0.0921
20	1.25	41	13	40%	110	391	1358	2.49	2.54	0.0244
9	0.5	40	24	40%	110	801	685	3.24	-	0.0650
18	0.425	30	13	40%	110	1863	2625	3.41	-	0.1590
16	1.25	30	13	40%	122	1099	2078	1.95	-	0.0535
17	1.25	30	13	40%	127	5297	2078	1.60	-	0.2120

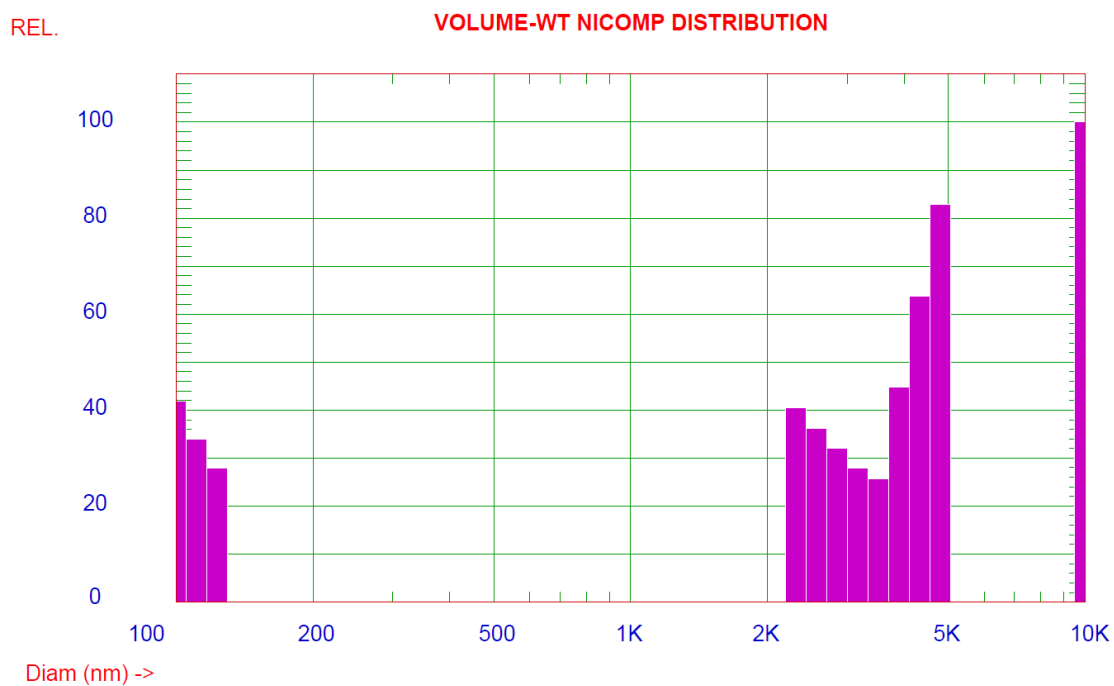
Table 19 - continued

<b>6</b>	0.5	20	24	40%	133	25362920	<i>4465*</i>	2.63	2.46	1670.00
<b>3</b>	0.5	20	2	40%	152	14250756	<i>4465*</i>	2.51	-	894.00
<b>7</b>	1.25	30	25.1	40%	155	575	2078	2.02	-	0.0290
<b>19</b>	1.25	30	0.9	40%	161	52246	<i>2078*</i>	0.27	-	0.3470
<b>10</b>	1.25	19	13	40%	169	12746253	<i>2799*</i>	1.48	-	473.00
<b>1</b>	1.25	30	13	40%	206	1464	2078	1.91	-	0.0700
<b>11</b>	2	20	2	40%	211	1099619	<i>1002*</i>	1.43	-	39.300
<b>14</b>	0.5	40	2	40%	292	1100	685	3.32	-	0.0913
<b>12</b>	2	40	2	40%	914	2529	2162	0.19	-	0.0118
<b>13</b>	1.25	30	13	40%	915	4164	2078	1.84	-	0.1920
<b>15</b>	2	40	24	40%	2880	2326	2162	1.28	-	0.07

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model



**Figure 40.** Particle size distribution of the matrix solution obtained through Run 5 (SWNT:PSS=1.25, sonication amplitude=30%, sonication duration=13 hr) using PSS dispersant, showing a low resistance of 393  $\Omega$ . The average particle size of ~18 nm was reported for this matrix solution.



**Figure 41.** Particle size distribution of the matrix solution obtained through Run 15 (SWNT:PSS=2, sonication amplitude=40%, sonication duration=24 hr) using PSS dispersant, showing a high resistance of 2326  $\Omega$ . The average particle size of ~2880 nm was reported for this matrix solution.

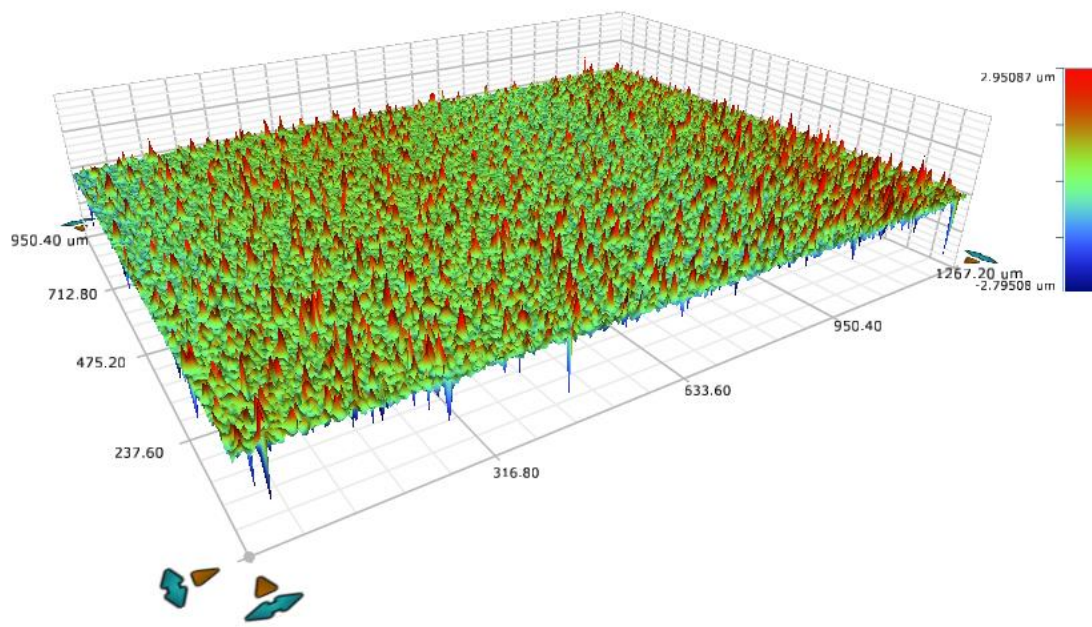
The sample with the minimum average resistance of 125  $\Omega$  in Run 20 was found to have a monodisperse particle size distribution with an average particle size of 110 nm (Figure 42). The samples with the maximum average resistance of 25 K $\Omega$  in Run 6 also had a monodisperse particle size distribution with an average particle size of ~133 nm (Figure 46). This fact indicates that PSS is very sensitive to the sonication conditions. This may result in a portion of PSS wrapping around itself, forming bundles rather than wrapping around SWNTs and dispersing them, or wrapping in multiple layers around SWNTs which will hinder electron transport of conductive carbon nanotubes [92, 93, 96, 98]. This may also create larger particles which results in the removal of SWNTs from the solution after filtrations. This fact can also be observed through the analysis of variance of the particle sizes obtained in the PSS-dispersed matrix solutions (Table 15 through Table 17) as the sonication amplitude and its interaction with the concentration of PSS are the only significant factors among all ( $\alpha=0.05$ ). In contrast, the sonication duration is the most significant factor in the analysis of variance of the particle sizes obtained in the CMC-dispersed matrix solutions (see Table 12 through Table 14). No interaction is observed for the CMC-dispersed matrix solutions. Therefore, the interaction that exists between the PSS and the sonication amplitude will make PSS act even more unpredictable at various situations. This makes CMC as a superior, more predictably behaving dispersant for this application. Considering the t-statistics for each model term and the provisions of the RSM, the following first-order model can be used to predict the particle size of the CMC-dispersed matrix solutions:

$$\text{Particle size (nm)} = 1490 - 60.1 X_3 \quad (11)$$

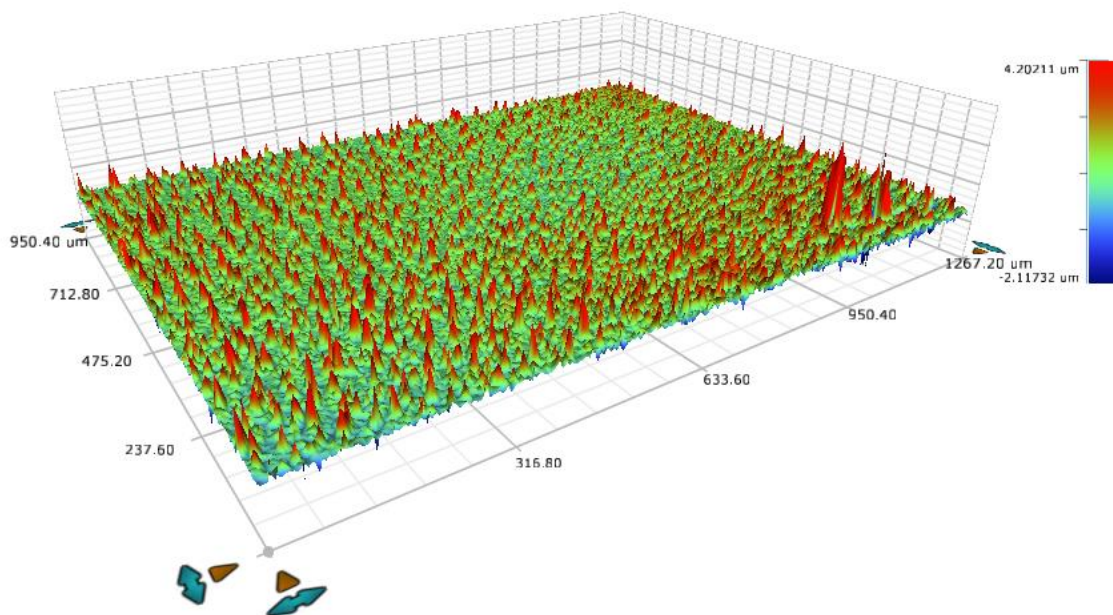
The surface profile of Run 20 of the experiments in which PSS was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution is presented in Figure 43 through Figure 45. As discussed earlier, in this run the minimum resistance of 125  $\Omega$  was achieved. As can be observed in Figure 43, a coating with an average roughness of 186 nm was obtained for the matrix coating at 100% (no dilution). This average roughness is almost three times more than its CMC counterpart (67 nm). The same formulation that was diluted down to 70% showed a roughness of 133 nm (Figure 44) (compare to 50 nm (Figure 34)), and the coating that was diluted down to 40% showed a roughness of 134 nm (Figure 45) (compare to 34 nm (Figure 35)).

On the other hand, Run 6, which was characterized by the maximum resistance among the set of experiments in which PSS was used, exhibit an even rougher surface. Figure 47 shows the surface morphology of the coating obtained from Run 6 at 100% (no dilution), revealing an average roughness of 601 nm (compare to the CMC counterpart at 274 nm). The coating non-uniformity of the PSS samples is obvious in the surface profiles of the all the PSS samples. The same formulation that was diluted down to 70% showed a roughness of 923 nm (Figure 48) (compare to 307 nm (Figure 38)), and the coating that was diluted down to 40% showed a roughness of 288 nm (Figure 49) (compared to 196 nm as shown in Figure 39).

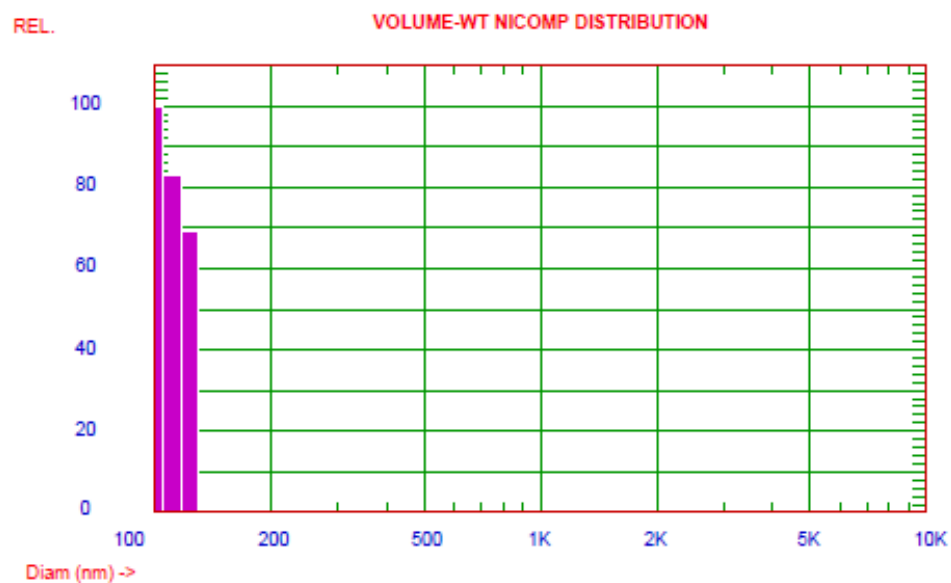




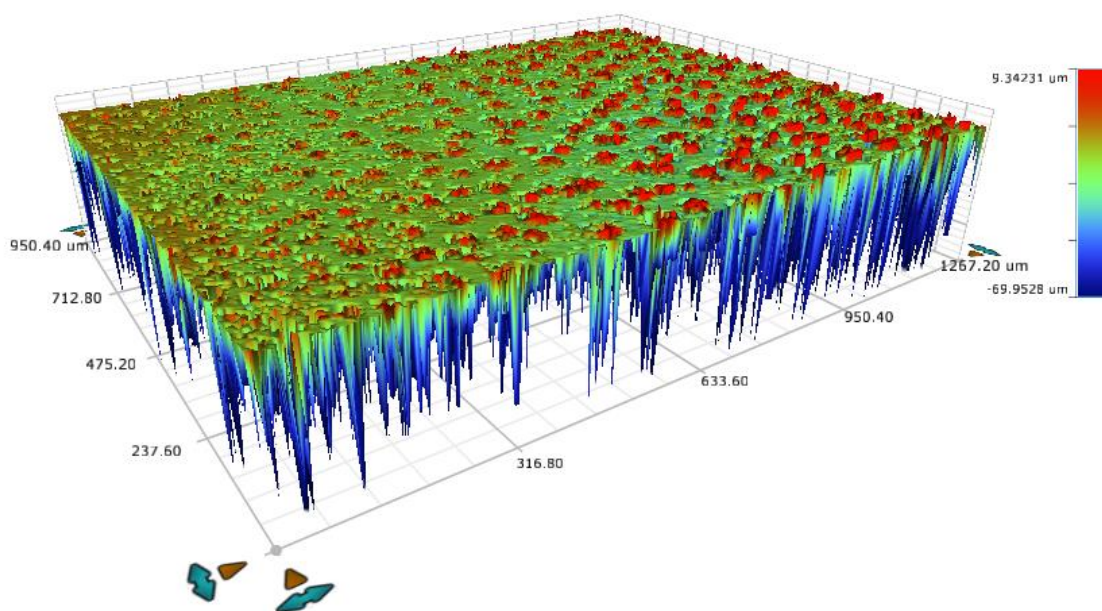
**Figure 44. Profilometry of Run 20 (SWNT:PSS=1.25, sonication amplitude=41%, sonication duration=13 hr), showing a PSS-dispersed coating diluted to 70% with an average roughness of 0.133  $\mu\text{m}$ .**



**Figure 45. Profilometry of Run 20 (SWNT:PSS=1.25, sonication amplitude=41%, sonication duration=13 hr), showing a PSS-dispersed coating diluted to 40% with an average roughness of 0.134  $\mu\text{m}$ .**

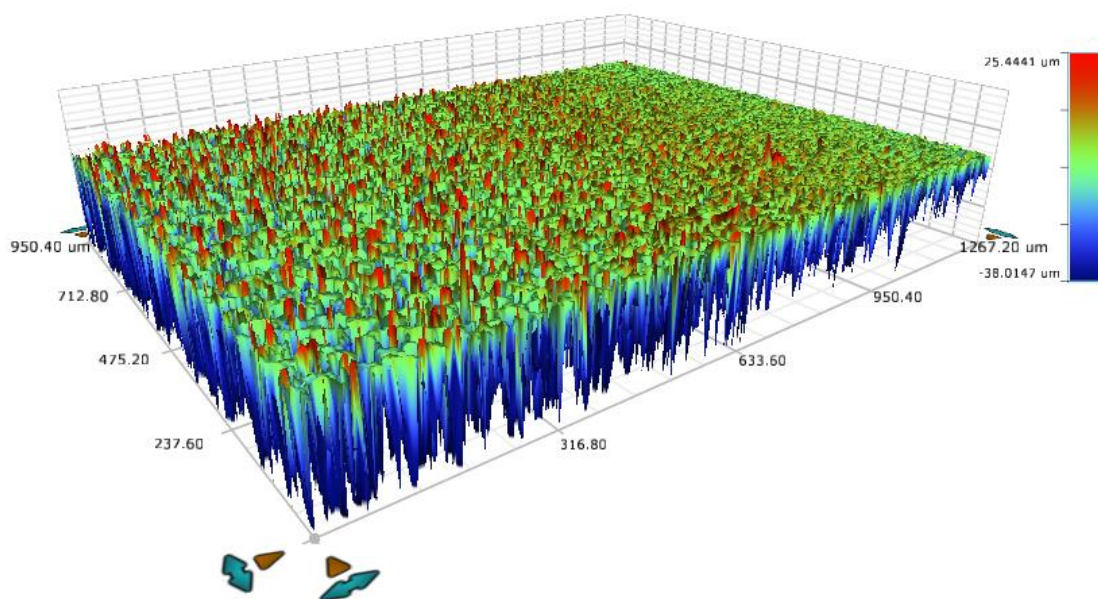


**Figure 46.** Particle size distribution of the matrix solution obtained through Run 6 (SWNT:PSS=0.5, sonication amplitude=20%, sonication duration=24 hr) using PSS dispersant, showing a high resistance of 25 K $\Omega$ . The average particle size of ~133 nm was reported for this matrix solution.

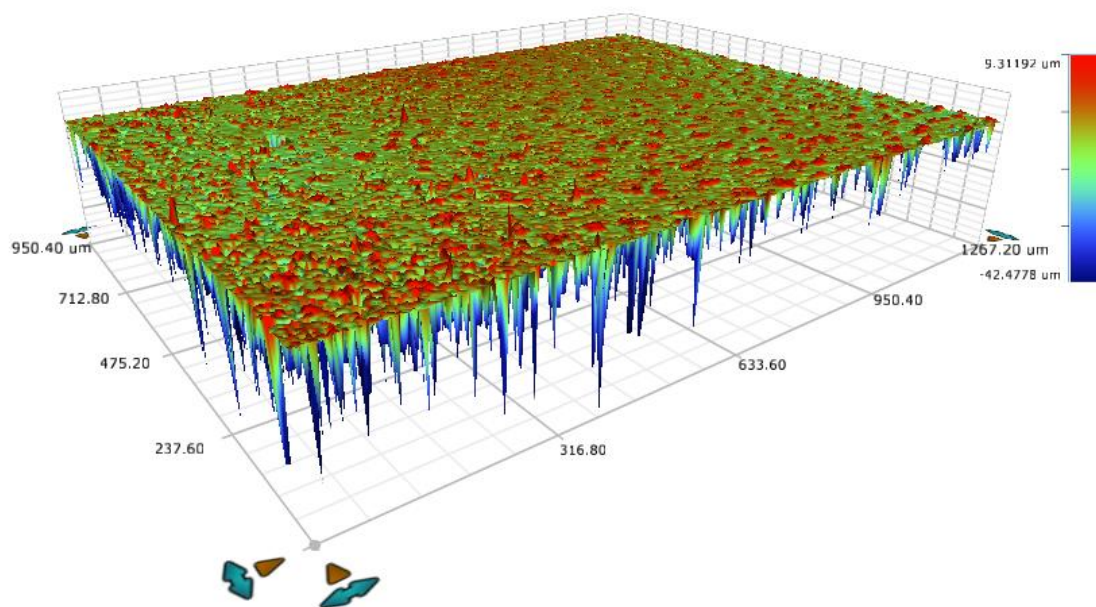


**Figure 47.** Profilometry of Run 6 (SWNT:PSS=0.5, sonication amplitude=20%, sonication duration=24 hr), showing a PSS-dispersed coating at 100% (no dilution) with an average roughness of 0.601  $\mu\text{m}$ .





**Figure 48. Profilometry of Run 6 (SWNT:PSS=0.5, sonication amplitude=20%, sonication duration=24 hr), showing a PSS-dispersed coating diluted to 70% with an average roughness of 0.923  $\mu\text{m}$ .**



**Figure 49. Profilometry of Run 6 (SWNT:PSS=0.5, sonication amplitude=20%, sonication duration=24 hr), showing a PSS-dispersed coating diluted to 40% with an average roughness of 0.288  $\mu\text{m}$ .**

Therefore, it can be concluded that a smoother surface can be achieved by using CMC as the dispersant for dispersing carbon nanotubes. Agglomerations and non-uniformity is obviously visible when SWNTs are dispersed using PSS at any concentration when compared to its CMC matrix solution counterparts. This makes CMC a superior, more predictable dispersant for SWNTs dispersion.

#### **2.3.4 Mass balance and thickness**

The thickness of the coatings is important because of their effect on the resistance of the obtained dried films. The higher the coating thickness is, the less the resistance of the coating will be. The thickness of the coatings can be experimentally and theoretically measured. The experimental measurements can be performed using optical profiler for each coating. Theoretical estimation of the coating is also possible through performing an accurate mass balance between the solid materials that were originally weighed for each formulation and the amount solids that were filtered out through filtration and centrifugation. For the purpose of theoretical thickness calculations, all the vials and filters used for each formulation were marked and weighed before the experiments. After the experiments, the left over matrix solution and their corresponding filters were dried in an oven for 2 days, under vacuum, until all the moisture was removed from the solution, leaving only solid particles in the vials. The vials were then weighed again and the amount of remaining solid that was eliminated from each solution was identified. This data was used to measure the theoretical thickness of the final coatings.

The density ( $\rho$ ) of the materials in the matrix solution has the following relationship with the mass ( $m$ ) and the volume of the solution:

$$\rho = \frac{m}{a \times b \times h} \quad (12)$$

Where (a) is the length, (b) is the width, and (h) is the thickness of the deposited matrix into each separate channels. This equation can be rearranged such that the thickness (h) represents the thickness of the deposited SWNT matrix solution after the solvent evaporation (dry thickness):

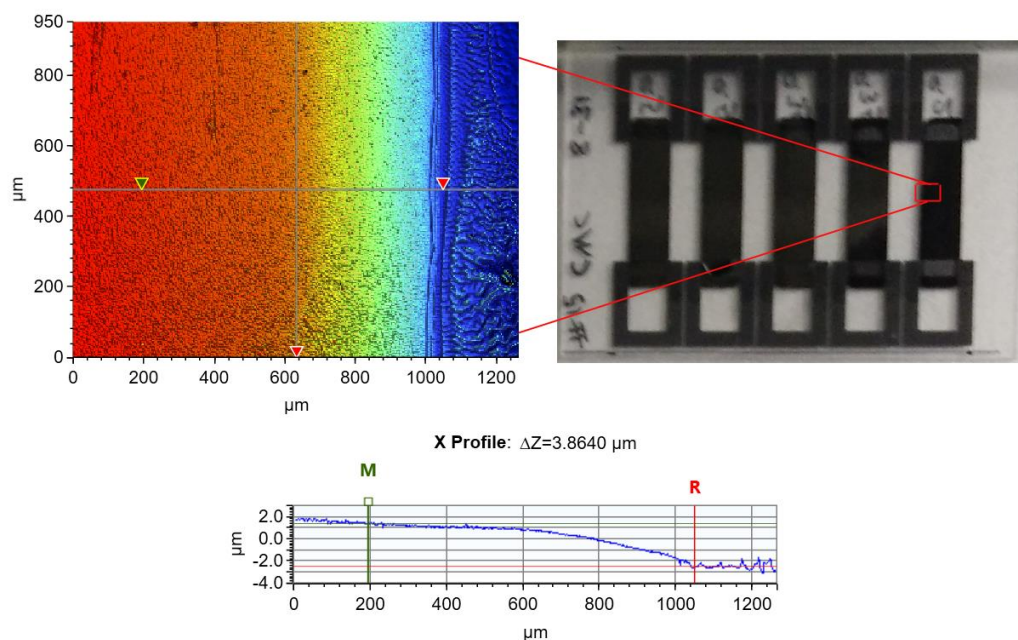
$$h = \frac{m}{a \times b \times \rho} \quad (13)$$

The theoretical thickness of the deposited matrix materials is presented in Table 20.

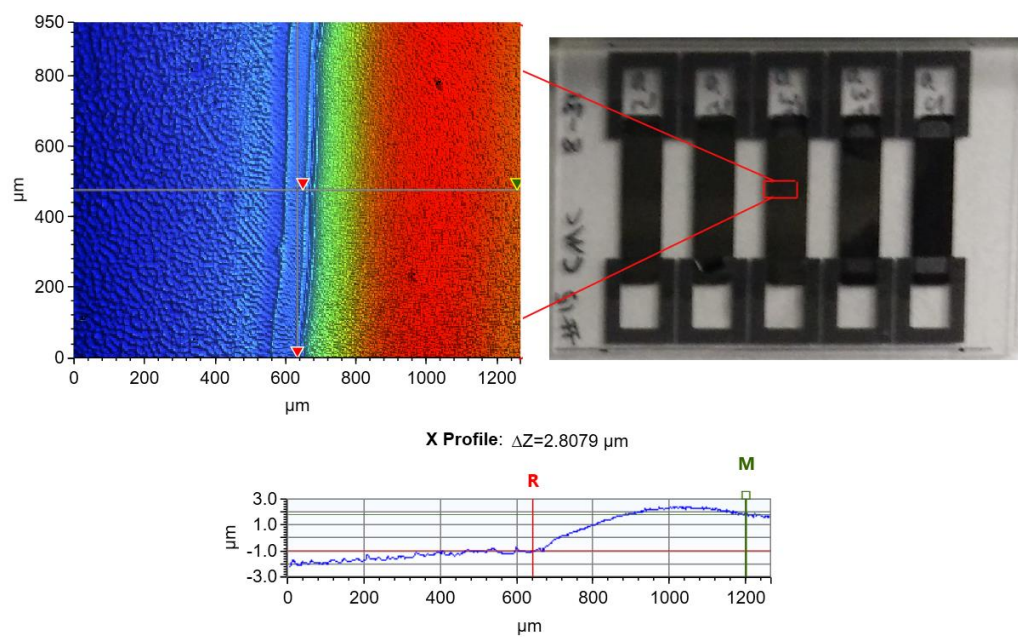
**Table 20- Theoretical calculation of the coating thicknesses for different dilutions using the mass balance and the densities of each solution. Measured thicknesses by 2D microscopy profiling are shown in parentheses.**

Sample	CMC coatings thickness (μm)			PSS coatings thickness (μm)		
	100%	70%	40%	100%	70%	40%
1	4.23	2.96	1.69	4.78	3.35	1.91
2	4.13	2.89	1.65	4.23	2.96	1.69
3	3.70 (3.81)	2.59 (2.42)	1.48(1.18)	6.27	4.39	2.51
4	2.26	1.58	0.90	3.82	2.67	1.53
5	4.04	2.83	1.62	4.29	3.00	1.72
6	4.27	2.99	1.71	6.57 (6.15)	4.60 (4.29)	2.63 (2.46)
7	4.26	2.98	1.70	5.04	3.53	2.02
8	2.53	1.77	1.01	3.44	2.41	1.38
9	4.81	3.36	1.92	8.11	5.68	3.24
10	3.95	2.76	1.58	3.71	2.60	1.48
11	2.55	1.79	1.02	3.57	2.50	1.43
12	3.56	2.49	1.42	0.47	0.33	0.19
13	4.26	2.98	1.70	4.61	3.23	1.84
14	4.60	3.22	1.84	8.30	5.81	3.32
15	3.84 (3.86)	2.69 (2.81)	1.54 (1.63)	3.19	2.23	1.28
16	4.13	2.89	1.65	4.87	3.41	1.95
17	4.19	2.93	1.68	4.00	2.80	1.60
18	4.78	3.34	1.91	8.51	5.96	3.41
19	0.85	0.60	0.34	0.66	0.46	0.27
20	4.15	2.91	1.66	6.23 (5.66)	4.36 (4.23)	2.49 (2.54)

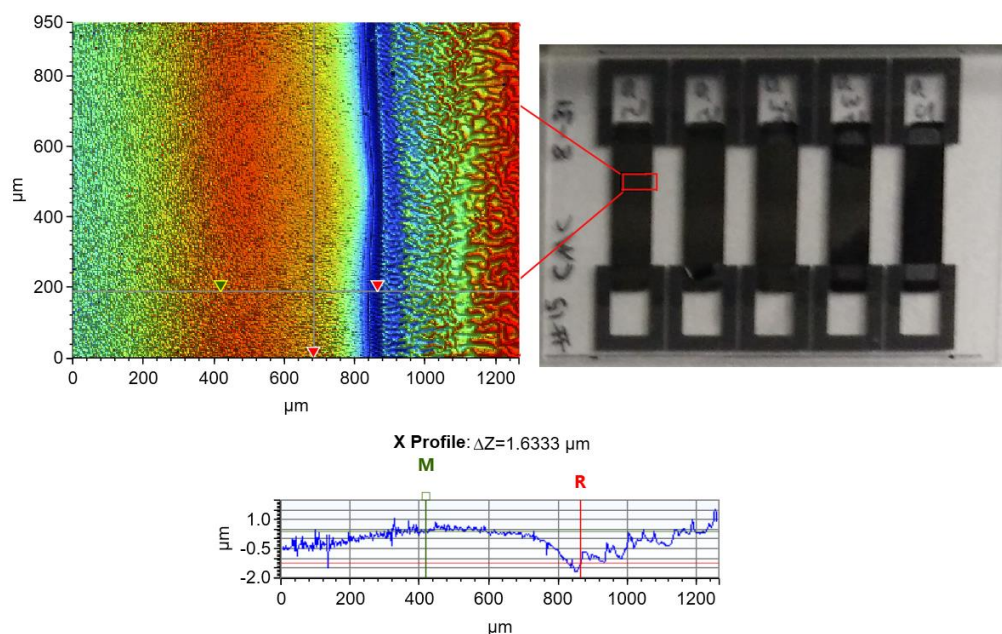
The thickness of the dried matrix coating was also investigated using the optical profiler. For this purpose, the optical profile of the edge of the coatings was obtained using optical profiler for each sample. The thickness of matrix coatings obtained from Run 15 of the formulation in which CMC was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution is presented in Figure 50 through Figure 52. The thickness of the deposited matrix solution is measured to be 3.86, 2.81, and 1.63  $\mu\text{m}$ , respectively. As expected, the thickness of the coatings decreased by the dilution of the matrix solution.



**Figure 50. The thickness of the CMC-dispersed coating obtained from run 15 at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 3.864  $\mu\text{m}$ .**



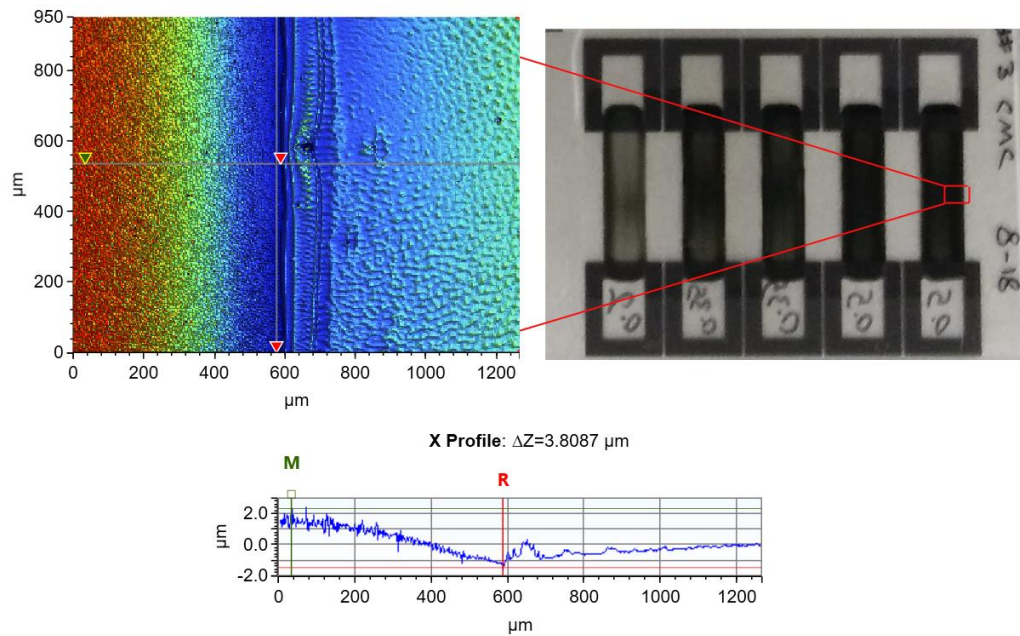
**Figure 51. The thickness of the CMC-dispersed coating obtained from run 15 diluted to 70% using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 2.8079  $\mu\text{m}$ .**



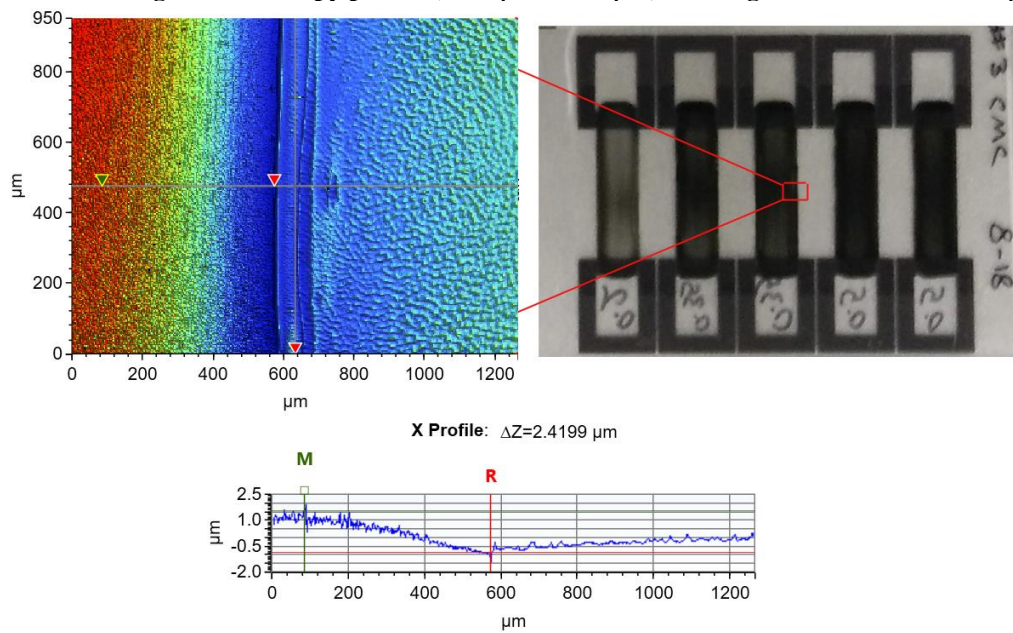
**Figure 52. The thickness of the CMC-dispersed coating obtained from run 15 diluted to 40% using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 1.6333  $\mu\text{m}$ .**

The thickness of matrix coatings obtained from Run 3 of the formulation in which CMC was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution is presented in Figure 53 through Figure 55. The thickness of the deposited matrix solution is measured to be 3.81, 2.42, and 1.18  $\mu\text{m}$ , respectively.

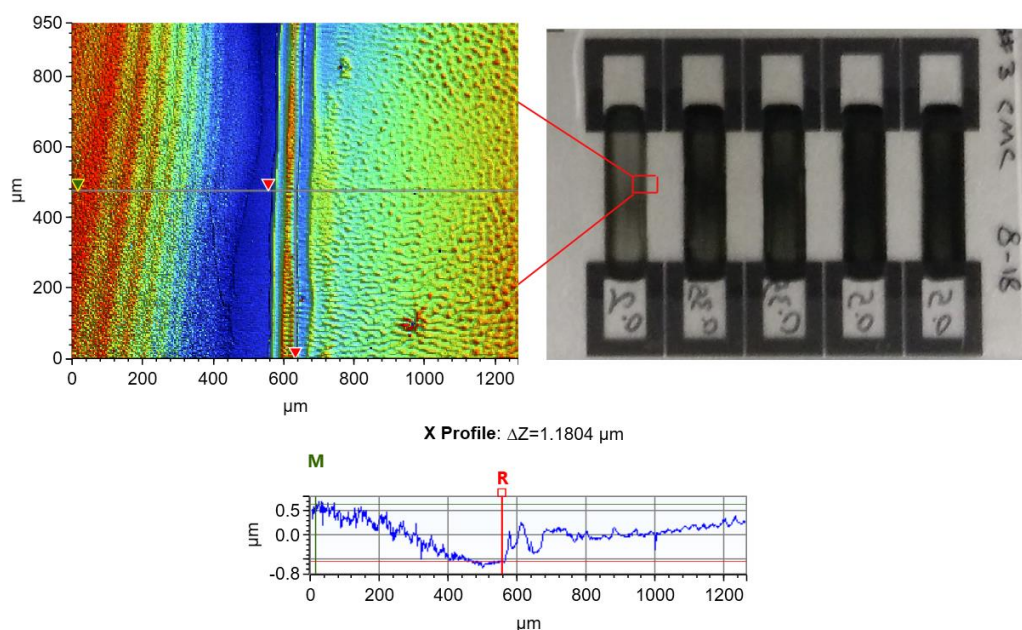




**Figure 53. The thickness of the CMC-dispersed coating obtained from run 3 at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 3.8087  $\mu\text{m}$ .**



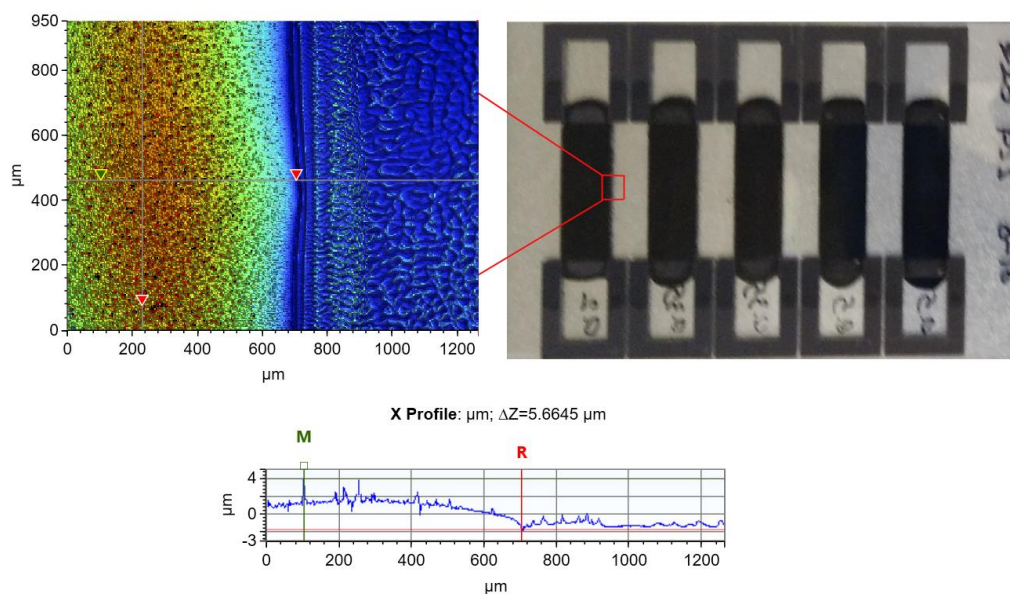
**Figure 54. The thickness of the CMC-dispersed coating obtained from run 3 diluted to 70% using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 2.4199  $\mu\text{m}$ .**



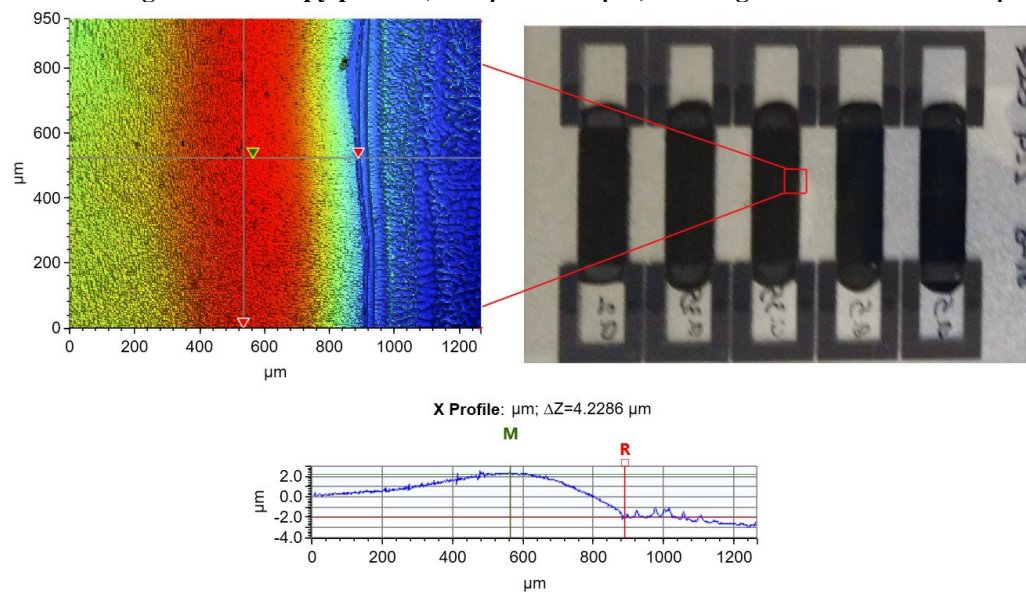
**Figure 55. The thickness of the CMC-dispersed coating obtained from run 3 diluted to 40% using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $1.1804 \mu\text{m}$ .**

The thickness of matrix coatings obtained from Run 20 of the formulation in which PSS was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution is presented in Figure 56 through Figure 58. The thickness of the deposited matrix solution is measured to be  $5.6645$ ,  $4.2286$ , and  $2.5407 \mu\text{m}$ , respectively. As expected, the thickness of the dried matrix coatings decreased as the matrix solutions were diluted.

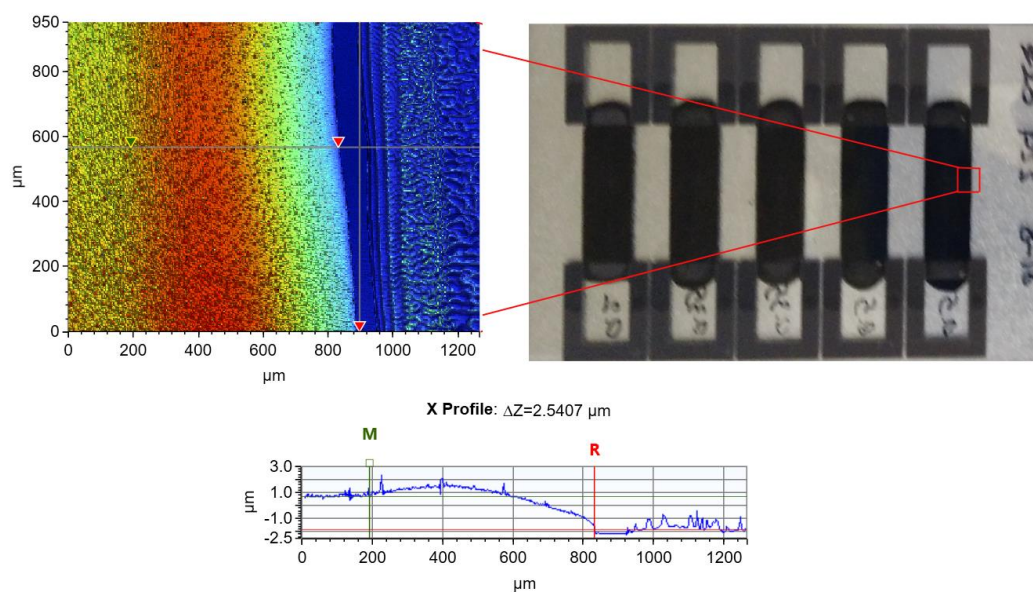




**Figure 56.** The thickness of the PSS-dispersed coating obtained from run 20 at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 5.6645  $\mu\text{m}$ .

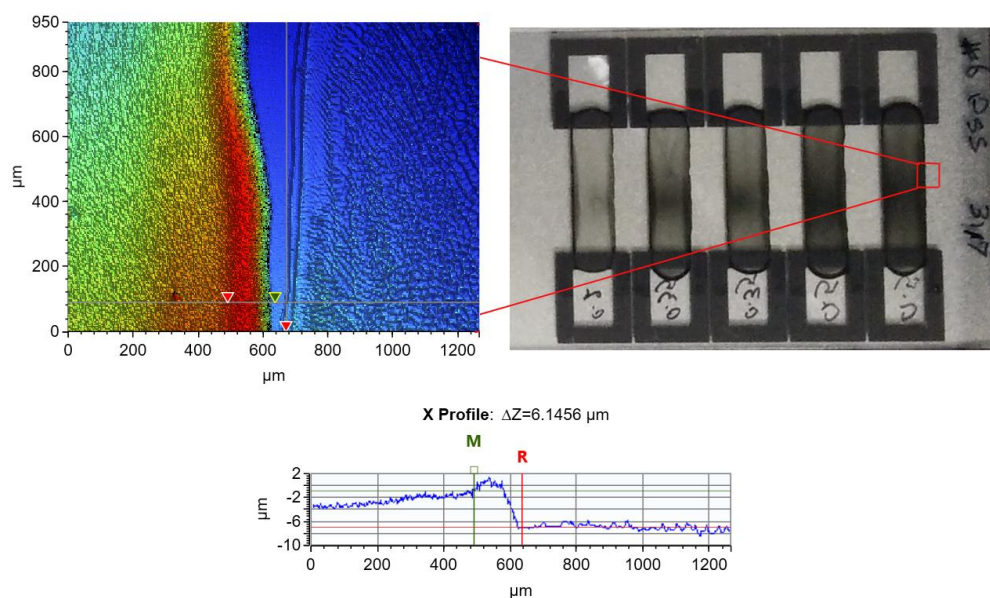


**Figure 57.** The thickness of the PSS-dispersed coating obtained from run 20 diluted to 70% using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 4.2286  $\mu\text{m}$ .

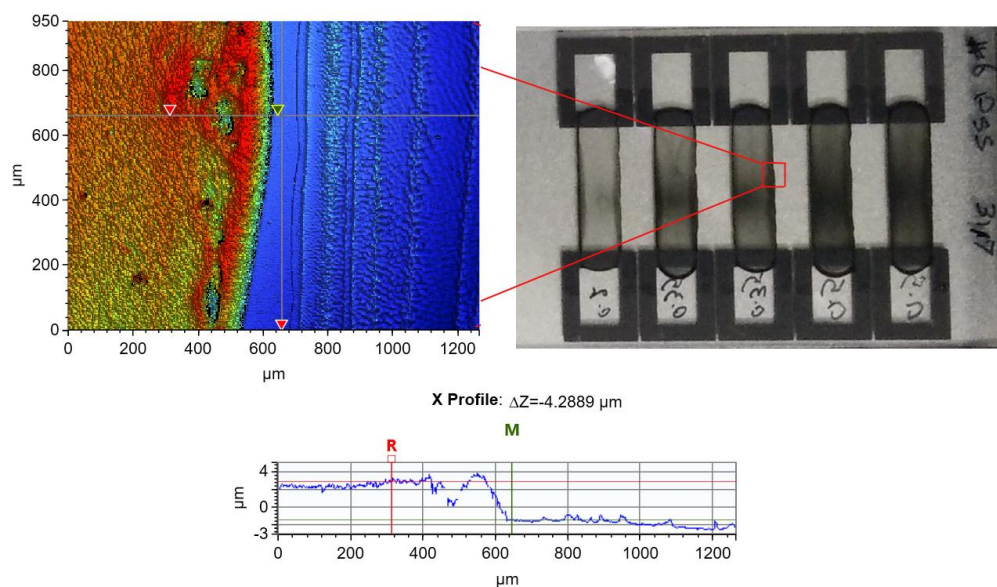


**Figure 58.** The thickness of the PSS-dispersed coating obtained from run 20 diluted to 40% using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $2.5407 \mu\text{m}$ .

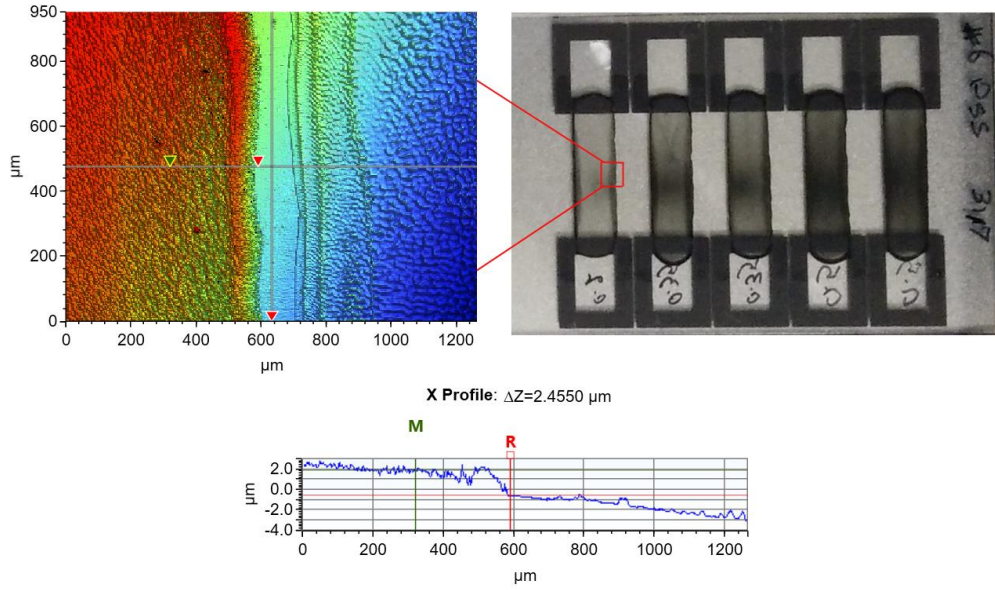
The thickness of matrix coatings obtained from Run 6 of the formulation in which PSS was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution is presented in Figure 59 through Figure 61. The thickness of the deposited matrix solution is measured to be 6.15, 4.29, and  $2.46 \mu\text{m}$ , respectively.



**Figure 59.** The thickness of the PSS-dispersed coating obtained from run 6 at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 6.1456  $\mu\text{m}$ .



**Figure 60.** The thickness of the PSS-dispersed coating obtained from run 6 diluted to 70% using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 4.2889  $\mu\text{m}$ .



**Figure 61. The thickness of the PSS-dispersed coating obtained from run 6 diluted to 40% using 2D microscopy profile (1250 μm × 950 μm) showing a thickness of 2.4550 μm.**

The thickness values obtained from the empirical measurements using the surface profilometry with the theoretical values of the thickness presented in Table 20 are found to be in a close agreement ( $\sim \pm 10\%$ ). Therefore the theoretical measurement of thickness can be used with a reasonable accuracy for prediction of the thickness for the deposited matrix solutions after drying.

### 2.3.5 Resistivity

By knowing the thickness of the deposited matrix solutions, and given that the dimensions of the deposited coatings are fixed for every channel, the resistivity of the coatings at different thicknesses can be calculated. The resistivity of such geometry can be obtained using the following equation [153, 154]:

$$\rho = \frac{V}{I} \frac{ah}{s} \quad (14)$$



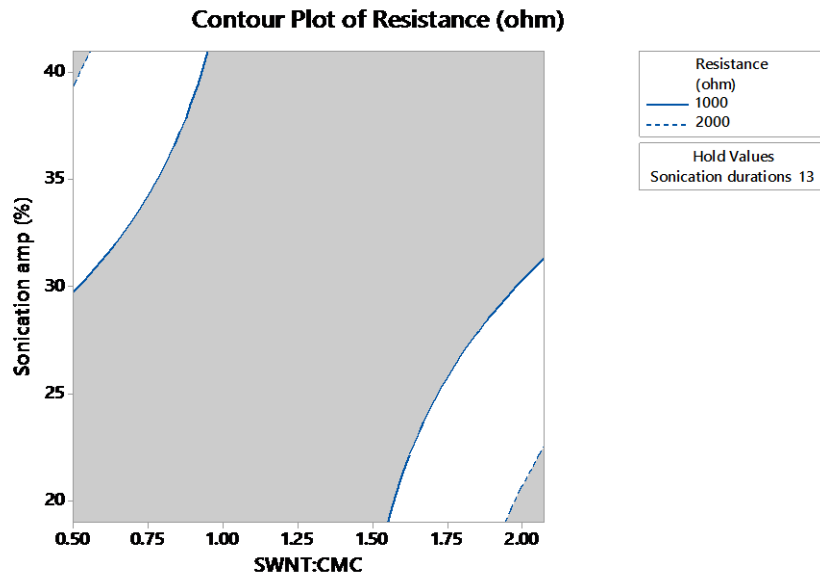
where  $\rho$  is the sample resistivity,  $V$  is the potential difference between the two voltage measurement probes,  $I$  is the current that passes through the sample,  $a$  and  $h$  are the sample width and height, and  $s$  is the length between the two probes. By using equation 14 the average resistivity of each sample has been calculated and presented in Table 21. Since the resistance of the electrodes/contacts are small compared to the resistance of the matrix coatings, a two point probe method was used to measure the resistance of the coatings [153, 154]. One would expect the resistivity of the film produced from each dilution of a sample to be similar, as they are in most cases.

**Table 21- Resistivity of the coatings at 100% (no dilution), 70%, and 40% dilutions after drying sorted from low to high for CMC at 100%(no dilution)**

Runs	X1 (SWNT:CMC)	X2 (Sonication amp (%))	X3 (Sonication durations (h))	CMC coating resistivity (ohm.cm)			PSS coating resistivity (ohm.cm)		
				100%	70%	40%	100%	70%	40%
15	2	40	24	0.0123	0.0134	0.0154	0.0735	0.0829	0.0742
12	2	40	2	0.0131	0.0126	0.0132	0.0026	0.0109	0.0118
5	1.25	30	13	0.0131	0.0123	0.0135	0.0422	0.0409	0.0516
1	1.25	30	13	0.0141	0.0140	0.0153	0.0549	0.0544	0.0700
10	1.25	19	13	0.0147	0.0145	0.0135	202.00	323.00	473.00
2	1.25	30	13	0.0153	0.0155	0.0169	0.0793	0.0962	0.0921
17	1.25	30	13	0.0154	0.0143	0.0177	0.1560	0.1470	0.2120
19	1.25	30	0.9	0.0169	0.0161	0.0158	0.2950	0.2520	0.3470
16	1.25	30	13	0.0170	0.0161	0.0175	0.0444	0.0428	0.0535
13	1.25	30	13	0.0177	0.0182	0.0205	0.1400	0.1370	0.1920
7	1.25	30	25.1	0.0212	0.0189	0.0197	0.0206	0.0212	0.0290
20	1.25	41	13	0.0234	0.0241	0.0247	0.0194	0.0199	0.0244
4	2.075	30	13	0.0313	0.0342	0.0320	0.0291	0.0261	0.0294
14	0.5	40	2	0.1230	0.1210	0.0963	0.0894	0.0807	0.0913
9	0.5	40	24	0.1580	0.1520	0.1360	0.0561	0.0519	0.0650
11	2	20	2	0.6970	0.6750	0.5220	29.700	31.900	39.300
18	0.425	30	13	0.7470	0.6810	0.5110	0.1190	0.1220	0.1590
6	0.5	20	24	2.5800	2.1100	1.5500	2680.0	1900.0	1670.00
8	2	20	24	3.7300	3.1700	3.1100	60.400	68.800	249.000
3	0.5	20	2	357.00	285.00	104.00	1010.0	753.00	894.00

### 2.3.6 RSM optimization

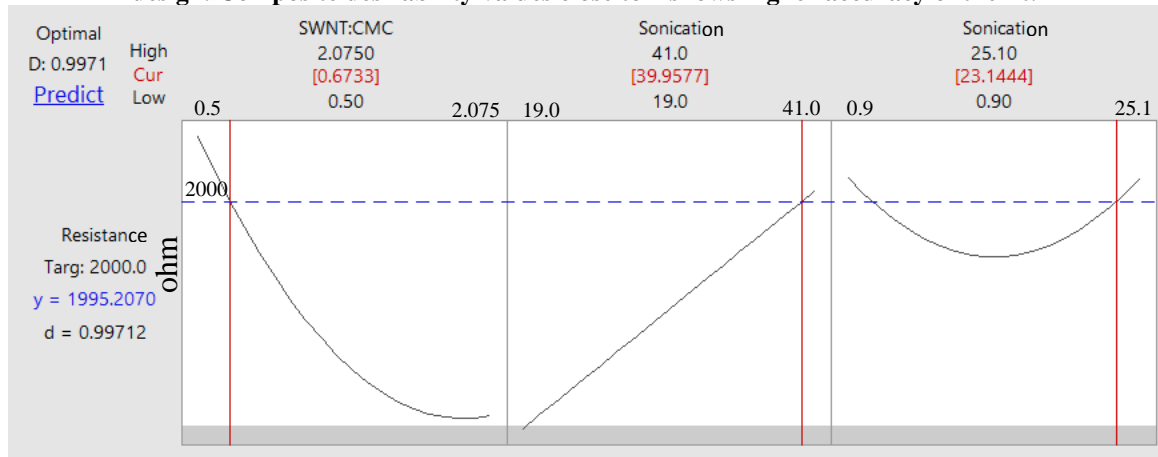
Thus far, as analyzed in the result and discussion section of this chapter, the predictability of the CMC-dispersed coatings, whether it be the resistance behavior or the particle size analysis, became apparent. The workability and the uniformity of the CMC-dispersed matrix solutions are also confirmed experimentally, as the CMC-dispersed SWNTs did not plug the filters and the coatings did not exhibit non-uniformity. These characteristics were not observed for the PSS-dispersed solution, leading the author to select the CMC as a superior dispersant for dispersing SWNTs in aqueous solutions. Using the response surface methodology (RSM) design of experiment, the behavior of the coatings can be modeled for dried matrix resistances up to several thousand ohms. As an example, the optimization of RSM was conducted based on the target resistance of  $\sim 2000\ \Omega$ . The optimized parameters for two possible solutions of achieving this goal are shown in the table insert in Figure 62. The unshaded areas on this plot identify several combinations of SWNT to CMC ratios and sonication amplitudes which result in a resistance between 1000 to 2000  $\Omega$ . Figure 63 also shows the overlay of the optimum conditions for Solution 1 at  $\sim 0.67$  SWNT:CMC ratio, sonication amplitude of  $\sim 40\%$ , and sonication duration of 23 hr. This modeling is valid for formulation sonication amplitudes in the 30-40% range.



Solution	SWNT:CMC	Sonication amp (%)	Sonication durations (hr)	Resistance (ohm) Fit	Composite Desirability
1	0.67	40	23	1995 <sup>1</sup>	1.00
2	0.67	40	2	2061	0.93

<sup>1</sup>(95% CI : 1590, 2400)

**Figure 62.** The condition optimized for the CMC solution at 40% showing the region of optimum (unshaded region) in which a resistance between 1000 to 2000  $\Omega$  can be achieved. The table insert shows the condition optimized for achieving a resistance of 2000  $\Omega$ , based on the response surface design. Composite desirability values close to 1 shows higher accuracy of the fit.



**Figure 63.** The main effect graph showing the optimized condition for achieving ~2000 ohm resistance using the CMC solution diluted to 40%. This figure refers to the Solution 1 in Figure 62 in order to achieve the target resistance of 2000  $\Omega$ , the parameters of SWNT:CMC, Sonication amp (%), and Sonication durations (hr) are set to 0.67, 40, and 23. Y axis represent resistance values (ohm).

## 2.4 Conclusion

Obtaining a uniform SWNT dispersion in aqueous solution using various sonication protocols along with poly(sodium 4-styrenesulfonate) (PSS) and carboxymethylcellulose (CMC) dispersants was investigated through a series of experiments designed with the aid of Response Surface Methodology (RSM). The resistance responses of the matrix solution formulations subjected to various conditions and coated and dried on UVO-treated PET substrate were modeled using this RSM design. The surface total free energy of the PET substrate used was increased from ~39 to 67 dyn/cm by UVO treatment for 5 minutes prior to use in these studies. From resistance measurements, the resistivity of the coatings was calculated by two methods, one using measured film thicknesses and the other using a thickness calculated from the mass of solids applied to the matrix film. The particle size distribution data using the dynamic light scattering technique was also correlated with the resistance response of each coating.

It was found that run 15 matrix solution (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr) resulted in dried matrix films with the lowest resistance (128  $\Omega$ ) and possessed an average particle size of ~115 nm. In contrast, the largest particle size (81% of the particles have an average particle size of ~5000 nm) for the set of formulation experiments in which CMC was used was observed in Run 3 (SWNT:CMC=0.5, Sonication amplitude=20%, and sonication duration= 2 hr) in which the highest dried film resistance was obtained. This perhaps indicates that a lower SWNT:CMC ratio (lower conductive:insulating components) is contributing to larger SWNT particle size that together are contributing to higher film resistivity. Higher



concentration of dispersant in comparison with the SWNTs may result in the dispersant wrapping in multiple layers around SWNTs which will hinder electron transport of conductive carbon nanotubes. This may also create larger particles which results in the removal of SWNTs from the solution after filtrations. The sonication factor plays an important role in the dispersion and conductivity of SWNTs. The analysis of variance of the particle sizes obtained in the PSS-dispersed matrix solutions shows that the sonication amplitude and its interaction with the concentration of PSS and the sonication amplitude are the only significant factors ( $\alpha=0.05$ ) among all the factors. Moreover, the sonication duration is the most significant factor in the analysis of variance of the particle sizes obtained in the CMC-dispersed matrix solutions.

3D optical microscopy was performed on the dried matrix films to study the morphology and roughness of the dried films thickness of the coatings as well as to measure the thickness of the coatings. Profilometry of Run 3 (CMC-dispersed coating) at 100% (no dilution) showed an average roughness of 0.274  $\mu\text{m}$  while profilometry of Run 15 at 100% (no dilution) showed an average roughness of 0.067  $\mu\text{m}$ . The same trend was noted for matrix coatings at higher dilutions (70% and 40%). Therefore, increased particle size of the matrix formulations had a strong correlation with increased roughness, and increased resistance of the CMC-dispersed coatings. Superior uniformity with low average roughness was observed for the coatings in which CMC was used as the dispersing agent in contrast to PSS.

The thickness of the coatings as measured by 3D optical microscope was found to decrease with the dilution of the matrix solution as would be expected due to reduced solids

content. As an example, the thickness of matrix coatings obtained from Run 15 of the formulation in which CMC was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution was measured to be 3.86, 2.81, and 1.63  $\mu\text{m}$ , respectively. Mass balance of each prepared solution was performed by determining the weight difference of the solids before and after the centrifugation and filtration and the thickness of the coatings after drying were calculated based on solids content, volume applied and areas. The results were compared with thickness measurements by the 3D optical microscope. The calculated values were found to be close to experimentally measured thickness measurements.

In conclusion, based on the coating quality and superior workability experimentally determined, CMC will be used in the matrix formulations as the dispersant for the next chapter (chapter 3) of this dissertation. An experiment will be designed to investigate several promising matrix formulations produced from stable SWNT/CMC dispersions in this chapter. Aliquots of the SWNT dispersion will be mixed with solutions of antibody, and glycerol. The resistance of dried matrix semi-solid films of these formulations with will be assessed with the same methodology and procedures as used in this chapter.

# EFFECT OF ANTIBODY AND GLYCEROL ADDITION TO MATRIX FORMULATIONS

## 3.1 Objective

In chapter 2 of this dissertation, obtaining uniform SWNT dispersion in aqueous solution using various sonication protocols along with poly(sodium 4-styrenesulfonate) (PSS) and carboxymethylcellulose (CMC), two dispersing agents, and other components comprising sodium azide and phosphate buffer was studied. In this chapter, selected SWNT dispersions will be combined with antibody and other constituents (such as humectant glycerol) and deposited within test sensor channels, dried into a semi-solid/fluid film, and resistance tested using a constant microamp current. Once promising matrix solution formulations were identified, further characterizations were performed. These characterizations focused on selecting promising matrix films that could be further characterized by a combination of a 3D optical microscope (white light interferometer Contour GT-I) for profilometry, to look at gross features of the cured matrix layer formulation, and SEM to attempt to show physical interaction of micro-components of the formulations within the dried matrixes.

## **3.2 Methodology**

### **3.2.1 Matrix formulation**

#### **3.2.1.1 Single-walled nano-tube (SWNT) dispersion**

Three promising formulations from chapter 2 of this dissertation were selected based on the minimum resistance measurements obtained within each SWNT:CMC ratio categories (0.5, 1.25, and 2). The same procedures as explained in Research Phase 1 (Chapter 2) were employed to make 10 ml of each of these three solutions. These solutions were then used as the stock solutions for this research phase of the dissertation.

#### **3.2.1.2 Matrix formulation materials (with SWNT, CMC, antibody and glycerol)**

The purpose of the experiments in chapter 2 was to find matrix film formulations that had a dried resistance of less than 2000 ohms and to correlate resistance with film properties. Three matrix formulations produced from stable SWNT dispersions in phase 1 research (Chapter 2) were selected. Aliquots of the SWNT dispersion were mixed with solutions of antibody and glycerol. The resistance of dried matrix semi-solid films of these formulations with antibody was assessed with the same procedure as mentioned in in phase 1 research. Matrix formulation parameters investigated were:

1. Three composition ratios (1:1, 1:1.6, 1:4) of two matrix components (SWNT:glycerol);
2. Four levels of antibody (rabbit anti-mouse IgG), (1:0.1, 1:0.014, 1:0.0075) (SWNT:Ab);

3. And two different matrix film thicknesses achieved by using 100% strength formulations and dilution of formulations with water to 25% (75% dilution).

The dried channel matrices produced in this phase of the dissertation contained rabbit anti-mouse IgG – as this is a relatively inexpensive antibody to work with (all IgG antibodies have a molecular weight of ~150,000 daltons). In all cases the matrix antibody is physically adsorbed onto and entrapped by the nanotube matrix. The sensor and formulation materials needed for this study are presented in Table 2 (Research Phase 1). In addition, antibody (Sigma Aldrich) I-5006 rabbit anti-mouse-IgG and Glycerol (Sigma Aldrich) were used to prepare the matrix formulation for Research Phase 2.

Aliquots of dispersed nanotubes in solution (0.15 ml) were deposited on the test channels (see Figure 21 and Figure 22 and Figure 23) and let dry in a desiccant chamber. Two different matrix film thicknesses based on the theoretically calculated densities of the varying components in the formulations for the remaining solids (SWNT and CMC) was achieved by preparing diluted aliquots of the dispersions prepared in this section at 100% (no dilution), and 25 % (75% dilution) using water before coating.

The resistance of all matrix-coated plates/channels was measured following drying in a desiccant chamber.

### **3.2.2 RSM**

In this experiment, the design was composed of two factors (SWNT:Glycerol ratio, and SWNT:antibody ratio), each with two levels (low, high), and a total of 13 runs were carried out to optimize the chosen variables. For the purpose of statistical computations,

the two independent variables were denoted as  $x_1$ , and  $x_2$ , respectively. The range of the values used for the levels of each factor investigated in the experiments was selected based on the preliminary experiments and are listed in Table 22. The quadratic equation can be used for this methodology to fit the response variables:

$$Y = \beta_0 + \sum_{i=1}^k \beta_i x_i + \sum_{i=1}^k \beta_{ii} x_i^2 + \sum_{i < j} \beta_{ij} x_i x_j \quad (15)$$

where  $Y$ ,  $k$ ,  $\beta_0$ ,  $\beta_i$ ,  $\beta_{ii}$ , and  $\beta_{ij}$  are the predicted response variable, number of variables, a constant term, coefficients of the linear parameters, coefficients of the quadratic parameters, and coefficients of the interaction parameters, respectively. The coefficient of determination ( $R^2$ ) can be estimated to test the certainty of the above polynomial model [151]. In this dissertation, RSM was employed to investigate the effect of SWNT:Glycerol ratio, and SWNT:antibody on the resistance of the prepared matrix solutions.

**Table 22- Factors and levels used in this experimental design (DOE)**

<b>Factor</b>	<b>Low level (-1)</b>	<b>High level (+1)</b>
<b>SWNT:Glycerol (<math>X_1</math>)</b>	0.25	1
<b>SWNT:antibody (<math>X_2</math>)</b>	10	133

Different mixtures using SWNT:Glycerol and SWNT:Antibody ratios were prepared for select matrix formulations developed in research phase 1 (formulations 12, 14, and 17 using CMC) and mixed with different ratios of SWNT:Glycerol (0.25 and 1) and SWNT:antibody (10 and 133). The matrix preparation parameters were set according to the response surface methodology (RSM) experimental design, with the parameters

tabulated in Table 23 through Table 28. Dilution of the prepared solutions (diluted to 25%), as explained earlier, and 0.15 ml of the resulting solutions were deposited into designated channels (in duplicate). The prepared templates were then dried in a desiccant chamber for 48 hr. Subsequently, the surface profile of the select dried matrix was obtained and analyzed.

**Table 23- The design of experiment based on RSM showing the factor levels and their responses. The two independent variables were denoted as  $x_1$ , and  $x_2$ , which represent SWNT:Glycerol, and SWNT:antibody, respectively for the CMC Run 12 matrix solution (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2) at 100% (no dilution)**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:GLY)	<i>X<sub>2</sub></i> ( SWNT:Ab )	<i>Resistance (ohm)</i>	<i>Predicted</i> <i>Resistance (ohm)</i> (eq. 16)
12-1	0.625	71.7	1827	2294
12-2	0.625	71.7	2322	2294
12-3	0.25	10.0	180753	145507
12-4	0.625	10.0	5283	4965
12-5	0.625	71.7	2421	2294
12-6	1	133.3	474	792
12-7	0.625	71.7	1902	2294
12-8	0.25	71.7	32026	3956*
12-9	1	71.7	1265	629
12-10	0.625	133.3	1634	1316
12-11	1	10.0	1841	2159
12-12	0.25	133.3	14718	1841*
12-13	0.625	71.	2362	2294

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 24- The design of experiment based on RSM showing the factor levels and their responses. The two independent variables were denoted as  $x_1$ , and  $x_2$ , which represent SWNT:Glycerol, and SWNT:antibody, respectively for the CMC Run 12 matrix solution (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2) diluted to 25%.**

<i>Runs</i>	<i>X<sub>1</sub></i> (SWNT:GLY)	<i>X<sub>2</sub></i> ( SWNT:Ab )	<i>Resistance (ohm)</i>	<i>Predicted</i> <i>Resistance (ohm)</i> (eq. 17)
12-1	0.625	71.7	3804	5869
12-2	0.625	71.7	7452	5869
12-3	0.25	10.0	184197	179903
12-4	0.625	10.0	6753	5869

Table 24 - continue

<b>12-5</b>	0.625	71.7	4355	5869
<b>12-6</b>	1	133.3	1135	1295
<b>12-7</b>	0.625	71.7	6753	5869
<b>12-8</b>	0.25	71.7	135247	8666*
<b>12-9</b>	1	71.7	3392	3073
<b>12-10</b>	0.625	133.3	5637	5869
<b>12-11</b>	1	10.0	4692	4852
<b>12-12</b>	0.25	133.3	38606	10443*
<b>12-13</b>	0.625	71.7	6331	5869

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 25- The design of experiment based on RSM showing the factor levels and their responses. The two independent variables were denoted as  $x_1$ , and  $x_2$ , which represent SWNT:Glycerol, and SWNT:antibody, respectively for the CMC Run 14 matrix solution (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2) at 100% (no dilution)**

<b><i>Runs</i></b>	<b><i>X<sub>1</sub></i></b> <b><i>(SWNT:GLY)</i></b>	<b><i>X<sub>2</sub></i></b> <b><i>( SWNT:Ab )</i></b>	<b><i>Resistance (ohm)</i></b>	<b><i>Predicted</i></b> <b><i>Resistance (ohm)</i></b> <b><i>(eq. 18)</i></b>
<b>14-1</b>	0.625	71.7	43576	45248
<b>14-2</b>	0.625	71.7	64756	45248
<b>14-3</b>	0.25	10.0	1650440	1283512
<b>14-4</b>	0.625	10.0	138495	145292
<b>14-5</b>	0.625	71.7	57602	45248
<b>14-6</b>	1	133.3	108749	4337*
<b>14-7</b>	0.625	71.7	28654	45248
<b>14-8</b>	0.25	71.7	4337	4337
<b>14-9</b>	1	71.7	5727	19322
<b>14-10</b>	0.625	133.3	8738	15535
<b>14-11</b>	1	10.0	13822	7024
<b>14-12</b>	0.25	133.3	97941	137666
<b>14-13</b>	0.625	71.7	17777	45211

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 26- The design of experiment based on RSM showing the factor levels and their responses. The two independent variables were denoted as  $x_1$ , and  $x_2$ , which represent SWNT:Glycerol, and SWNT:antibody, respectively for the CMC Run 14 matrix solution (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2) diluted to 25%.**

<b><i>Runs</i></b>	<b><i>X<sub>1</sub></i></b> <b><i>(SWNT:GLY)</i></b>	<b><i>X<sub>2</sub></i></b> <b><i>( SWNT:Ab )</i></b>	<b><i>Resistance (ohm)</i></b>	<b><i>Predicted</i></b> <b><i>Resistance (ohm)</i></b> <b><i>(eq. 19)</i></b>
<b>14-1</b>	0.625	71.7	17574	17838
<b>14-2</b>	0.625	71.7	21209	17838
<b>14-3</b>	0.25	10.0	356148	292113



Table 26 - continue

<b>14-4</b>	0.625	10.0	60731	142287
<b>14-5</b>	0.625	71.7	21104	17838
<b>14-6</b>	1	133.3	9190	7557
<b>14-7</b>	0.625	71.7	20363	17838
<b>14-8</b>	0.25	71.7	51550	28118
<b>14-9</b>	1	71.7	6862	7557
<b>14-10</b>	0.625	133.3	9890	17838
<b>14-11</b>	1	10.0	9983	7557
<b>14-12</b>	0.25	133.3	31483	28119
<b>14-13</b>	0.625	71.7	10159	17838

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 27- The design of experiment based on RSM showing the factor levels and their responses. The two independent variables were denoted as  $x_1$ , and  $x_2$ , which represent SWNT:Glycerol, and SWNT:antibody, respectively for the CMC Run 17 matrix solution (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13) at 100% (no dilution)**

<b><i>Runs</i></b>	<b><i>X<sub>1</sub></i></b> <b><i>(SWNT:GLY)</i></b>	<b><i>X<sub>2</sub></i></b> <b><i>( SWNT:Ab )</i></b>	<b><i>Resistance (ohm)</i></b>	<b><i>Predicted</i></b> <b><i>Resistance (ohm)</i></b> <b><i>(eq. 20)</i></b>
<b>17-1</b>	0.625	71.7	16360	21645
<b>17-2</b>	0.625	71.7	21138	21645
<b>17-3</b>	0.25	10.0	968476	921401
<b>17-4</b>	0.625	10.0	46313	43557
<b>17-5</b>	0.625	71.7	27339	21645
<b>17-6</b>	1	133.3	4162	3835
<b>17-7</b>	0.625	71.7	22043	21645
<b>17-8</b>	0.25	71.7	702175	37042*
<b>17-9</b>	1	71.7	5570	6224
<b>17-10</b>	0.625	133.3	2488	268
<b>17-11</b>	1	10.0	8939	8612
<b>17-12</b>	0.25	133.3	551042	4350*
<b>17-13</b>	0.625	71.7	15832	21645

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

**Table 28- The design of experiment based on RSM showing the factor levels and their responses. The two independent variables were denoted as  $x_1$ , and  $x_2$ , which represent SWNT:Glycerol, and SWNT:antibody, respectively for the CMC Run 17 matrix solution (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13) diluted to 25%.**

<b><i>Runs</i></b>	<b><i>X<sub>1</sub></i></b> <b><i>(SWNT:GLY)</i></b>	<b><i>X<sub>2</sub></i></b> <b><i>( SWNT:Ab )</i></b>	<b><i>Resistance (ohm)</i></b>	<b><i>Predicted</i></b> <b><i>Resistance (ohm)</i></b> <b><i>(eq. 21)</i></b>
<b>17-1</b>	0.625	71.7	59871	101831
<b>17-2</b>	0.625	71.7	273081	23543

Table 28 - continued

<b>17-3</b>	0.25	10.0	1376718	1041319
<b>17-4</b>	0.625	10.0	175400	101831
<b>17-5</b>	0.625	71.7	131665	101831
<b>17-6</b>	1	133.3	11912	23544
<b>17-7</b>	0.625	71.7	80214	<i>101831</i>
<b>17-8</b>	0.25	71.7	215690	101831
<b>17-9</b>	1	71.7	13971	23544
<b>17-10</b>	0.625	133.3	25774	101831
<b>17-11</b>	1	10.0	21869	23544
<b>17-12</b>	0.25	133.3	46424	23544
<b>17-13</b>	0.625	71.7	138061	101831

\*Fitted values in italic denotes an extremely unusual point relative to predictor levels used to fit the model

### 3.3 Results and Discussion

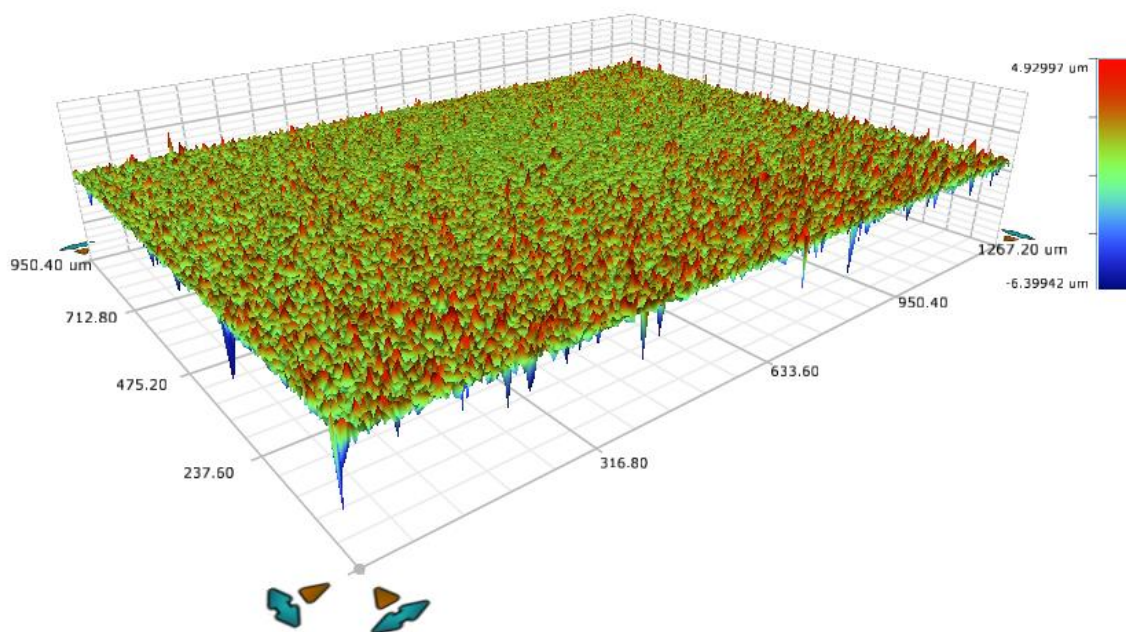
#### 3.3.1 Matrix solution selection

The stock matrix solution was selected based on the minimum resistance and minimum sonication duration for each SWNT to CMC ratios from the experiments in phase 1 research. Considering the factors mentioned and the resistance results from Table 4 through Table 9, Run 14 was selected from set matrix solutions in which SWNT:CMC ratio was 0.5. Run 17 was selected from set matrix solutions in which SWNT:CMC ratio was 1.25. Run 12 was selected from set matrix solutions in which SWNT:CMC ratio was 2. These selected solution will be referred to as CMC-14 (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2), CMC-17 (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13), and CMC-12 (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2), respectively, for phase 2 research discussion. The corresponding resistance results were 1069, 794, and 148  $\Omega$ , respectively. The surface profile of run 12, 14, and 17 at 100%, 70%, and 40% is shown in Figure 64 through Figure 72. As mentioned earlier in phase 1, the surface roughness of the samples decreases with

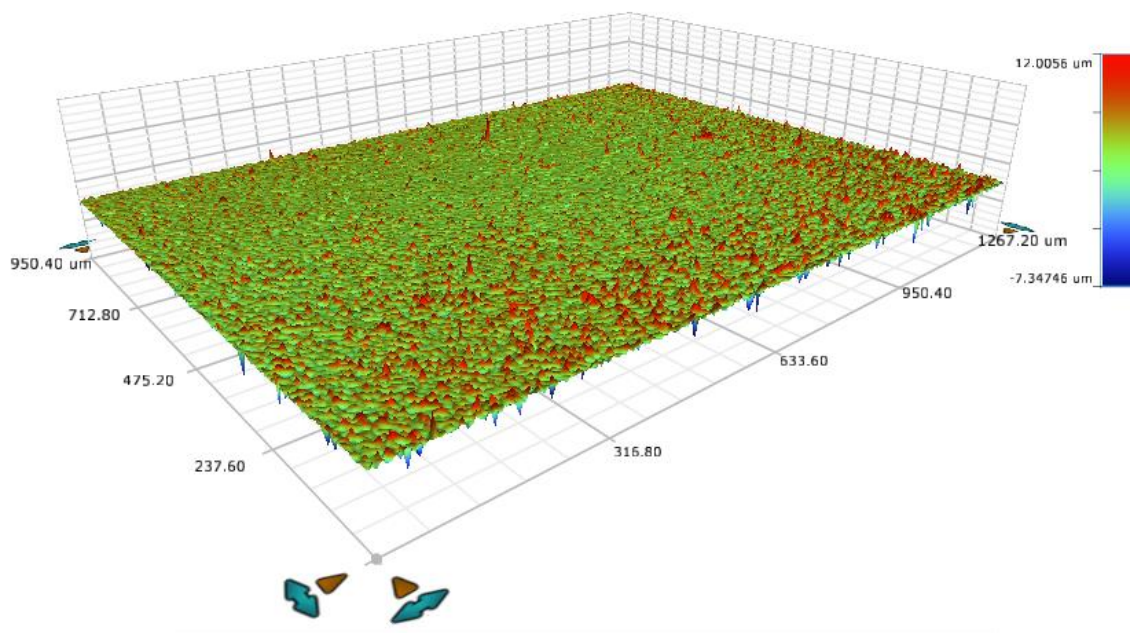
the increase in the dilutions Table 29. This fact can also be observed of the samples from Run 12, 14, and 17.

**Table 29- Summary of coating thickness and roughness of the selected matrix formulations from research phase 1.**

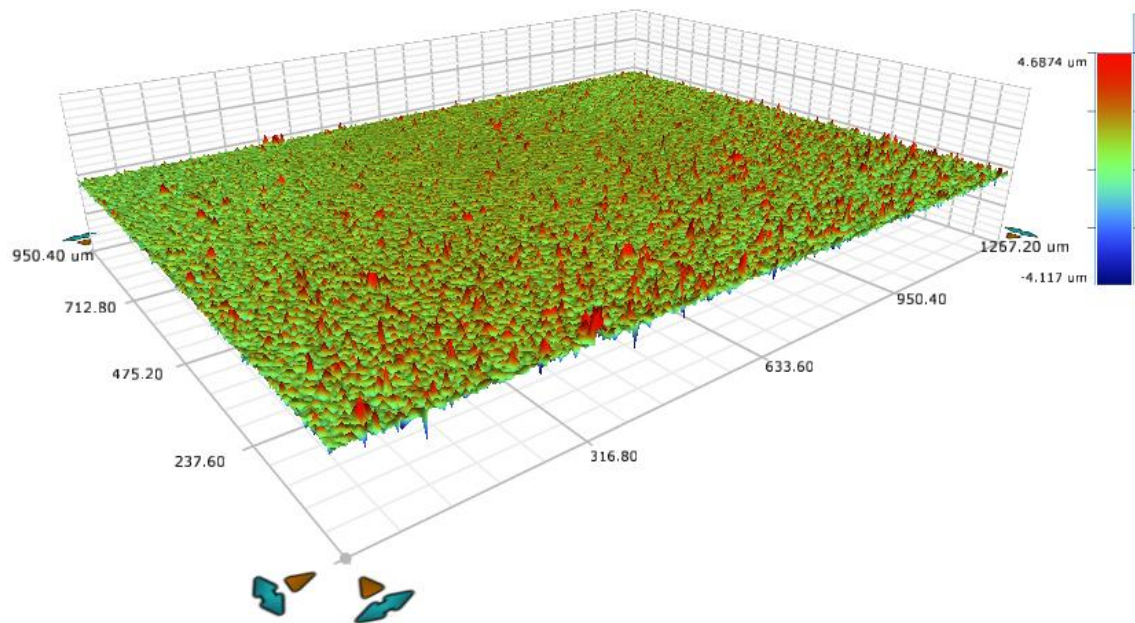
	dilution	roughness ( $\mu\text{m}$ )	Thickness* ( $\mu\text{m}$ )	roughness/thickness ratio	Resistance (ohm)
<u>CMC-12</u> (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2)	100%	0.220	3.56	0.062	148
	70%	0.189	2.49	0.076	202
	40%	0.119	1.42	0.084	372
<u>CMC-14</u> (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2)	100%	0.258	4.60	0.056	1069
	70%	0.208	3.22	0.065	1498
	40%	0.168	1.84	0.091	2095
<u>CMC-17</u> (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13)	100%	0.113	4.19	0.027	147
	70%	0.102	2.93	0.035	195
	40%	0.066	1.68	0.039	423
<i>*calculated thickness based on mass balance as described in chapter 2</i>					



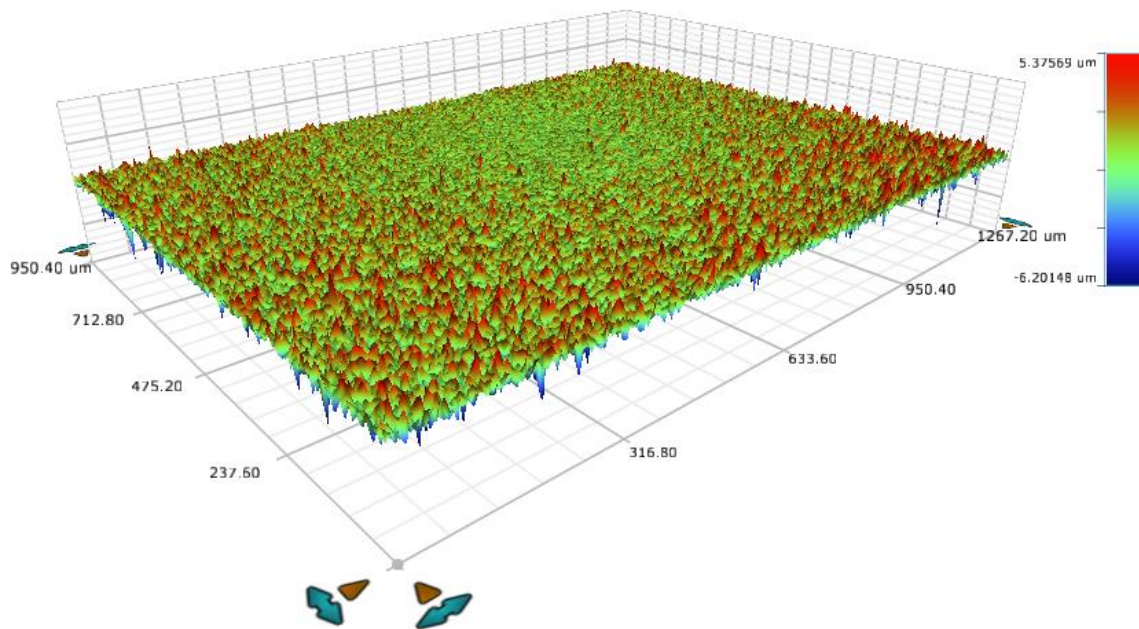
**Figure 64. Profilometry of Run 12, showing a uniform CMC-dispersed coating at 100% (no dilution) with an average roughness of 0.220  $\mu\text{m}$ .**



**Figure 65. Profilometry of Run 12, showing a uniform CMC-dispersed coating diluted to 70% with an average roughness of 0.189  $\mu\text{m}$ .**

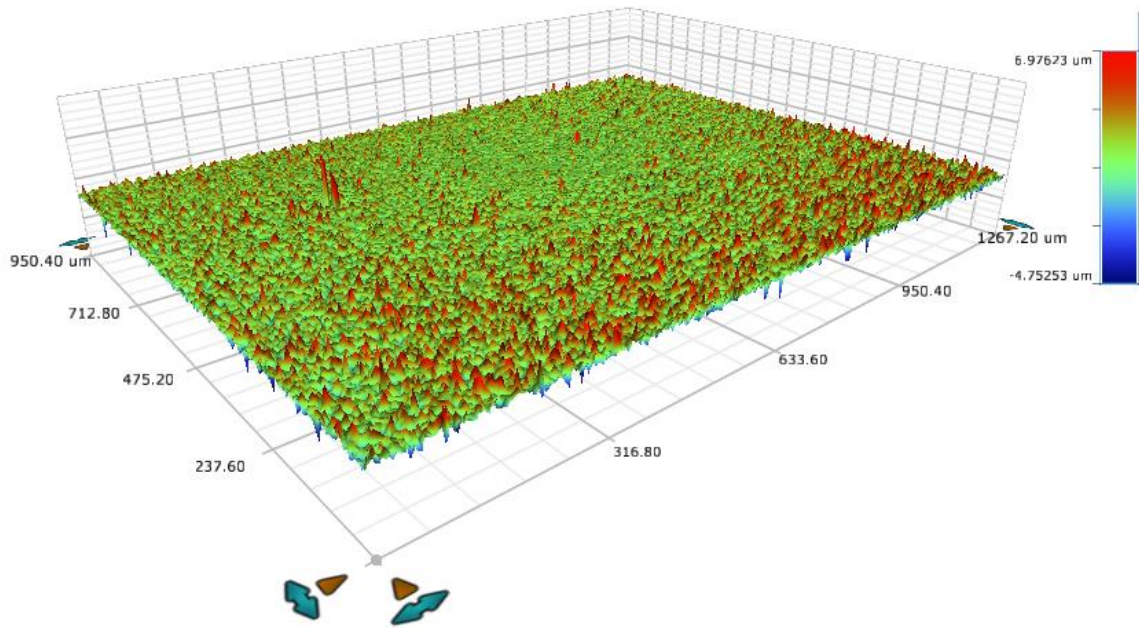


**Figure 66. Profilometry of Run 12, showing a uniform CMC-dispersed coating diluted to 40% with an average roughness of 0.119  $\mu\text{m}$ .**

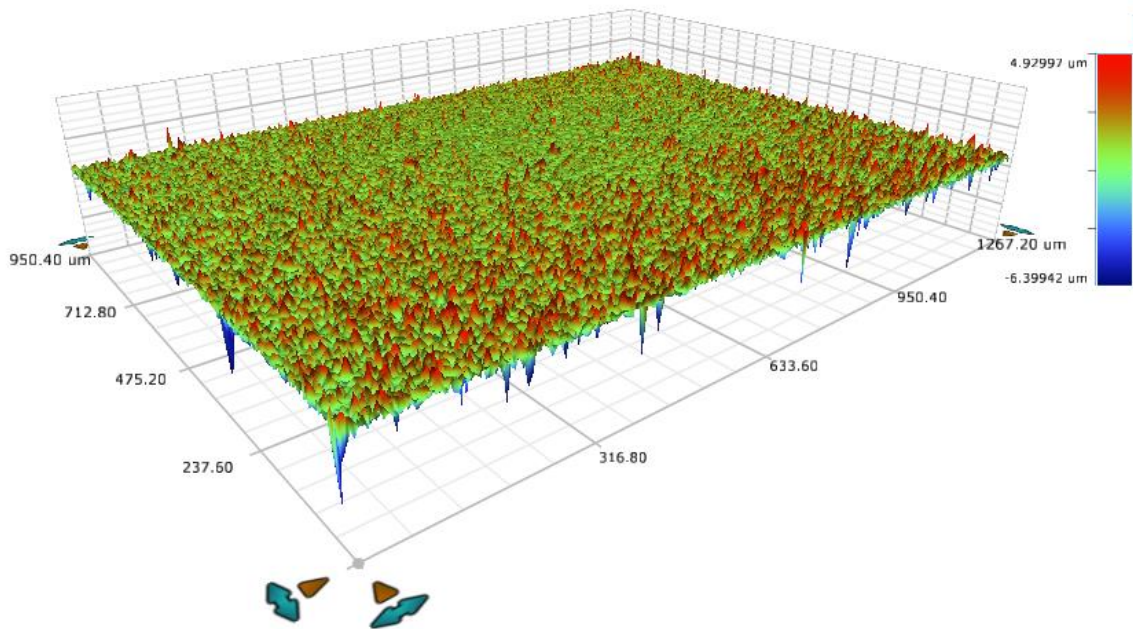


**Figure 67. Profilometry of Run 14, showing a uniform CMC-dispersed coating at 100% (no dilution) with an average roughness of 0.258  $\mu\text{m}$ .**

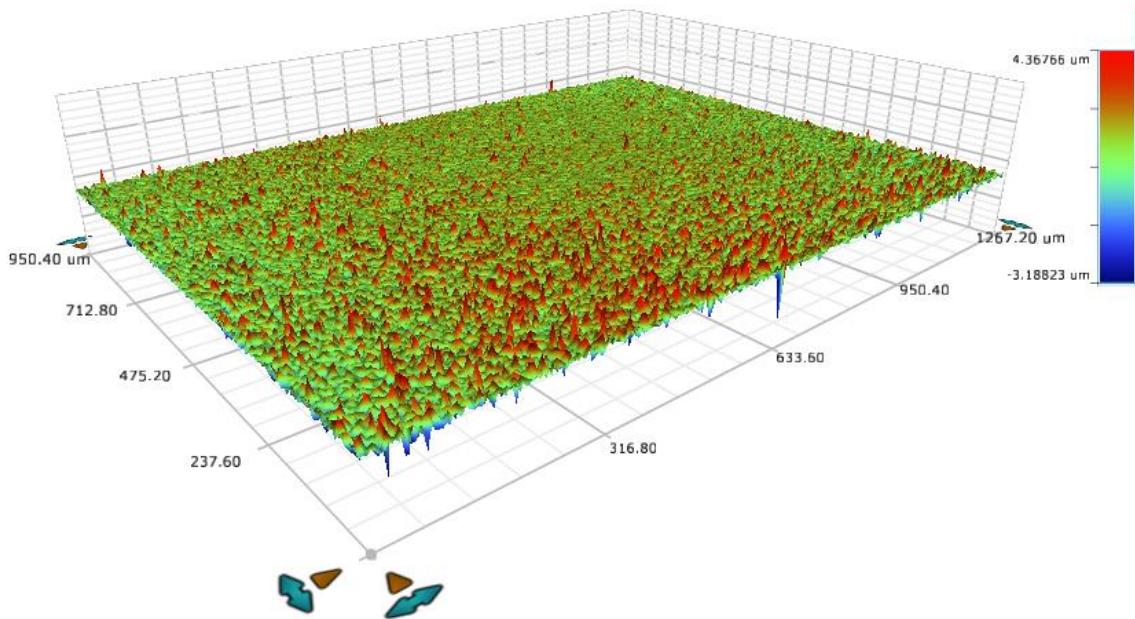




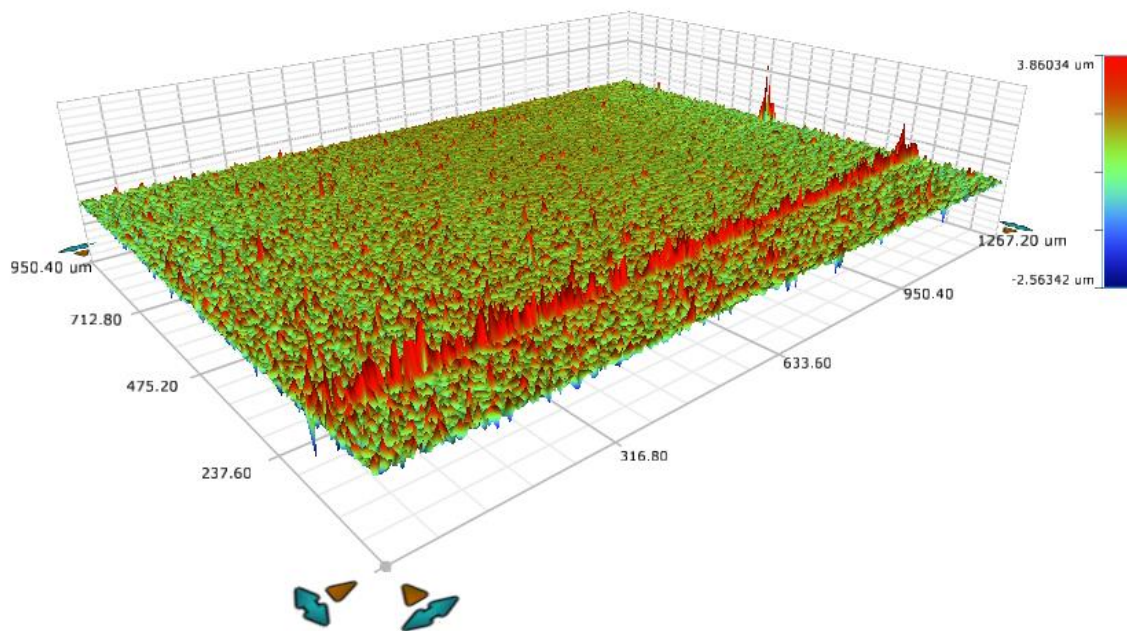
**Figure 68. Profilometry of Run 14, showing a uniform CMC-dispersed coating diluted to 70% with an average roughness of 0.208  $\mu\text{m}$ .**



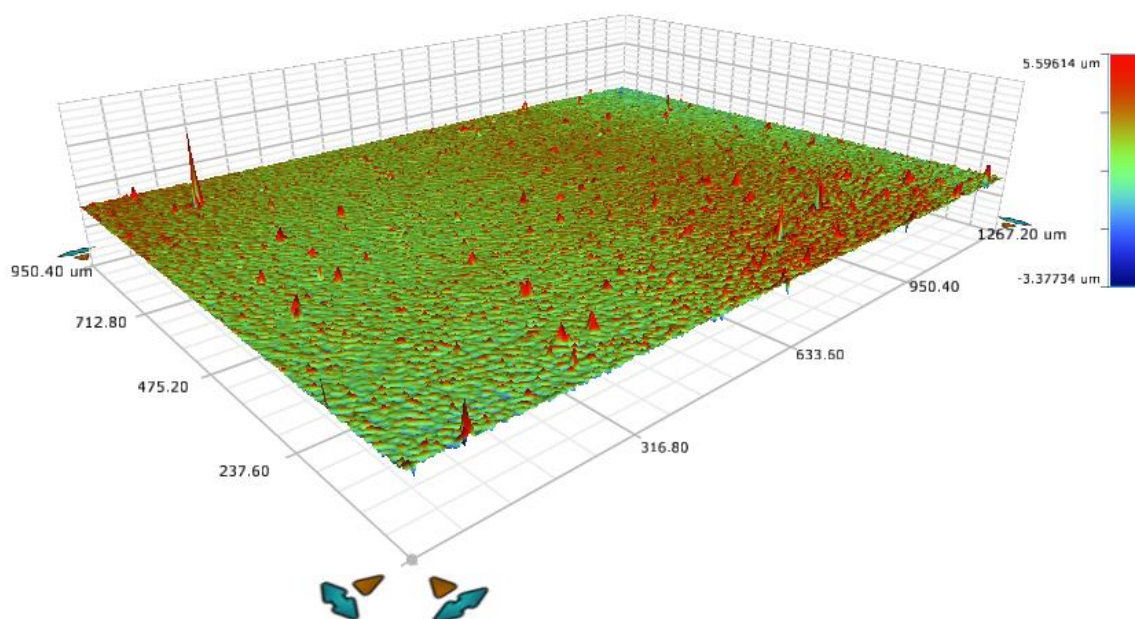
**Figure 69. Profilometry of Run 14, showing a uniform CMC-dispersed coating diluted to 40% with an average roughness of 0.168  $\mu\text{m}$ .**



**Figure 70. Profilometry of Run 17, showing a uniform CMC-dispersed coating at 100% (no dilution) with an average roughness of 0.113  $\mu\text{m}$ .**



**Figure 71. Profilometry of Run 17, showing a uniform CMC-dispersed coating diluted to 70% with an average roughness of 0.102  $\mu\text{m}$ .**



**Figure 72. Profilometry of Run 17, showing a uniform CMC-dispersed coating diluted to 40% with an average roughness of 0.066  $\mu\text{m}$ .**

### 3.3.2 Resistance response analysis of variance

The dimensions of the test templates are all equal for all of the experiments, enabling us to use the resistance results as the basis of the electrical properties comparison for all the experiments. The minimum average resistance of 474  $\Omega$  for the set of experiments in which formulation CMC-12 was used at 100% (no dilution) was noted in Run 12-6 (see Table 23, 12-6) corresponding to the minimum concentration of glycerol and antibody at: SWNT:Gly ratio of 1, SWNT:Ab ratio of 133. The maximum average resistance of 1.65 M $\Omega$  for the set of experiments in which formulation CMC-14 was used at 100% was noted in Run 12-3 (see Table 25) corresponding to the maximum concentration of glycerol and antibody at: SWNT:Gly ratio of 0.25, and SWNT:Ab ratio



of 10. These results show that increasing concentrations of both glycerol and antibody increases the resistance of the matrix film.

The analysis of variance, presented in Table 30 through Table 35, shows that the resistance of the matrix solutions containing CMC as the dispersant are significantly dependent on the SWNT:Gly, and SWNT:Ab ratios as well as the interaction terms. Considering the t-statistics for each model term and the provisions of the RSM, the following models were generated from the data (coded values) and can be used to fit the resistance responses:

12 at 100% (no dilution):

$$\text{Resistance (ohm)} = 277540 - 546895 X_1 - 1575 X_2 + 255837 X_1 \times X_1 + 1780 X_1 \times X_2 \quad (16)$$

12 diluted to 25%

$$\text{Resistance (ohm)} = 354379 - 757014 X_1 - 1366 X_2 + 393546 X_1 \times X_1 + 1536 X_1 \times X_2 \quad (17)$$

14 at 100%

$$\text{Resistance (ohm)} = 1999590 - 1998354 X_1 - 26903 X_2 + 80.1 X_2 \times X_2 + 17811 X_1 \times X_2 \quad (18)$$

14 diluted to 25%

$$\text{Resistance (ohm)} = 444497 - 434550 X_1 - 5403 X_2 + 15.33 X_2 \times X_2 + 3501 X_1 \times X_2 \quad (19)$$

17 at 100%

$$\text{Resistance (ohm)} = 1900768 - 4425497 X_1 - 4048 X_2 + 2501328 X_1 \times X_1 + 4461 X_1 \times X_2 \quad (20)$$

17 diluted to 25%

$$\text{Resistance (ohm)} = 1567645 - 1730114 X_1 - 12948 X_2 + 14274 X_1 \times X_2 \quad (21)$$

**Table 30- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC-12 matrix solution at 100% (no dilution)**

<b>Source</b>	<b>Resistance (ohm)</b>		
	<b>DF</b>	<b>F-Value</b>	<b>P-Value</b>
<b>Model</b>	4	9.16	0.004
<b>Linear</b>	2	9.84	0.007
<b>SWNT:GLY</b>	1	12.34	0.008
<b>SWNT:Ab</b>	1	7.35	0.027
<b>Square</b>	1	6.37	0.036
<b>SWNT:GLY*SWNT:GLY</b>	1	6.37	0.036
<b>2-Way Interaction</b>	1	10.57	0.012
<b>SWNT:GLY*SWNT:Ab</b>	1	10.57	0.012

**Table 31- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC-12 matrix solution diluted to 25%**

<b>Source</b>	<b>Resistance (ohm)</b>		
	<b>DF</b>	<b>F-Value</b>	<b>P-Value</b>
<b>Model</b>	4	24.78	0.001
<b>Linear</b>	2	29.98	0.001
<b>SWNT:GLY</b>	1	50.37	0.001
<b>SWNT:Ab</b>	1	9.58	0.006
<b>Square</b>	1	25.35	0.001
<b>SWNT:GLY*SWNT:GLY</b>	1	25.35	0.001
<b>2-Way Interaction</b>	1	13.8	0.003
<b>SWNT:GLY*SWNT:Ab</b>	1	13.8	0.003

**Table 32- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC-14 matrix solution at 100% (no dilution)**

<b>Source</b>	<b>Resistance (ohm)</b>		
	<b>DF</b>	<b>F-Value</b>	<b>P-Value</b>
<b>Model</b>	4	17.45	0.012
<b>Linear</b>	2	18.73	0.023
<b>SWNT:GLY</b>	1	24	0.035
<b>SWNT:Ab</b>	1	13.46	0.039
<b>Square</b>	1	12.57	0.071
<b>SWNT:GLY*SWNT:GLY</b>	1	12.57	0.071
<b>2-Way Interaction</b>	1	19.78	0.014
<b>SWNT:GLY*SWNT:Ab</b>	1	19.78	0.014

**Table 33- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC-14 matrix solution diluted to 25%**

Source	Resistance (ohm)		
	DF	F-Value	P-Value
<b>Model</b>	4	8.46	0.003
<b>Linear</b>	2	9.37	0.004
<b>SWNT:GLY</b>	1	9.83	0.006
<b>SWNT:Ab</b>	1	8.92	0.010
<b>Square</b>	1	4.19	0.052
<b>SWNT:GLY*SWNT:GLY</b>	1	4.19	0.052
<b>2-Way Interaction</b>	1	10.93	0.008
<b>SWNT:GLY*SWNT:Ab</b>	1	10.93	0.008

**Table 34- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC-17 matrix solution at 100% (no dilution)**

Source	Resistance (ohm)		
	DF	F-Value	P-Value
<b>Model</b>	4	118.13	0.001
<b>Linear</b>	2	153.31	0.001
<b>SWNT:GLY</b>	1	291.23	0.001
<b>SWNT:Ab</b>	1	15.39	0.001
<b>Square</b>	1	145.05	0.001
<b>SWNT:GLY*SWNT:GLY</b>	1	145.05	0.001
<b>2-Way Interaction</b>	1	20.87	0.001
<b>SWNT:GLY*SWNT:Ab</b>	1	20.87	0.001

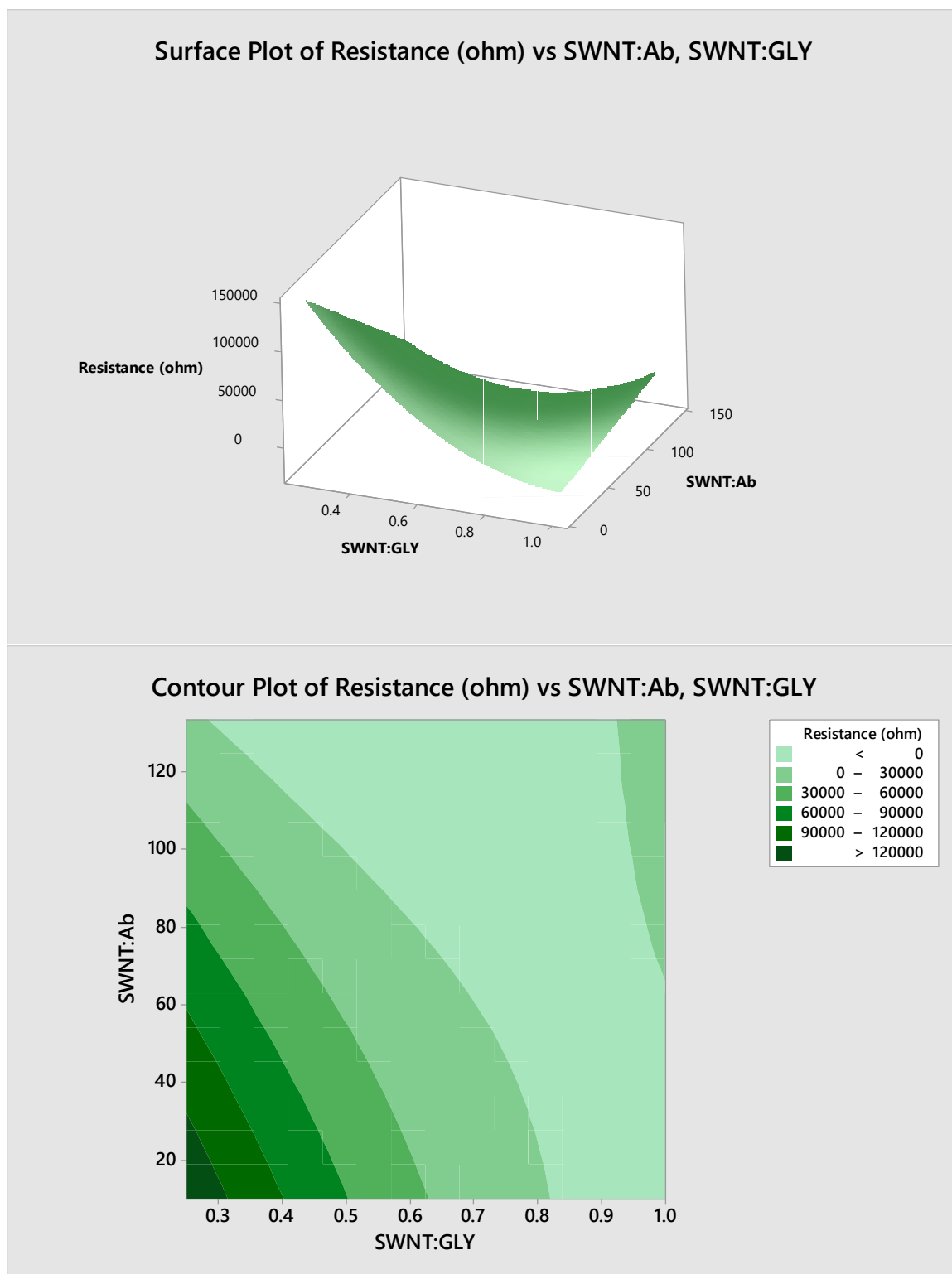
**Table 35- Analysis of variance for the resistance and particle size of the samples obtained under various preparation conditions for the CMC-17 matrix solution diluted to 25%**

Source	Resistance (ohm)		
	DF	F-Value	P-Value
<b>Model</b>	4	40.47	0.001
<b>Linear</b>	2	52.55	0.001
<b>SWNT:GLY</b>	1	68.33	0.001
<b>SWNT:Ab</b>	1	36.78	0.001
<b>Square</b>	1	18.34	0.003
<b>SWNT:GLY*SWNT:GLY</b>	1	18.34	0.003
<b>2-Way Interaction</b>	1	38.43	0.001
<b>SWNT:GLY*SWNT:Ab</b>	1	38.43	0.001

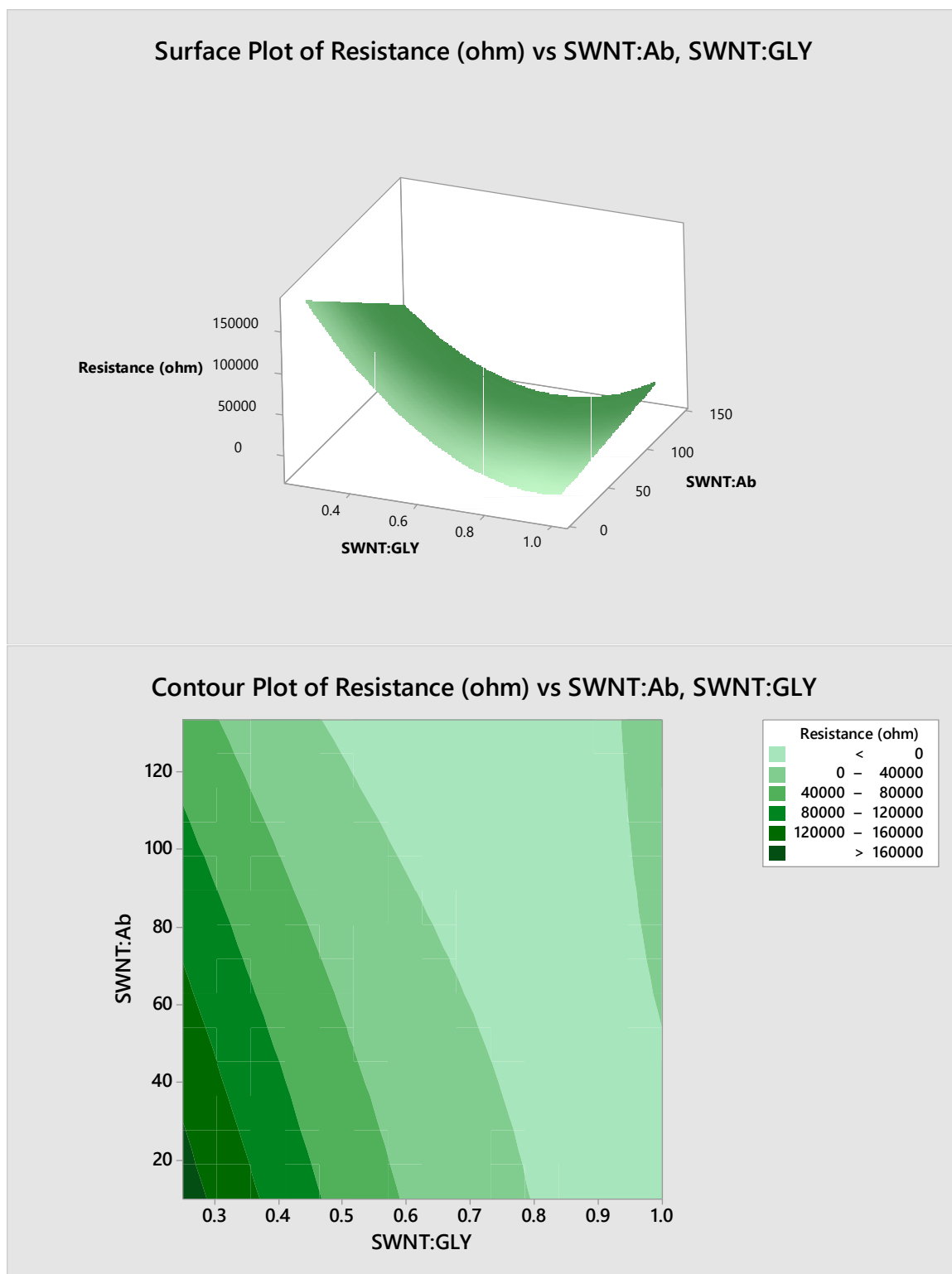
The surface and contour plots of the most significant process parameters are presented in Figure 73 to Figure 78. These figures show the effect of each factor on the resistance of the matrix solution. In general, the higher the SWNT:Glycerol/Antibody ratio, the lower the resistance of the corresponding matrix solution. This seems logical as more glycerol or antibody should act as insulator-like materials (low-conductivity) between SWNT-to-SWNT contacts. Table 36 shows the predicted resistances using the regression models equations (16) – (21) and the measured experimental resistances for select coatings. The predicted resistance values are in close agreement with the measured resistance for matrix solutions diluted to 25% as well as matrix solutions at no dilutions (100%) in majority of cases. Therefore, the fitted model can be used to predict the relative magnitude of the resistance of the dried matrix coatings.

**Table 36- Comparison of the coatings' measured resistance and predicted resistance for select coatings**

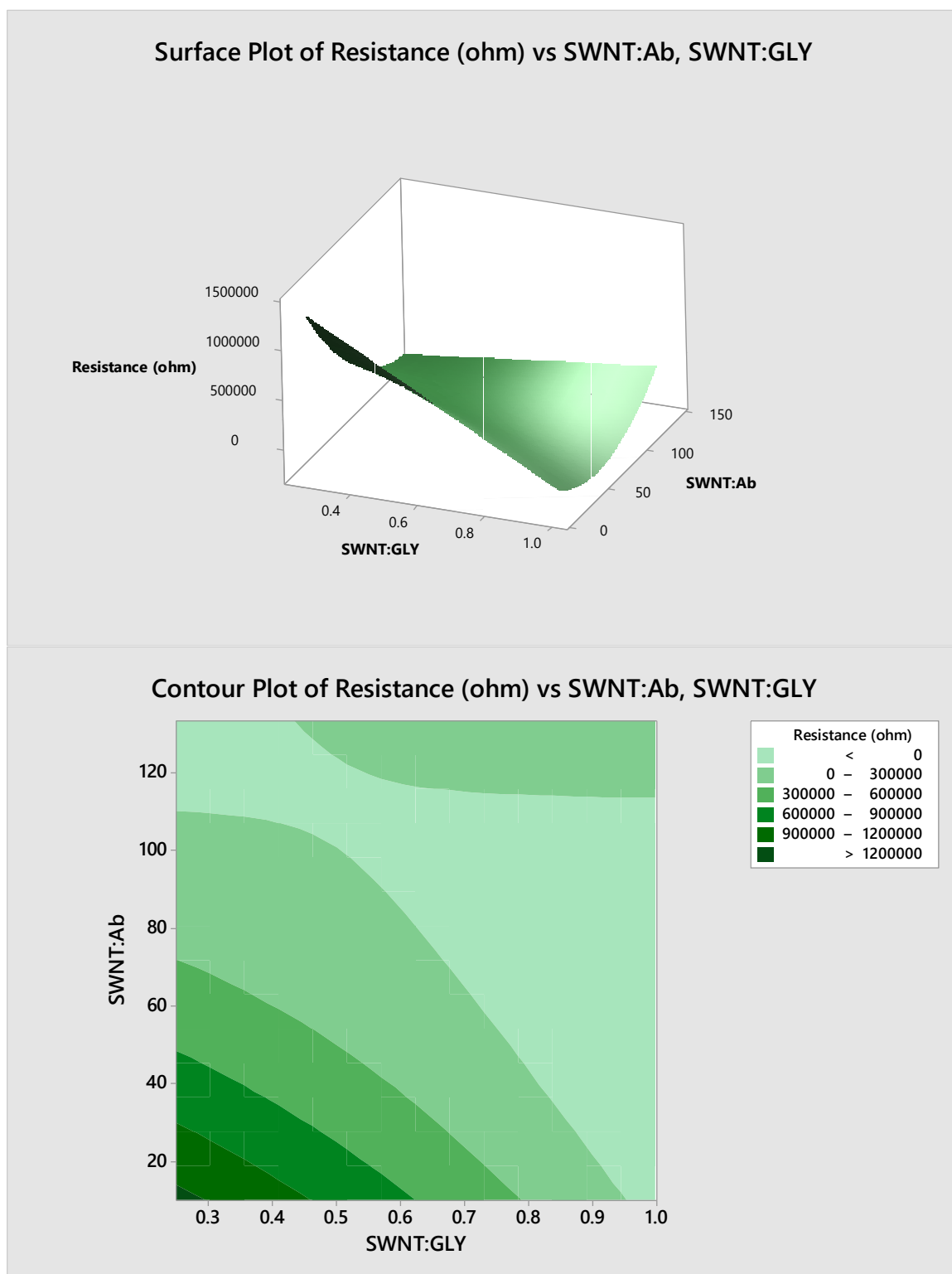
Run	SWNT:GLY	SWNT:Ab	dilution	<u>Resistance (<math>\Omega</math>)</u>			<u>Predicted Resistance (ohm)</u>		
				CMC-12	CMC-17	CMC-14	CMC-12	CMC-17	CMC-14
3	0.25	10	100%	180753	968476	1650440	145507	921401	1283512
4	0.625	10	100%	5283	46313	138495	4965	43557	145292
11	1	10	100%	1841	8939	13822	2159	8612	7024
	$\infty$	$\infty$	100%	148	147	1069			
11	1	10	100%	1841	8939	13822	2159	8612	7024
9	1	71	100%	1265	5570	5727	629	6224	19322
6	1	133	100%	474	4162	4337	792	3835	4337
	$\infty$	$\infty$	100%	148	147	1069			
3	0.25	10	25%	184197	1376718	356148	179903	1041319	292113
4	0.625	10	25%	6753	175400	60731	5869	101831	142287
11	1	10	25%	4692	21869	9983	4852	23544	7557
	$\infty$	$\infty$	25%	n.d.	n.d.	n.d.			
SWNT:CMC ratio =				2.00	1.25	0.50	2.00	1.25	0.50



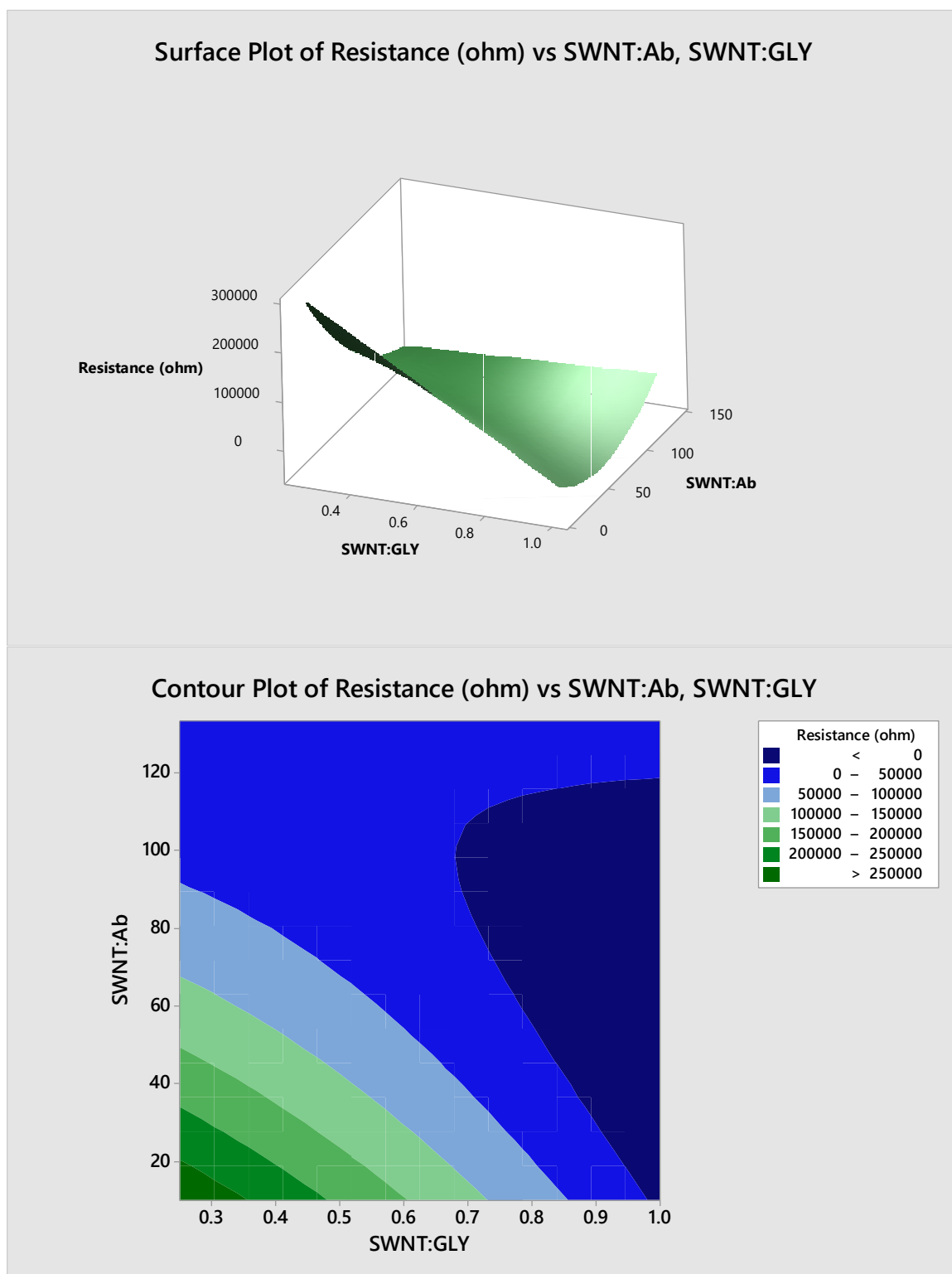
**Figure 73. Contour plots and three-dimensional response surface of the resistance response for CMC-12 (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2) at 100% (no dilution)**



**Figure 74. Contour plots and three-dimensional response surface of the resistance response for CMC-12 (SWNT:CMC= 2, Sonication amp (%)=40, Sonication durations (h)=2) diluted to 25%**



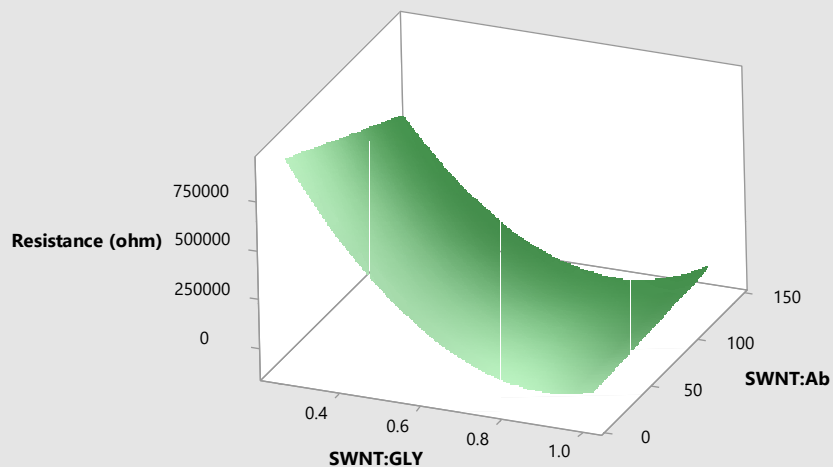
**Figure 75. Contour plots and three-dimensional response surface of the resistance response for CMC-14 (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2) at 100% (no dilution)**



**Figure 76. Contour plots and three-dimensional response surface of the resistance response for CMC-14 (SWNT:CMC= 0.5, Sonication amp (%)=40, Sonication durations (h)=2) diluted to 25%**



Surface Plot of Resistance (ohm) vs SWNT:Ab, SWNT:GLY



Contour Plot of Resistance (ohm) vs SWNT:Ab, SWNT:GLY

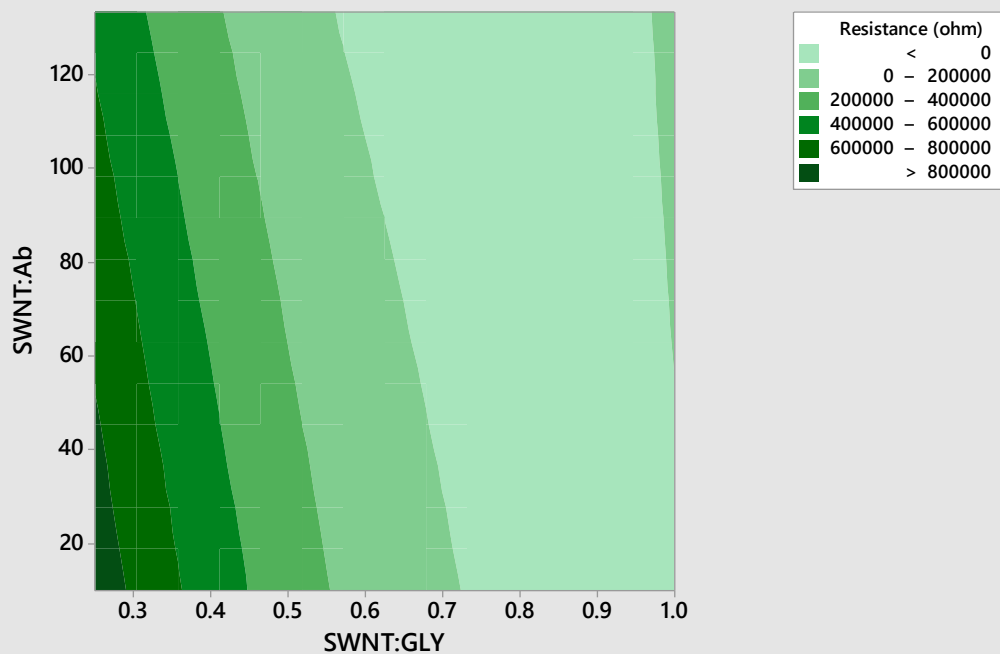
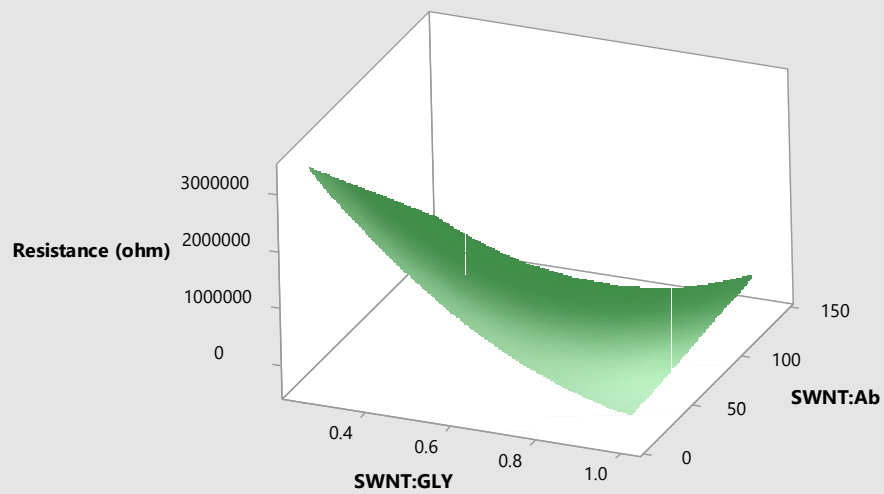


Figure 77. Contour plots and three-dimensional response surface of the resistance response for CMC-17 (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13) at 100% (no dilution)

Surface Plot of Resistance (ohm) vs SWNT:Ab, SWNT:GLY



Contour Plot of Resistance (ohm) vs SWNT:Ab, SWNT:GLY

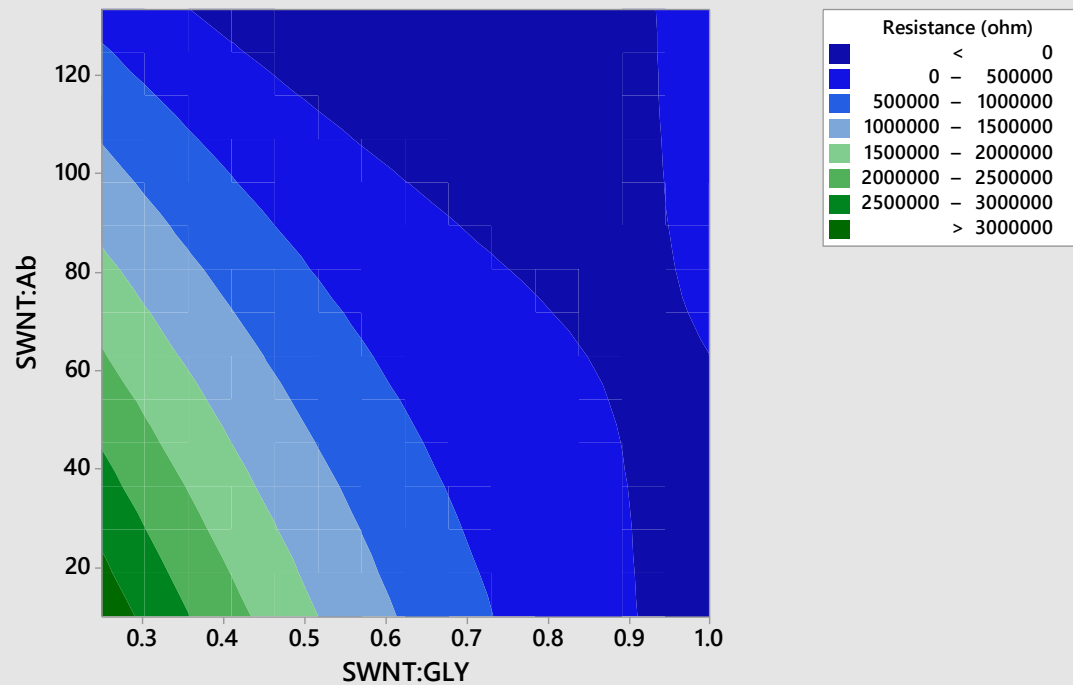


Figure 78. Contour plots and three-dimensional response surface of the resistance response for CMC-17 (SWNT:CMC= 1.25, Sonication amp (%)=30, Sonication durations (h)=13) diluted to 25%

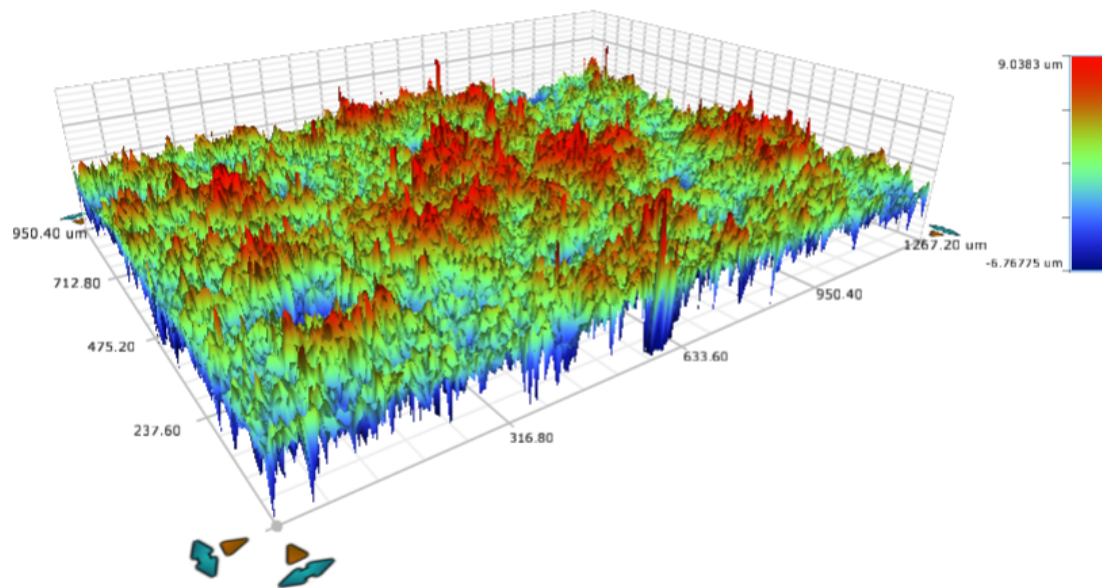
### 3.3.3 Surface profile

The topography of the deposited matrix solutions in the channels was investigated using an optical profiler.

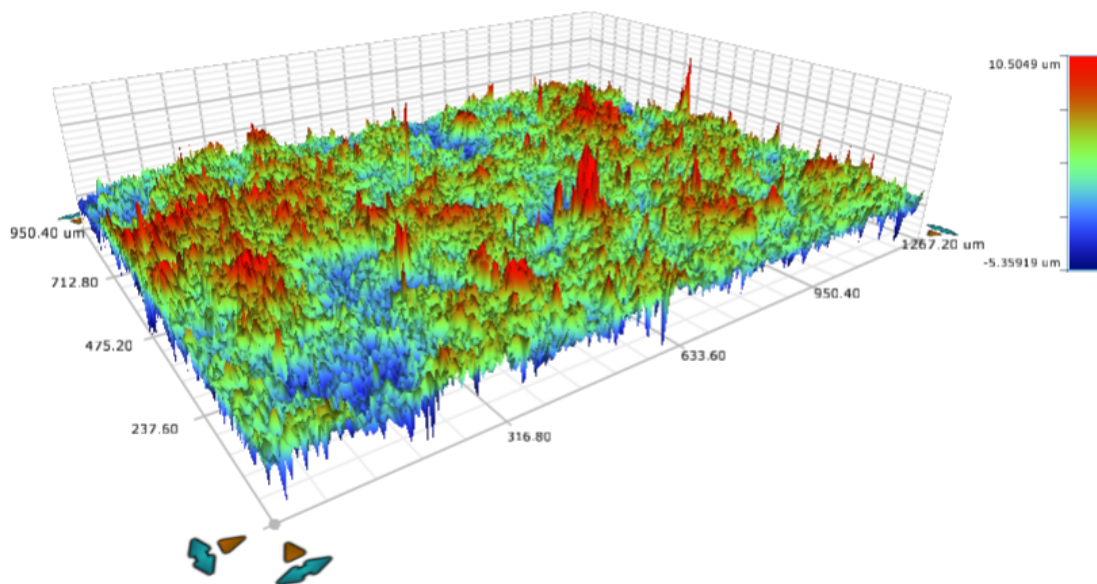
The average surface roughnesses of selected coatings are presented in Table 37 for comparison. In this table the coatings prepared from CMC-12, 17 and 14 formulations with select SWNT:Glycerol ratios, SWNT:antibody ratios, and water dilution were compared. The surface profile of the set of experiments in which CMC-12, CMC-14, and CMC-17 matrix formulations, at 100% (no dilution), and 25% dilution are presented in Figure 79 through Figure 94. From Table 37 and Figure 79 - Figure 94, the lowest resistances can be observed for the set of experiments in which the CMC-12 solution was used (see Table 23). The set of experiments in which solution CMC-17 was used resulted in the midrange resistances (see Table 25 and Table 37). CMC-14 solutions resulted in set of matrix samples with the highest resistances (see Table 27 and Table 37). It is important to note from these tables that the average roughness of the samples decreased with a decrease in the glycerol concentration. The roughness of the samples also decreased with an increase in the concentration of antibody. This is probably because antibody molecules can further disperse carbon nanotubes and therefore, they decrease aggregation, leading to a smoother surface. The roughness of the coating also decreased with matrix dilution. Interesting CMC-14 resistances decreased with dilution unlike other formulations and contrary to logic.

**Table 37- Comparison of the coatings' average roughness and its effect on resistance for select coatings**

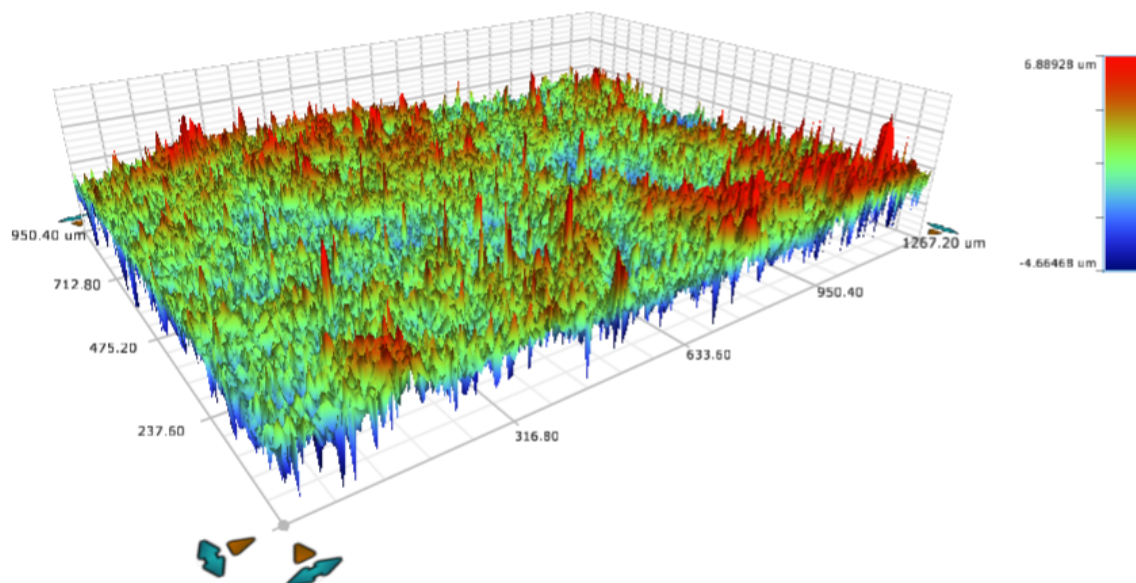
Run	SWNT:GLY	SWNT:Ab	dilution	Resistance ( $\Omega$ )			Roughness ( $\mu\text{m}$ )		
				CMC-12	CMC-17	CMC-14	CMC-12	CMC-17	CMC-14
3	0.25	10	100%	180753	968476	1650440	1.048		
4	0.625	10	100%	5283	46313	138495	0.816		
11	1	10	100%	1841	8939	13822	0.740	0.611	0.504
	$\infty$	$\infty$	100%	148	147	1069	0.220	0.113	0.258
11	1	10	100%	1841	8939	13822	0.740	0.611	0.504
9	1	71	100%	1265	5570	5727	0.624	0.455	0.432
6	1	133	100%	474	4162	4337	0.449	0.419	0.387
	$\infty$	$\infty$	100%	148	147	1069	0.220	0.113	0.258
3	0.25	10	25%	184197	1376718	356148	0.583		
4	0.625	10	25%	6753	175400	60731	0.424		
11	1	10	25%	4692	21869	9983	0.412	0.285	0.192
	$\infty$	$\infty$	25%	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
SWNT:CMC ratio =				2.00	1.25	0.50	2.00	1.25	0.50



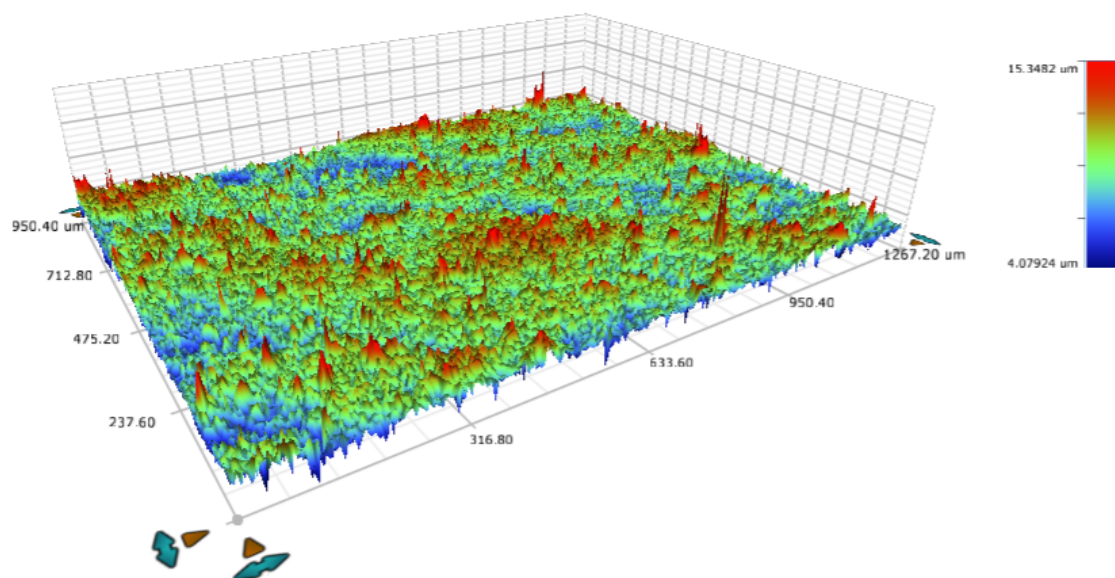
**Figure 79. Profilometry of CMC-12 solution, Run 3 (SWNT:Gly = 0.25, SWNT:Ab = 10) at 100% (no dilution) with an average roughness of 1.048  $\mu\text{m}$ .**



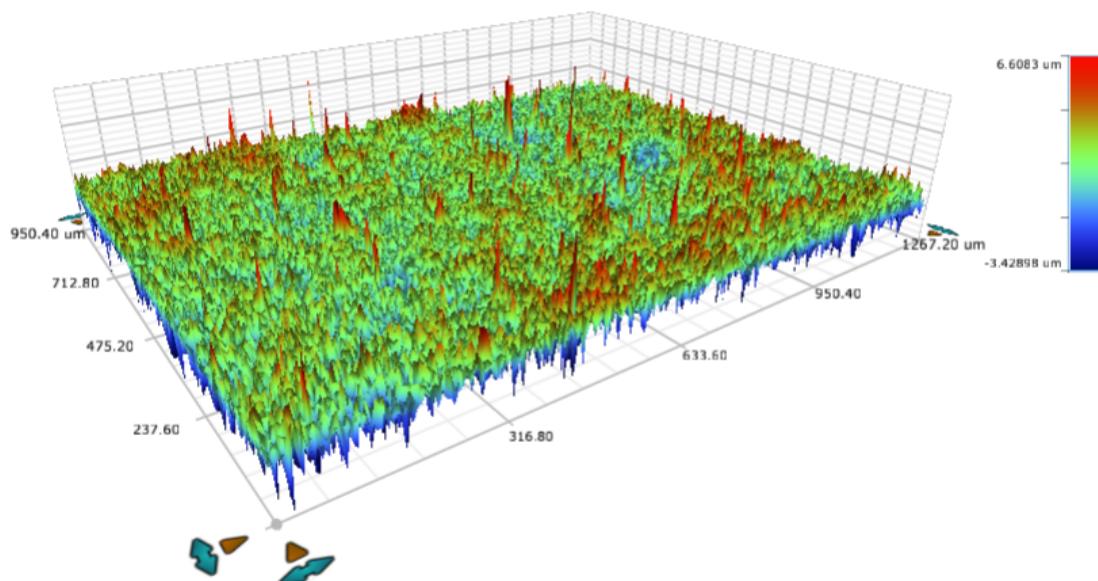
**Figure 80. Profilometry of CMC-12 solution, Run 4 (SWNT:Gly = 0.625, SWNT:Ab = 10) at 100% (no dilution) with an average roughness of 0.816  $\mu\text{m}$ .**



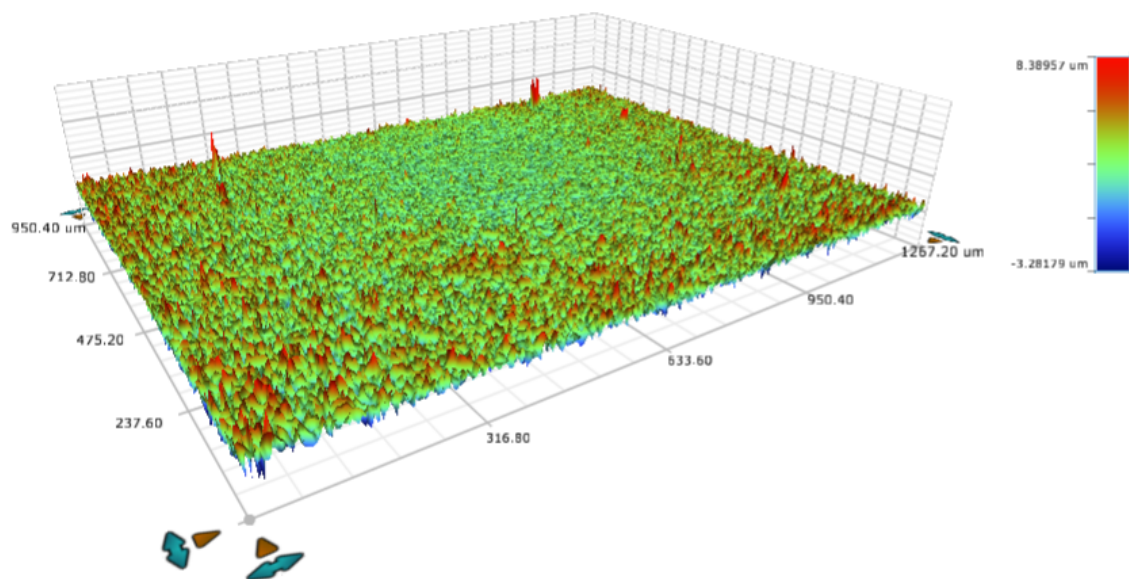
**Figure 81. Profilometry of CMC-12 solution, Run 11 (SWNT:Gly = 1, SWNT:Ab = 10) at 100% (no dilution) with an average roughness of 0.74 μm.**



**Figure 82. Profilometry of CMC-12 solution, Run 9 (SWNT:Gly = 1, SWNT:Ab = 71) at 100% (no dilution) with an average roughness of 0.624 μm.**

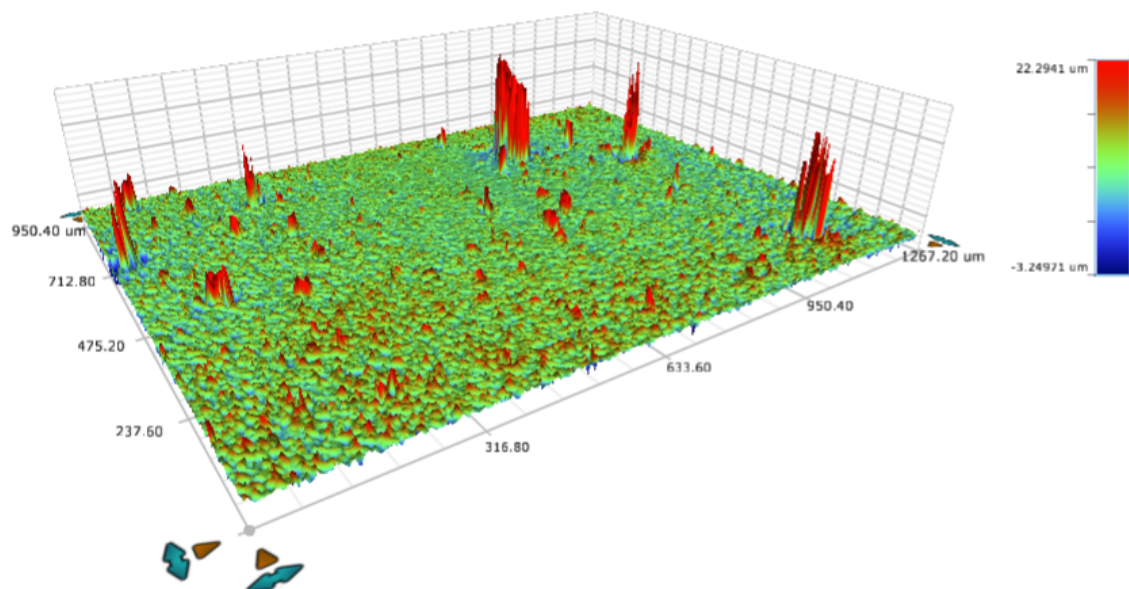


**Figure 83. Profilometry of CMC-12 solution, Run 6 (SWNT:Gly = 1, SWNT:Ab = 133) at 100% (no dilution) with an average roughness of 0.449 μm.**

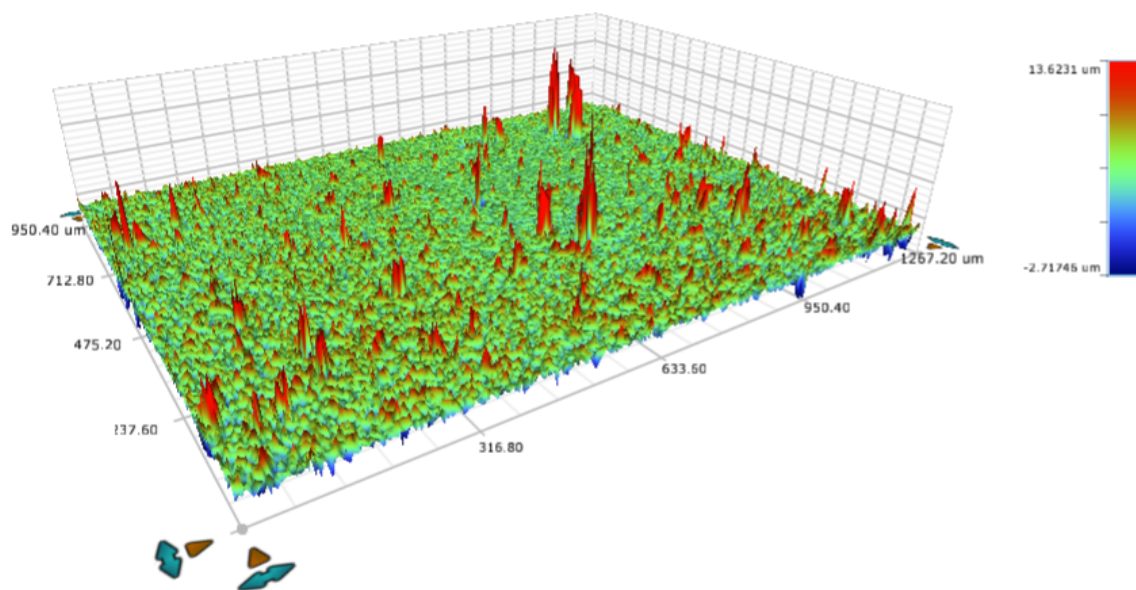


**Figure 84. Profilometry of CMC-14 solution, Run 11 (SWNT:Gly = 1, SWNT:Ab = 10) at 100% (no dilution) with an average roughness of 0.504 μm.**



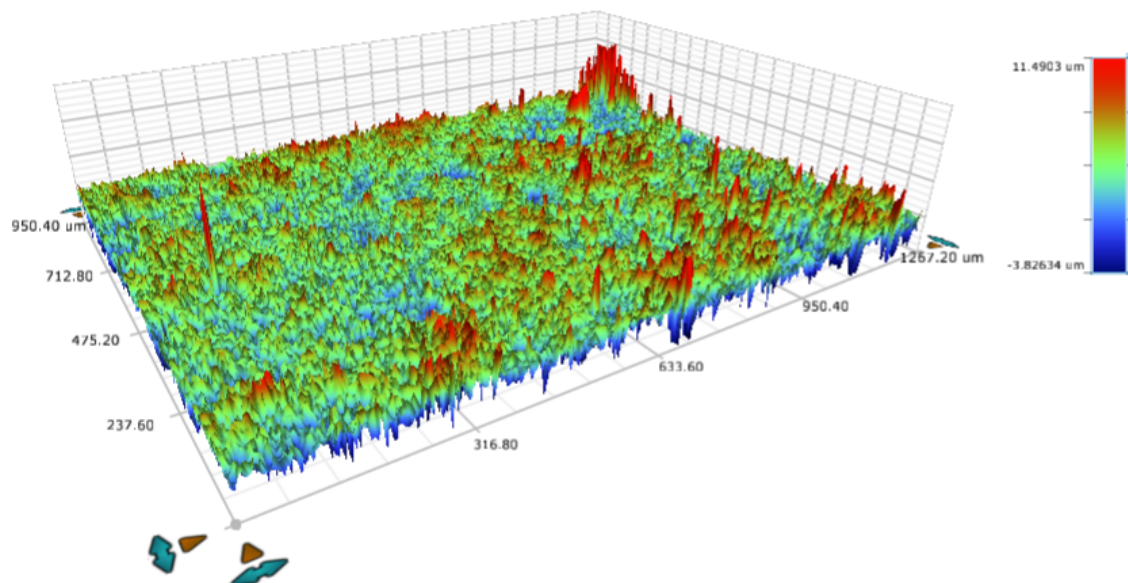


**Figure 85. Profilometry of CMC-14 solution, Run 9 (SWNT:Gly = 1, SWNT:Ab = 71) at 100% (no dilution) with an average roughness of 0.432  $\mu\text{m}$ .**

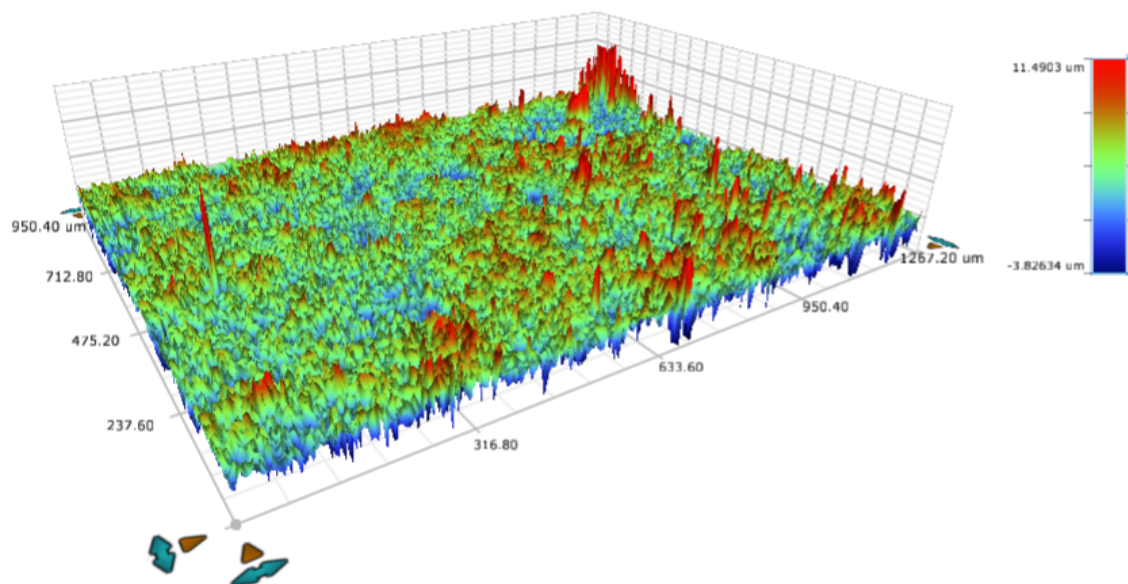


**Figure 86. Profilometry of CMC-14 solution, Run 6 (SWNT:Gly = 1, SWNT:Ab = 133) at 100% (no dilution) with an average roughness of 0.387  $\mu\text{m}$ .**

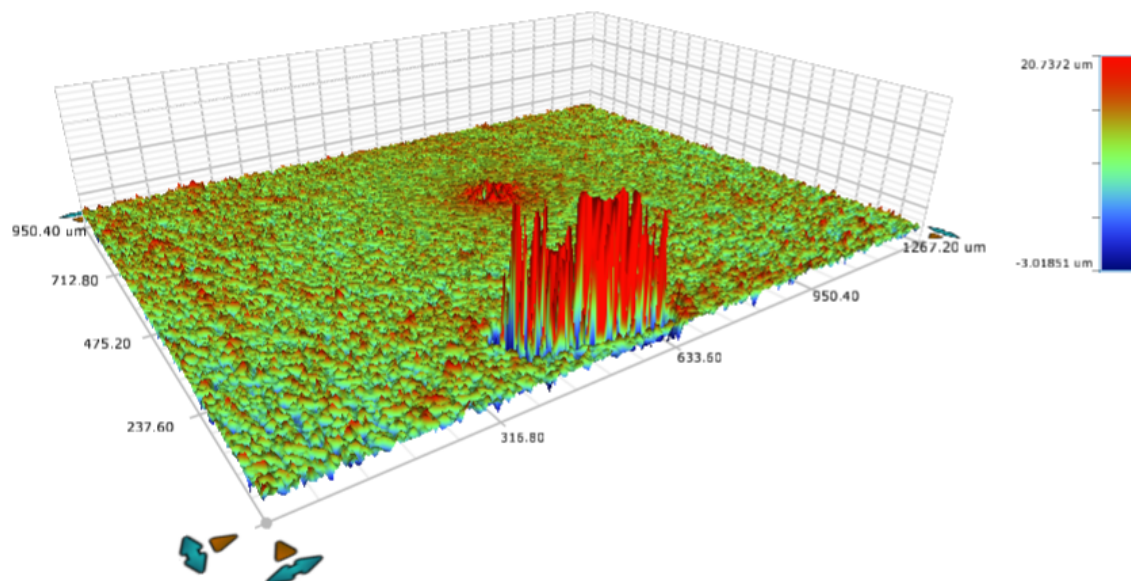




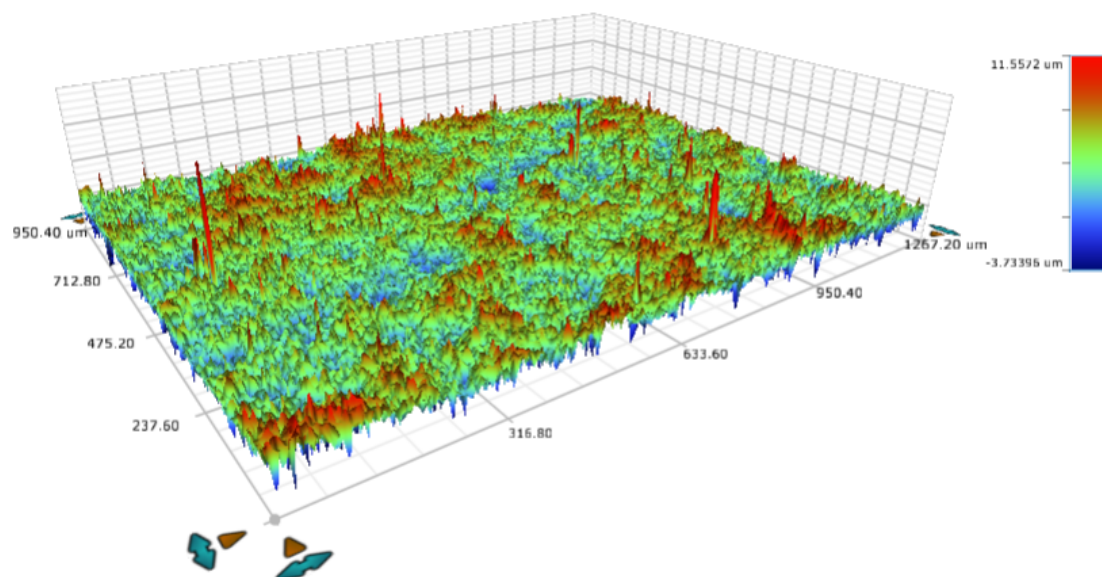
**Figure 87. Profilometry of CMC-17 solution, Run 11 (SWNT:Gly = 1, SWNT:Ab = 10) at 100% (no dilution) with an average roughness of 0.611  $\mu\text{m}$ .**



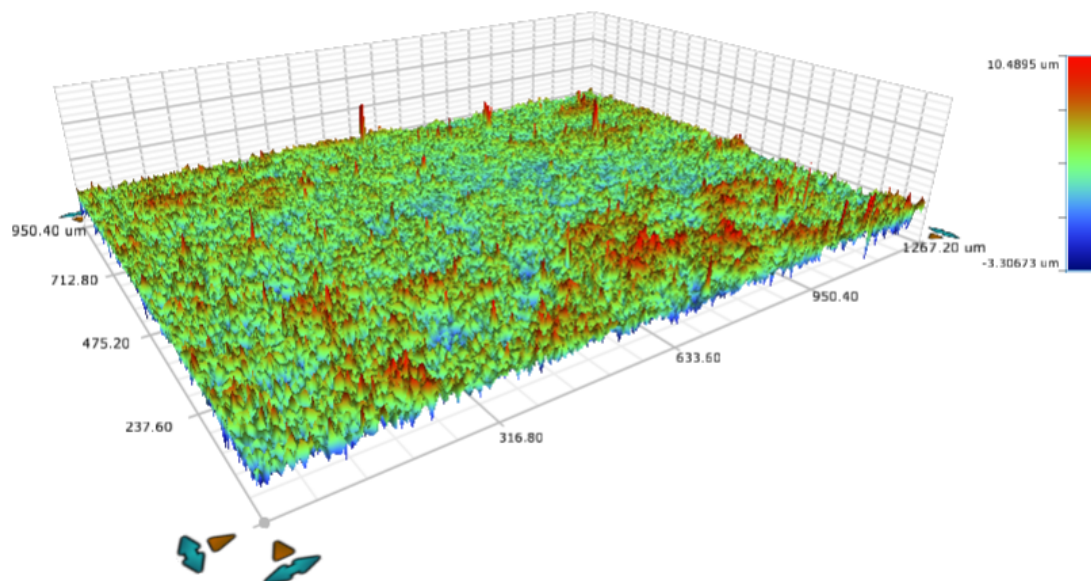
**Figure 88. Profilometry of CMC-17 solution, Run 9 (SWNT:Gly = 1, SWNT:Ab = 71) at 100% (no dilution) with an average roughness of 0.455  $\mu\text{m}$ .**



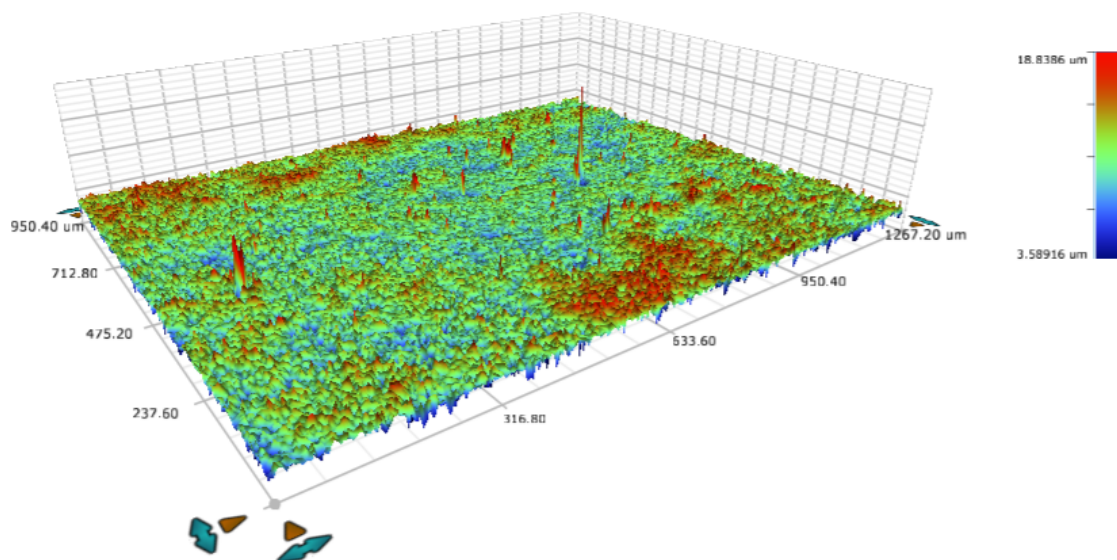
**Figure 89. Profilometry of CMC-17 solution, Run 6 (SWNT:Gly = 1, SWNT:Ab = 133) at 100% (no dilution) with an average roughness of 0.419  $\mu\text{m}$ .**



**Figure 90. Profilometry of CMC-12 solution, Run 3 (SWNT:Gly = 0.25, SWNT:Ab = 10) diluted to 25% with an average roughness of 0.583  $\mu\text{m}$ .**

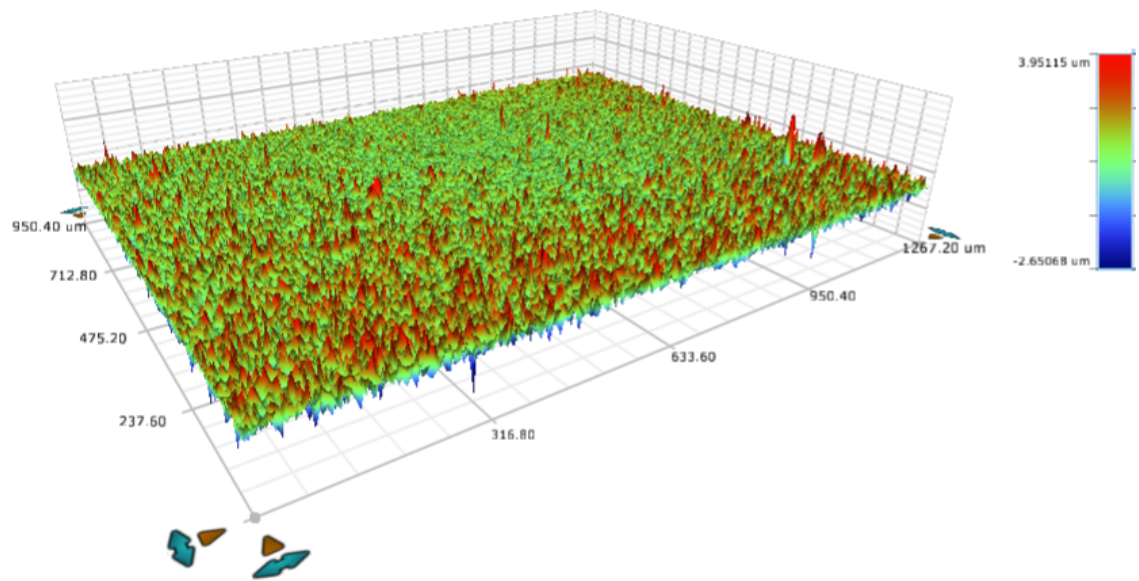


**Figure 91. Profilometry of CMC-12 solution, Run 4 (SWNT:Gly = 0.625, SWNT:Ab = 10) diluted to 25% with an average roughness of 0.424  $\mu\text{m}$ .**

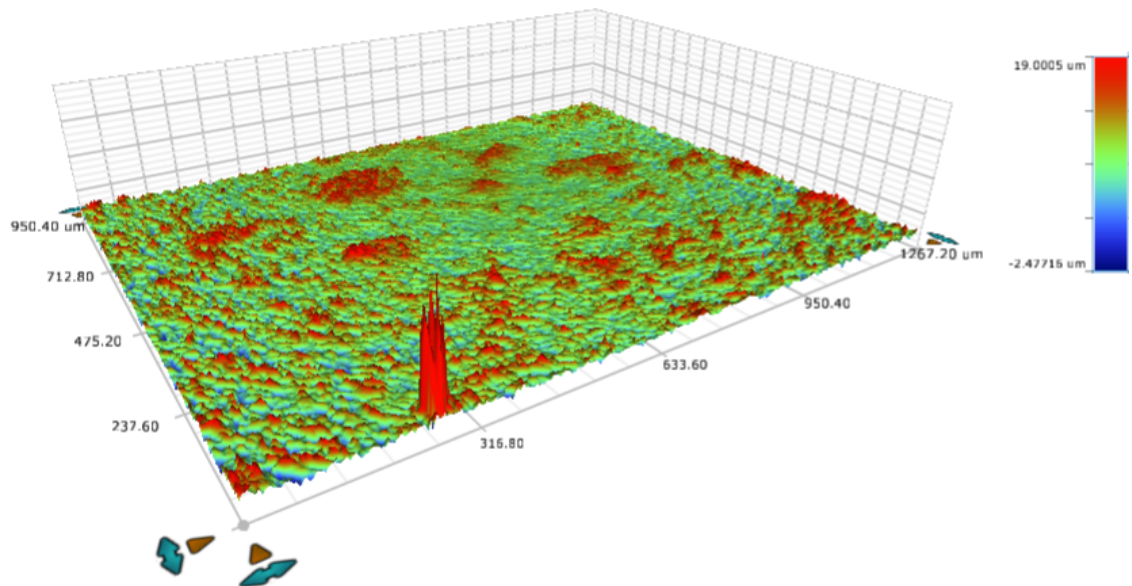


**Figure 92. Profilometry of CMC-12 solution, Run 11 (SWNT:Gly = 1, SWNT:Ab = 10) diluted to 25% with an average roughness of 0.412  $\mu\text{m}$ .**





**Figure 93. Profilometry of CMC-14 solution, Run 11 (SWNT:Gly = 1, SWNT:Ab = 10) diluted to 25% with an average roughness of 0.192  $\mu\text{m}$ .**



**Figure 94. Profilometry of CMC-17 solution, Run 11 (SWNT:Gly = 1, SWNT:Ab = 10) diluted to 25% with an average roughness of 0.285  $\mu\text{m}$ .**

It can be concluded that the surface topography of the dried matrix films is in correlation with the resistance response of the matrix. Uniform and smooth surfaces can be translated to low SWNT agglomeration and low resistance of the dried matrix solutions. Therefore it is desirable to obtain a uniform and smooth coating.

### 3.3.4 Coating thickness

The thickness of the coatings is important because of their effect on the resistance of the obtained dried films. The higher the coating thickness is, the less the resistance of the coating will be. The thickness of the dried matrix coating was investigated using the optical profiler. For this purpose, the optical profile of the edge of the coatings was obtained using optical profiler for select samples. The thickness of matrix coatings from profilometry obtained for CMC-12, CMC-14, and CMC-17 dried matrix formulations at 100% (no dilution), and 25% dilution is presented in Figure 95 through Figure 110, and summarized in Table 38.

The thickness of the coatings was observed to decrease with an increase in the glycerol concentration. This trend suggests that the mass of material applied at higher glycerol concentrations may have been lower due to precipitates being filtered out during the matrix preparation (in phase 1 of this research it was shown that the measured thickness was consistent with mass balance calculated thickness). This would also be consistent with the increasing surface roughness trend that was observed with increasing glycerol, suggesting an increase in particle size.

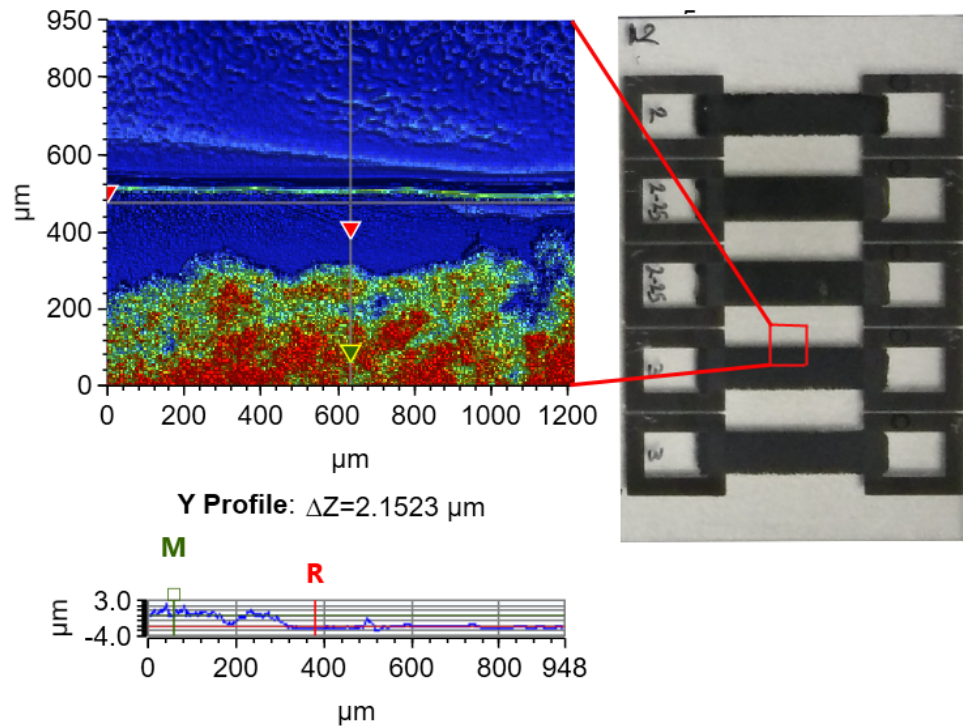
The contribution of the concentration of antibody to the thickness is fairly small, showing a slight increase with antibody concentration for CMC-12 and slight decrease with concentration for CMC-17 and CMC-14, and perhaps statistically insignificant.

As expected, the thickness of the coatings is observed to decrease by slightly more than fourfold with the fourfold dilution of the matrix solution from 100% to 25%.

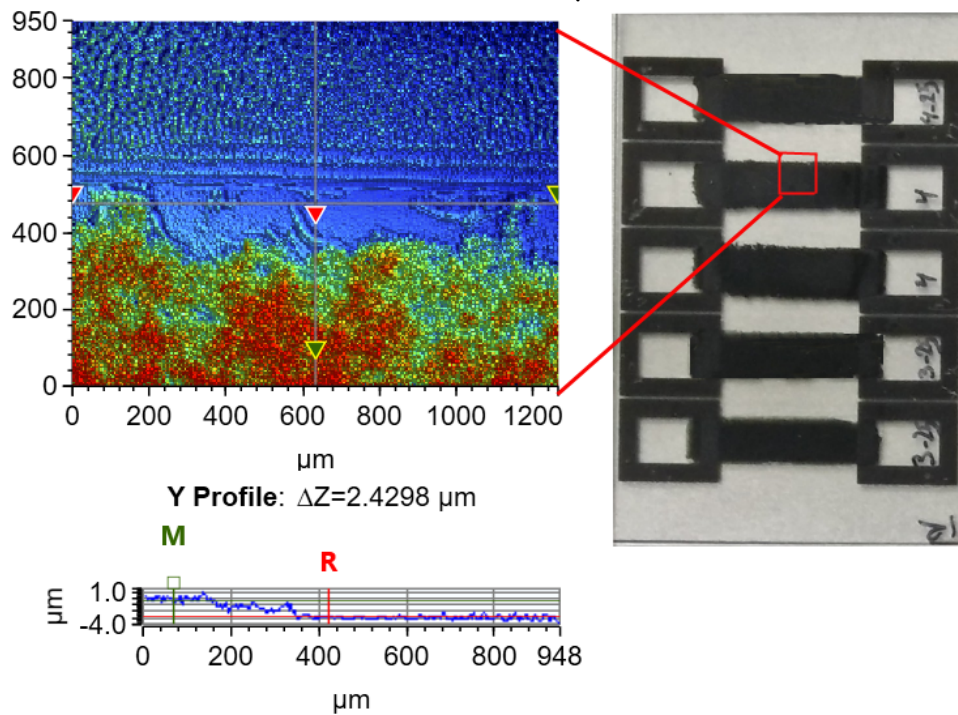
Resistivity, in contrast, did not increase with dilution as much as expected (4x or higher) and at highest glycerol ratios actually decreased slightly.

**Table 38- Comparison of the coatings' average thickness and its effect on resistance for select coatings**

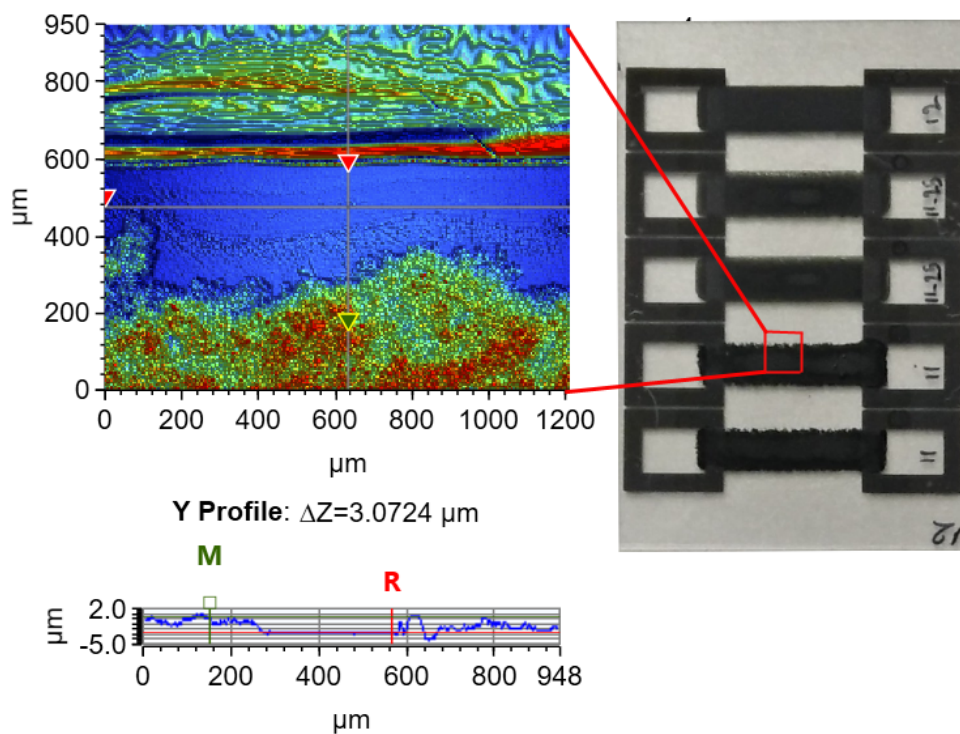
Run	SWNT: GLY	SWNT: Ab	dilution	Resistance ( $\Omega$ )			Thickness ( $\mu\text{m}$ )			Theoretical Thickness ( $\mu\text{m}$ )		
				CMC-12	CMC-17	CMC-14	CMC-12	CMC-17	CMC-14	CMC-12	CMC-17	CMC-14
3	0.25	10	100%	180753	968476	1650440	2.15			2.30		
4	0.625	10	100%	5283	46313	138495	2.43			2.59		
11	1	10	100%	1841	8939	13822	3.07	1.85	1.24	2.78	4.45	3.59
	$\infty$	$\infty$	100%	148	147	1069				3.56	4.19	4.6
11	1	10	100%	1841	8939	13822	3.07	1.85	1.24	2.78		
9	1	71	100%	1265	5570	5727	2.84	1.92	1.31	2.78		
6	1	133	100%	474	4162	4337	2.79	2	1.46	2.78	4.45	3.59
	$\infty$	$\infty$	100%	148	147	1069				3.56	4.19	4.6
3	0.25	10	25%	184197	1376718	356148	0.62			0.57		
4	0.625	10	25%	6753	175400	60731	0.63			0.65		
11	1	10	25%	4692	21869	9983				0.69	1.11	0.90
SWNT:CMC ratio =				2	1.25	0.5	2	1.25	0.5	2	1.25	0.5



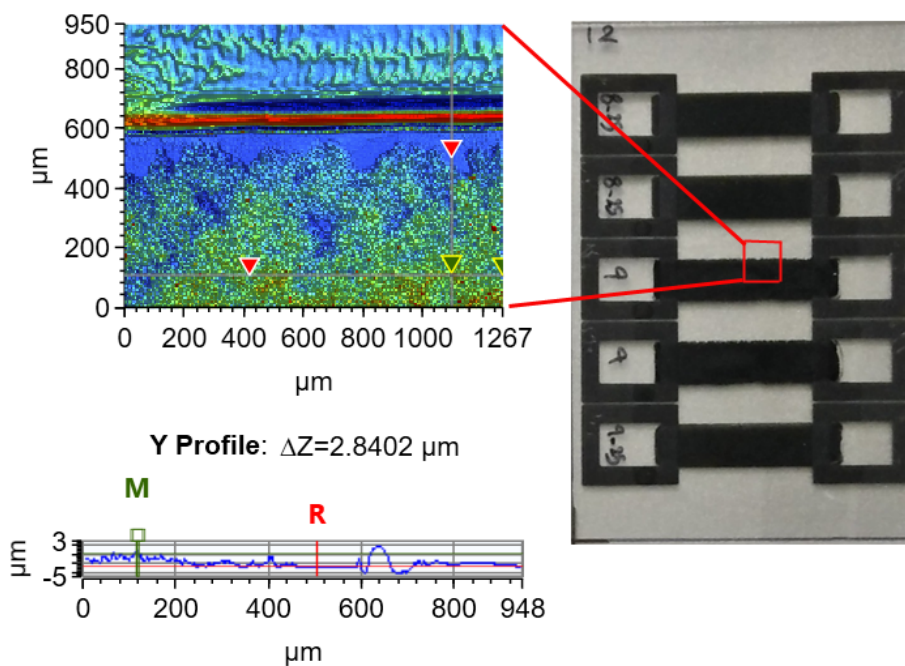
**Figure 95.** The thickness of Run 3 coating ((SWNT:Gly = 0.25, SWNT:Ab = 10) obtained from CMC-12 solution at 100% (no dilution) using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $2.15 \mu\text{m}$ .



**Figure 96.** The thickness of Run 4 coating ((SWNT:Gly = 0.25, SWNT:Ab = 10) obtained from CMC-12 solution at 100% (no dilution) using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $2.43 \mu\text{m}$ .

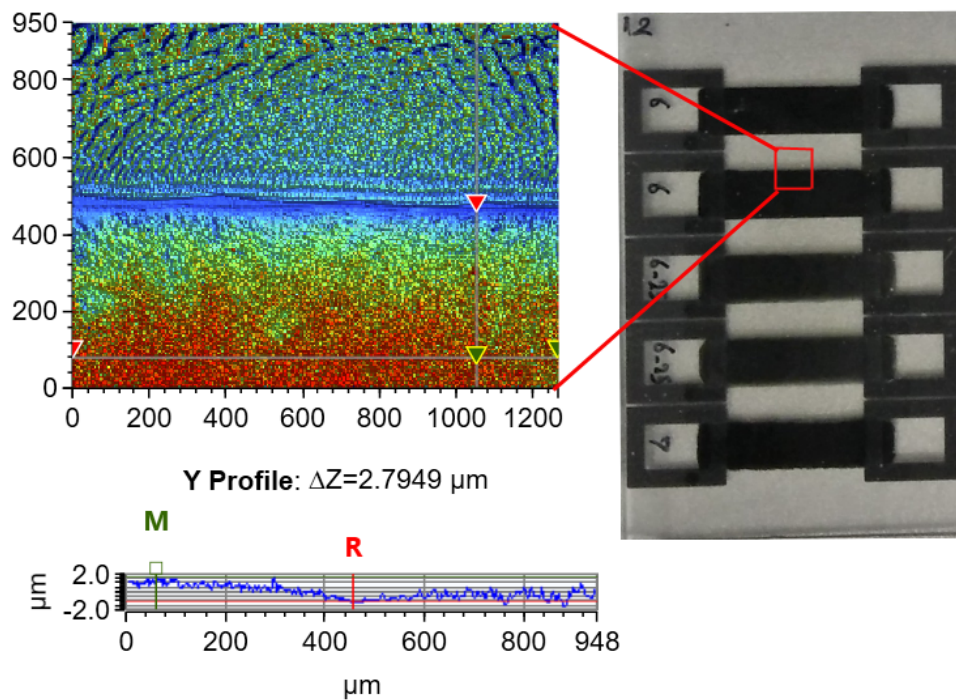


**Figure 97.** The thickness of Run 11 coating ((SWNT:Gly = 1, SWNT:Ab = 10) obtained from CMC-12 solution at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 3.07  $\mu\text{m}$ .

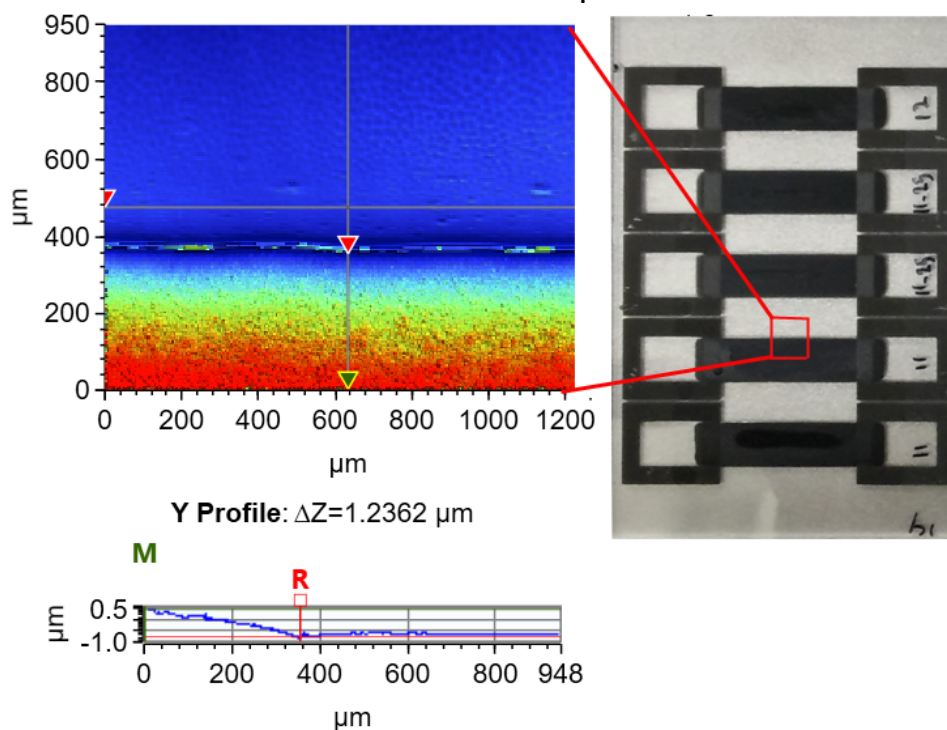


**Figure 98.** The thickness of Run 9 coating ((SWNT:Gly = 1, SWNT:Ab = 71) obtained from CMC-12 solution at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 2.84  $\mu\text{m}$ .

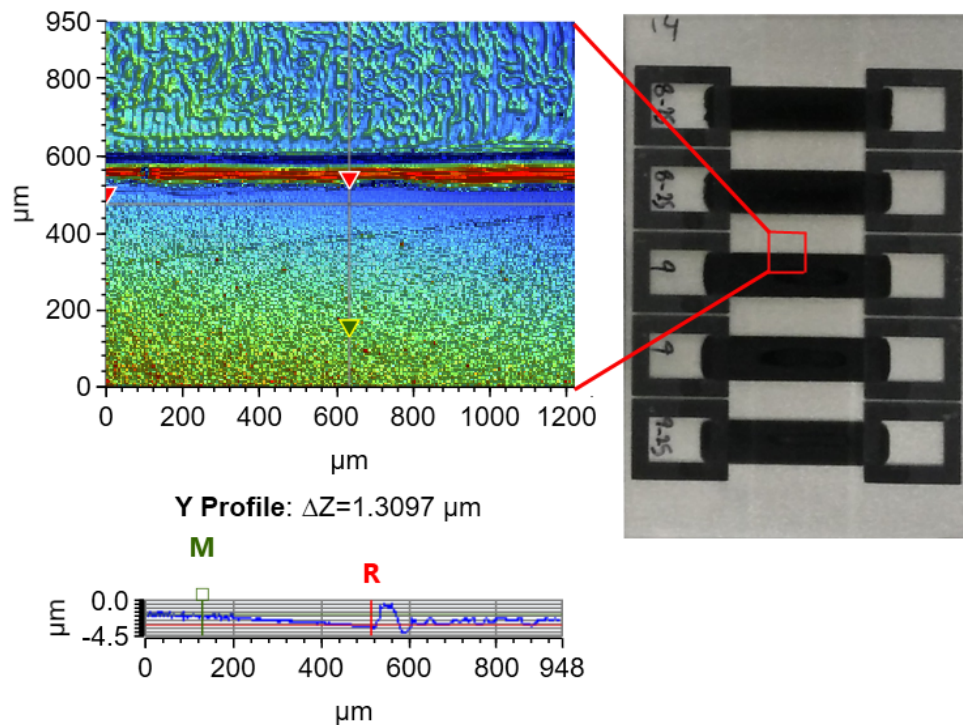




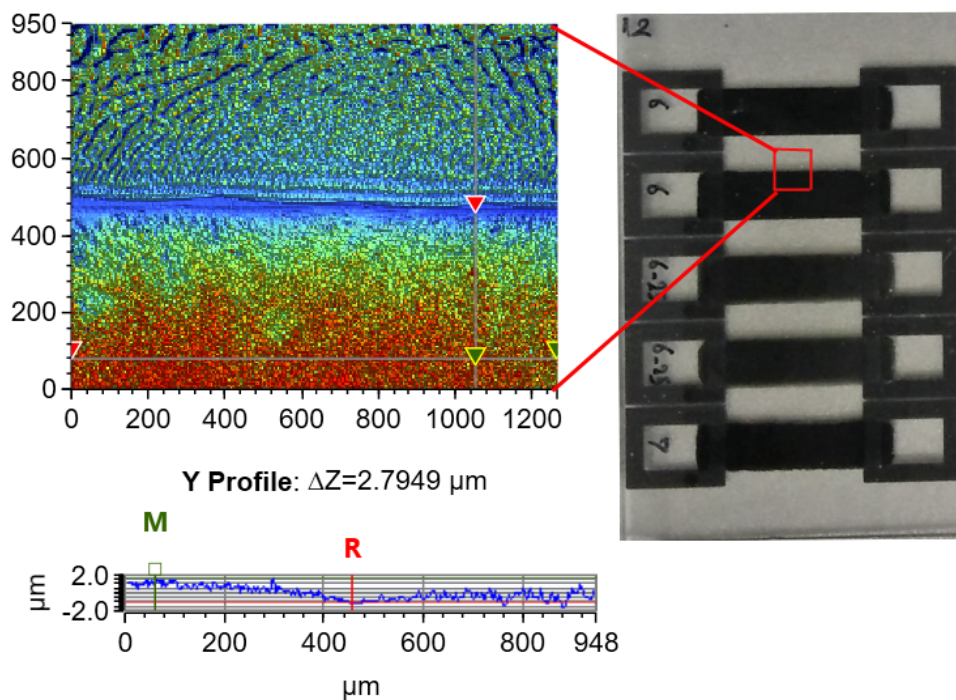
**Figure 99.** The thickness of Run 6 coating ((SWNT:Gly = 1, SWNT:Ab = 133) obtained from CMC-12 solution at 100% (no dilution) using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $2.79 \mu\text{m}$ .



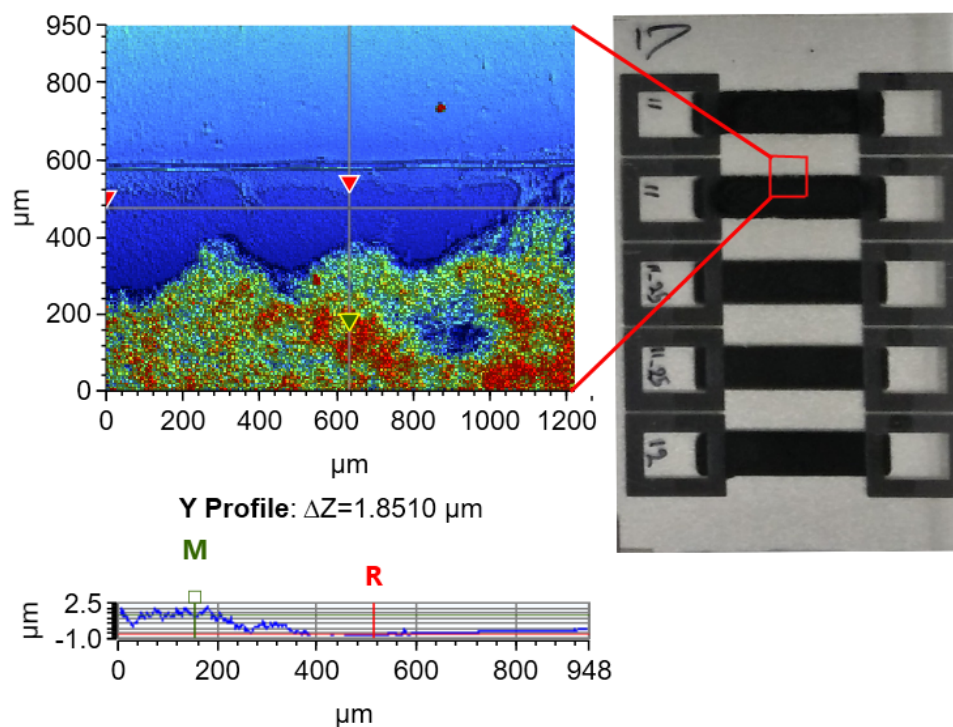
**Figure 100.** The thickness of Run 11 coating ((SWNT:Gly = 1, SWNT:Ab = 10) obtained from CMC-14 solution at 100% (no dilution) using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $1.24 \mu\text{m}$ .



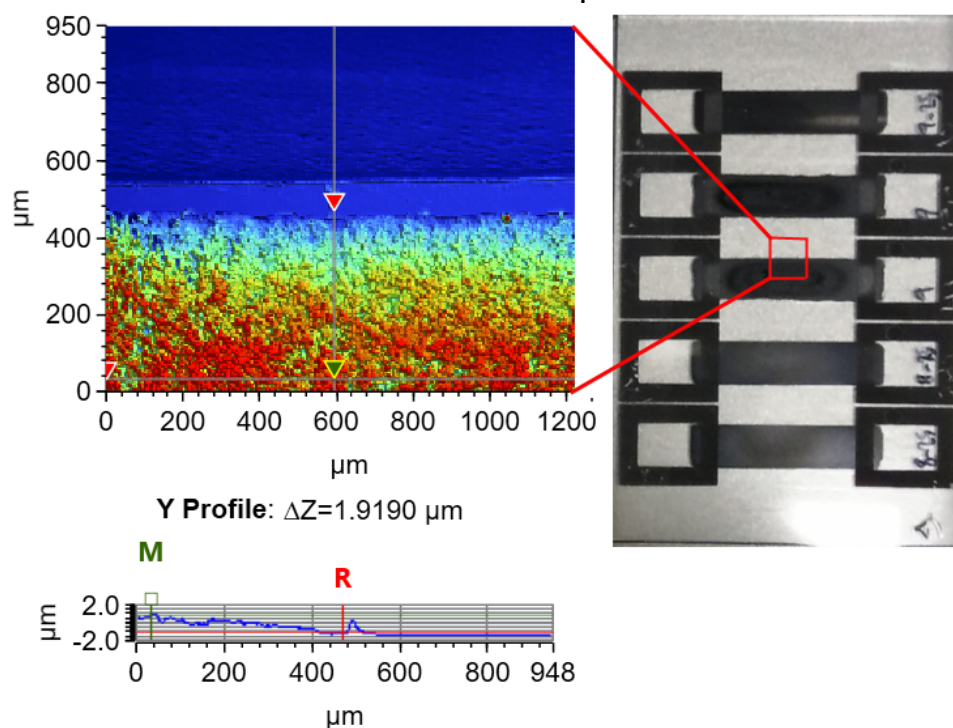
**Figure 101.** The thickness of Run 9 coating ((SWNT:Gly = 1, SWNT:Ab = 71) obtained from CMC-14 solution at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 1.31  $\mu\text{m}$ .



**Figure 102.** The thickness of Run 6 coating ((SWNT:Gly = 1, SWNT:Ab = 133) obtained from CMC-14 solution at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 2.79  $\mu\text{m}$ .

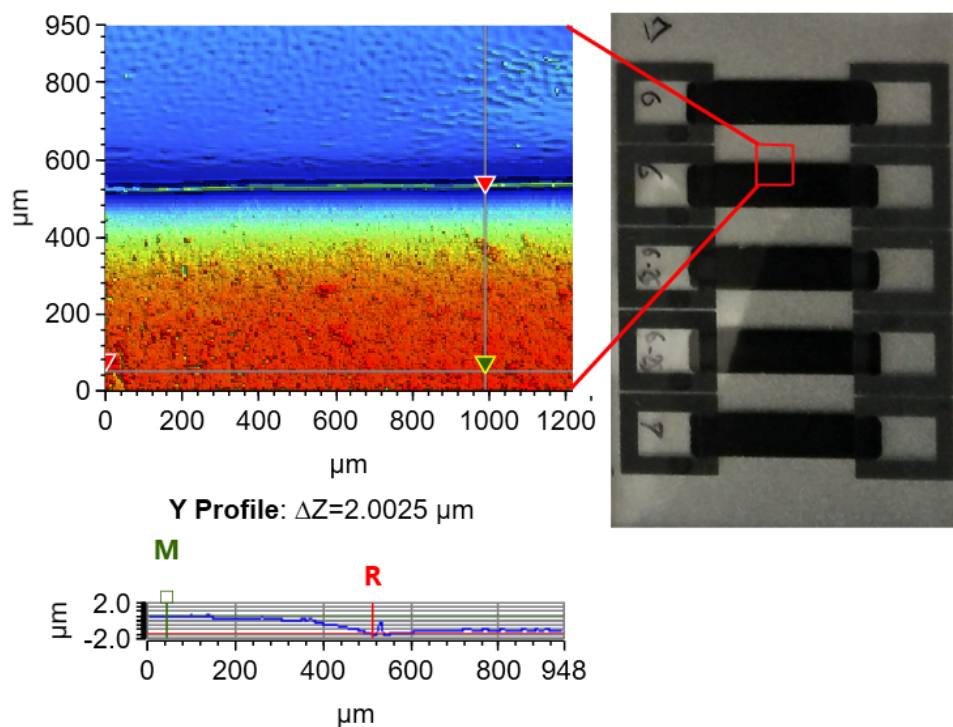


**Figure 103.** The thickness of Run 11 coating ((SWNT:Gly = 1, SWNT:Ab = 10) obtained from CMC-17 solution at 100% (no dilution) using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $1.85 \mu\text{m}$ .

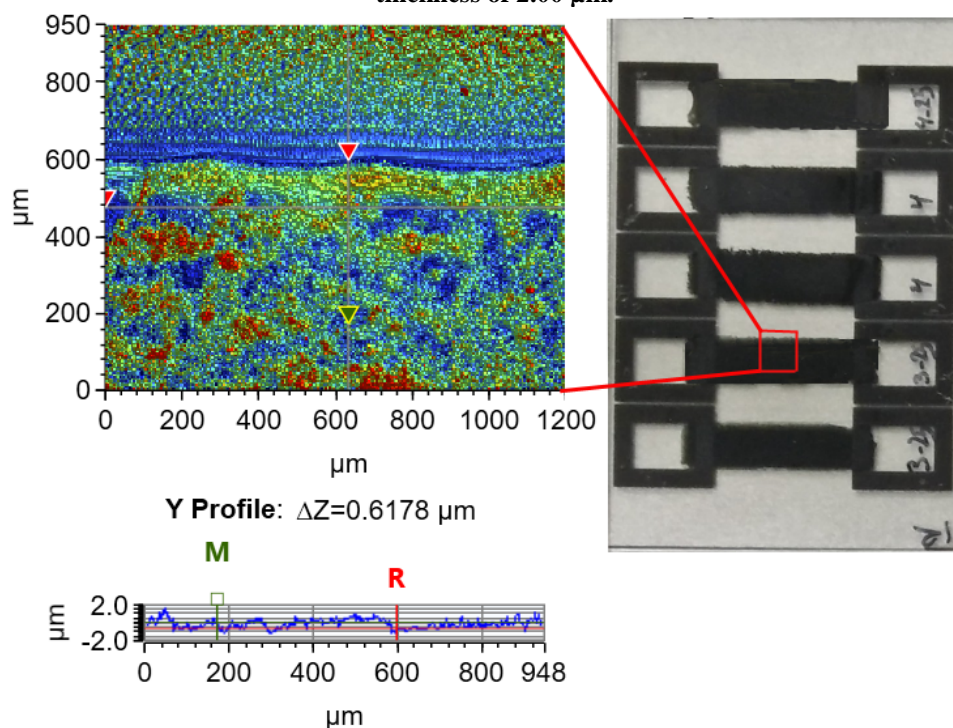


**Figure 104.** The thickness of Run 9 coating ((SWNT:Gly = 1, SWNT:Ab = 71) obtained from CMC-17 solution at 100% (no dilution) using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $1.92 \mu\text{m}$ .

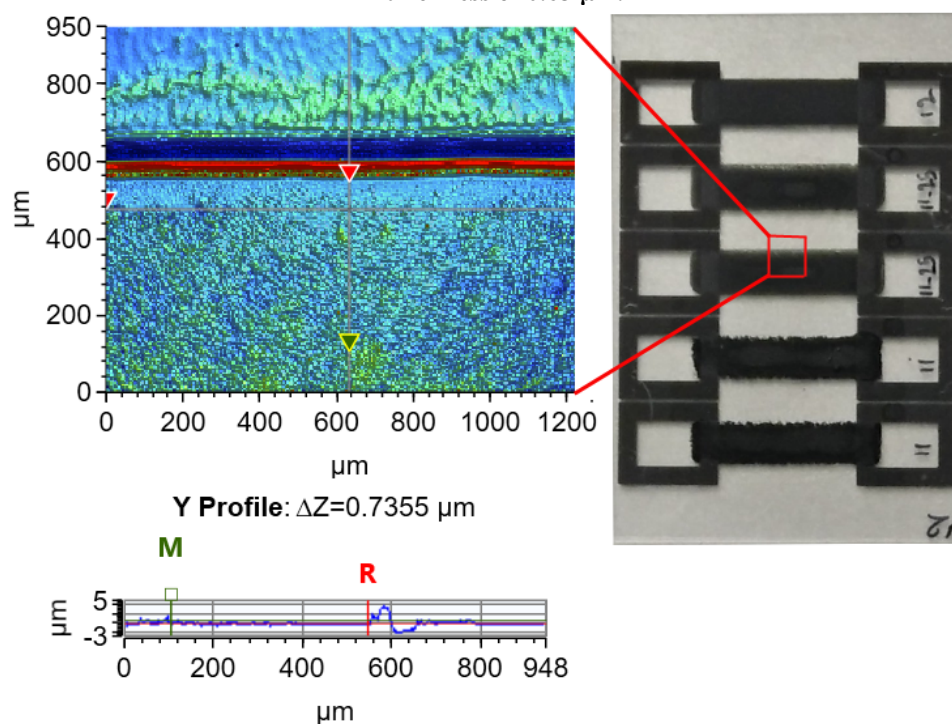
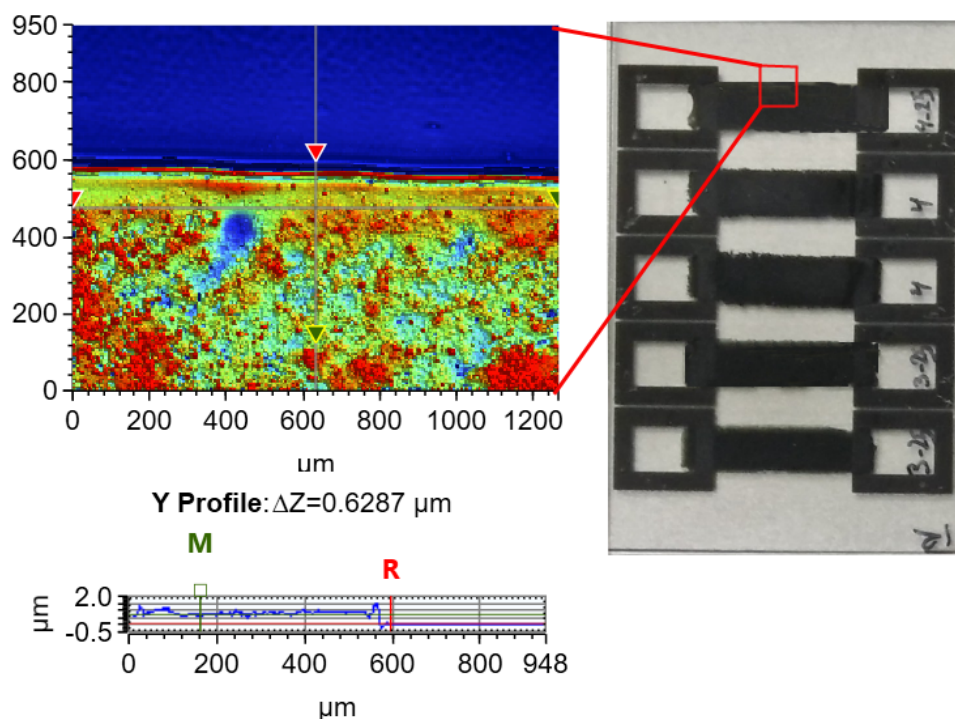


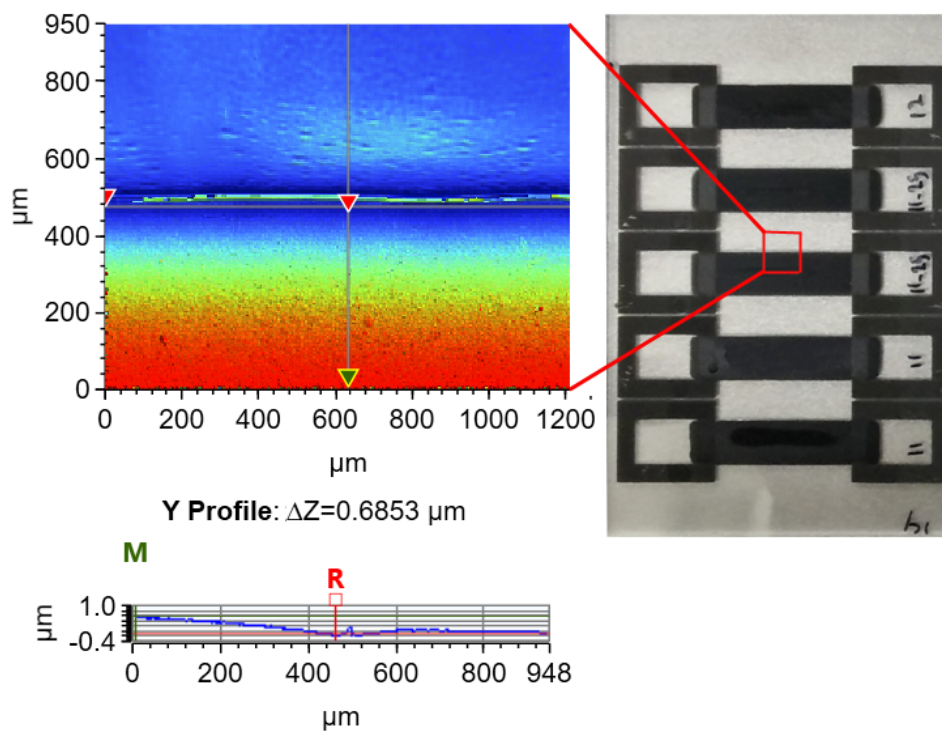


**Figure 105.** The thickness of Run 6 coating ((SWNT:Gly = 1, SWNT:Ab = 133) obtained from CMC-17 solution at 100% (no dilution) using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 2.00  $\mu\text{m}$ .

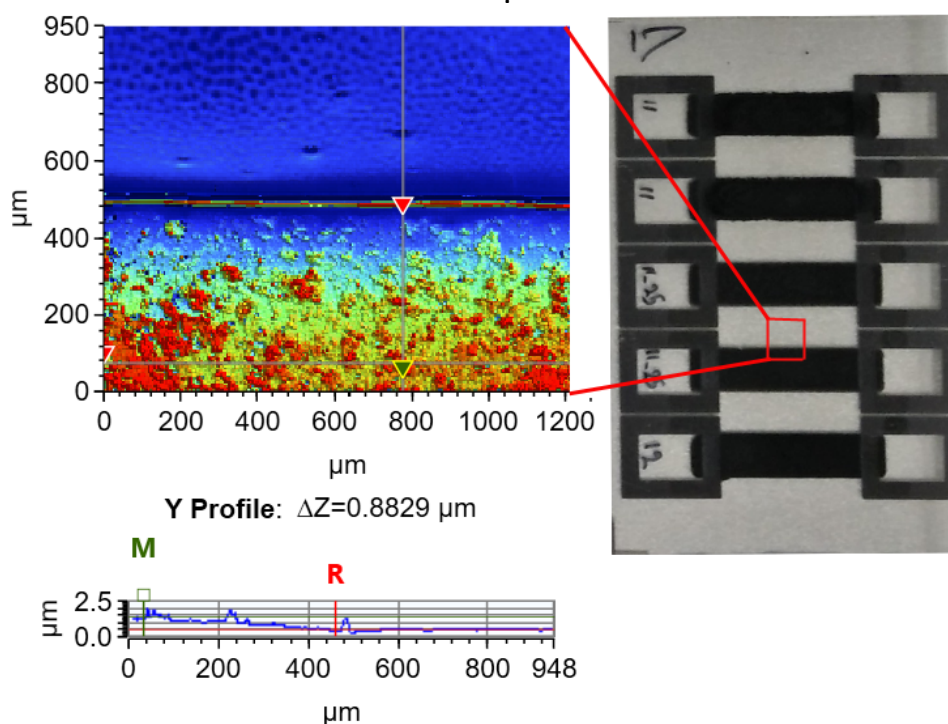


**Figure 106.** The thickness of Run 3 coating ((SWNT:Gly = 0.25, SWNT:Ab = 10) obtained from CMC-12 solution diluted to 25% using 2D microscopy profile (1250  $\mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of 0.62  $\mu\text{m}$ .





**Figure 109.** The thickness of Run 11 coating ((SWNT:Gly = 1, SWNT:Ab = 10) obtained from CMC-14 solution diluted to 25% using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $0.68 \mu\text{m}$ .



**Figure 110.** The thickness of Run 11 coating ((SWNT:Gly = 1, SWNT:Ab = 10) obtained from CMC-17 solution diluted to 25% using 2D microscopy profile ( $1250 \mu\text{m} \times 950 \mu\text{m}$ ) showing a thickness of  $0.88 \mu\text{m}$ .

### 3.3.5 RSM optimization

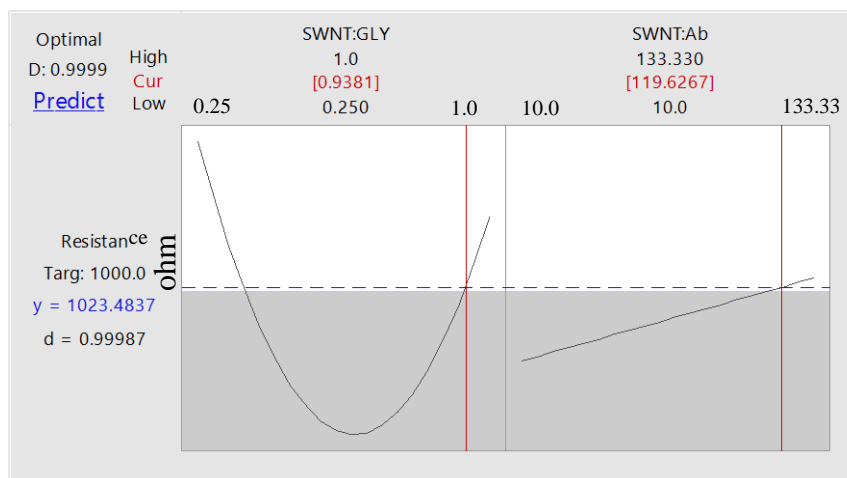
Thus far, as analyzed in the result and discussion section of this chapter, the resistance behavior of coatings prepared from CMC-12 were found to be superior when compared to that of the coatings prepared from solutions CMC-14 and CMC-17. The workability and the uniformity of the coatings prepared from CMC-12 matrix solutions were also compared with other matrix coatings and were found to be superior, as the coatings exhibit the highest uniformity among different matrix solutions. These facts lead the author to select the CMC-12 matrix solution as a potential candidate for biosensing applications. Every 5 ml of the CMC-12 matrix solution comprise of 0.0134 g of SWCNT, 0.1674 g of CMC, 0.0321 g of azide, and 0.0045 grams of sodium phosphate dibasic dihydrate to adjust the pH to between 7 and 8. Among different ratio variations of SWNT:Glycerol, the highest ratio tested, 1:1, was found to lead to the minimum electrical resistance. Although the addition of antibody in the system results in an increase in the resistance of the matrix coating, any desired concentration of antibody, up to the SWNT:antibody ratio of 10, can be selected while maintaining the resistance below 2000  $\Omega$ , using matrix at full strength (no dilution with water). Therefore, it can be concluded that the matrix coating obtained using the CMC dispersant at the ratio of SWNT:CMC of 2.0 and the sonication amplitude of 40% for 2 hours can be potentially used for integration into immunodiagnostic biosensors. Glycerol at the ratio of SWNT:glycerol of 1.0 can be used as a humectant to help preserve the activity of the incorporated antibody (not yet evaluated).

Using the response surface methodology (RSM) design of experiment in the second phase of this study with antibody and glycerol, the behavior of the coatings were modeled.

The comparison between the experimental resistance values and the predicted values in section 3.3.2, Table 36, shows that the regression models can be used to predict the resistance behavior of the matrix coatings. 63% of the predicted resistances in the Table 36 fall within  $\pm 20\%$  of the experimental values. The worst predicted value differs by 68% from the experimental value, however considering the range of resistances covered by this model in Table 36 was from  $474\Omega$  to  $1.65\text{M}\Omega$ , the model is fairly good.

As an example, the RSM models generated in this chapter can be used to identify formulation conditions that will result in dried matrix resistances of  $\sim 1000\ \Omega$ . The optimized parameters for one possible solution of achieving this goal using CMC-12 at 100% dilution is shown in the table insert in Figure 111. This figure also shows the overlay of the optimum conditions for Solution 1 at a SWNT:glycerol ratio of 1 and SWNT:antibody ratio of 120.





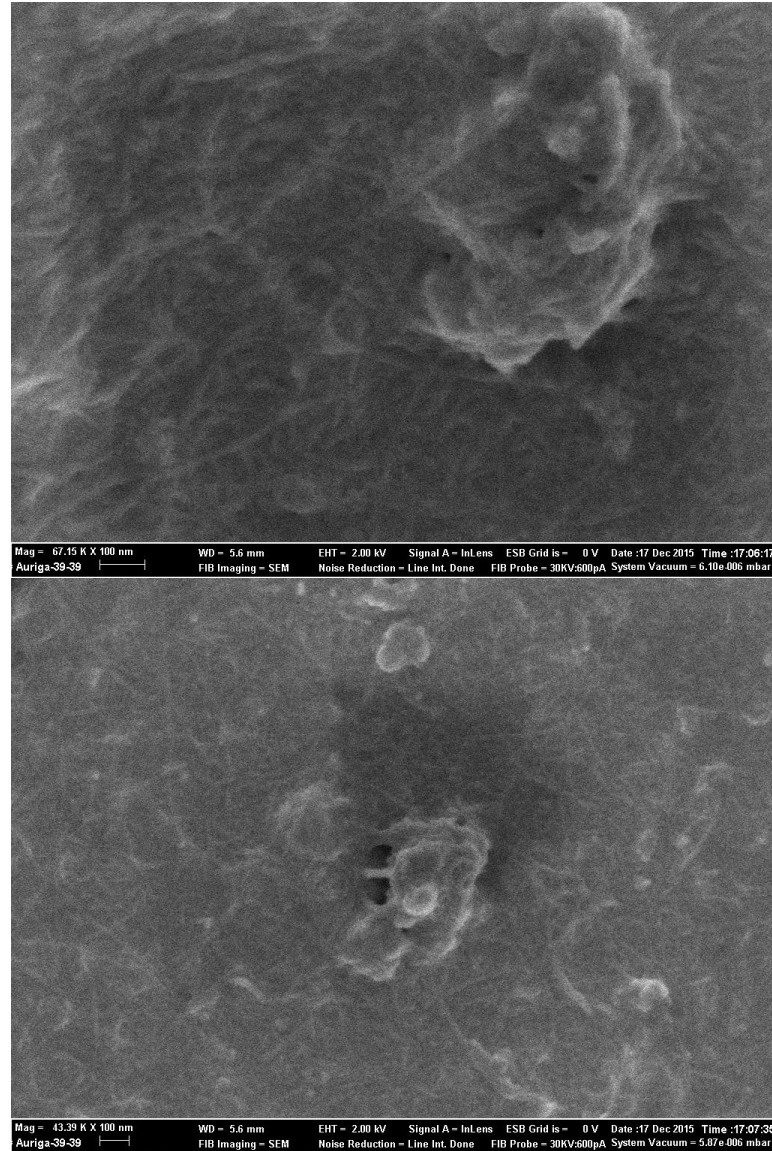
Solution	SWNT:Gly	SWNT:Ab	Resistance (ohm) Fit	Composite Desirability
1	1	120	1023	1

**Figure 111. The main effect graph showing the optimized condition for achieving ~1000 ohm resistance using the CMC-12 solution at 100% (no dilution). The table insert shows the condition optimized for achieving a resistance of 1000  $\Omega$ , based on the response surface design.**

### 3.3.6 Scanning electron microscopy

Efforts were made to obtain SEM micrographs of the matrix coatings. Five Institutions and Universities (Michigan State University, Western Michigan University (Dr. Eversole and Dr. Arigur's labs), Georgia Institute of Technology, and University of Georgia) have made effort to visualize a number of the samples of the prepared coating films. However, technical difficulties prohibited a successful visualization of the samples. Among the mentioned institutes, MSU obtained the best micrographs. Unfortunately, the SEM device used at MSU was not able to employ a higher magnification and sharper image due to a technical problem. Figure 112 shows SEM micrographs of Run 6 coating ((SWNT:Gly = 1, SWNT:Ab = 133) obtained from CMC-12 solution at 100% (no dilution). Uniformly dispersed individual SWNTs are visible in this figure. A larger clump of SWNT, representing SWNT agglomeration is visible in these two images, showing the

nature of SWNT bundles. The antibody and CMC molecules are not visible in these images due to insufficient magnification.



**Figure 112. SEM micrographs of Run 6 coating ((SWNT:Gly = 1, SWNT:Ab = 133) obtained from CMC-12 solution at 100% (no dilution) at (top) 67KX and (bottom) 43KX. Images show fairly uniformly dispersed individual SWNTs and a larger agglomerated clump of SWNT.**

### 3.4 Conclusion

The base matrix solution without glycerol and antibody was selected from the experiments the 1<sup>st</sup> phase of this research based on a set of matrix film formulations that achieved low resistance for each SWNT to CMC ratio. Matrix formulation protocols from

Run 14 (SWNT:CMC = 0.5), Run 17 (SWNT:CMC = 1.25), and Run 12 (SWNT:CMC = 2) were selected and labeled as CMC-14, CMC-17, and CMC-12, respectively. The corresponding resistance results for the dried matrices were 1069, 794, and 148  $\Omega$ , respectively. Seventy-eight different solutions were prepared using the CMC-14, CMC-17, and CMC-12 base matrix formulations based on an experimental design containing various SWNT:glycerol and SWNT:antibody ratios. Their resistance responses as well as surface morphologies were investigated using Profilometry. Although scanning electron microscopy of the prepared films with antibody/glycerol, the low quality of the SEM images at three different Universities (apparently due to technical difficulties as explained by the SEM technicians at each University) did not allow a comprehensive study of the coated matrix film morphology. However, the uniformly coated individualized SWNTs are still detectable in the lower magnification images obtained at Michigan State University.

The resistance behavior of coatings prepared from CMC-12 solution were found to be statistically more predictable when compared to that of the coatings prepared from solutions CMC-14 and CMC-17. 3D optical microscopy was performed on the dried matrix films to investigate the thickness of the coatings as well as to study the morphology of the dried films. The thickness of the coatings was found to increase with a decrease in the glycerol concentration. The uniformity of the dried films of antibody/glycerol formulations prepared from CMC-12 matrix solutions were found to be the best among the different matrix solutions and this superior uniformity helped achieve lower resistance. Therefore, CMC-12 matrix solution may be a potential candidate for biosensing applications.

The resistance response of different matrix dried film formulations were modeled using the RSM design. Among the three different ratio variations of SWNT:Glycerol, the

highest 1:1 ratio was found to lead to the lowest electrical resistance. Although the addition of antibody to the matrix results in an increase in the resistance of the matrix coating, increased levels of antibody up to the SWNT:antibody ratio of 10 can be added while maintaining the resistance below 2000  $\Omega$  if the formulation is not diluted. Therefore, it was determined that the matrix coating obtained using the CMC dispersant at the ratio of SWNT:CMC=2 and the sonication amplitude of 40% for 2 hours will lead to a relative low resistance, and uniform dried matrix film compared to others that were studied. Glycerol at the ratio of SWNT:glycerol of 1:1 is included in this matrix formulation and potentially could serve as a stabilizer to preserve the activity of antibodies after drying (not studied in this thesis). It is hoped that information from this research will be helpful in the design of immunodiagnostic biosensors relying on antibody-SWNT design elements.

# FUTURE STUDY

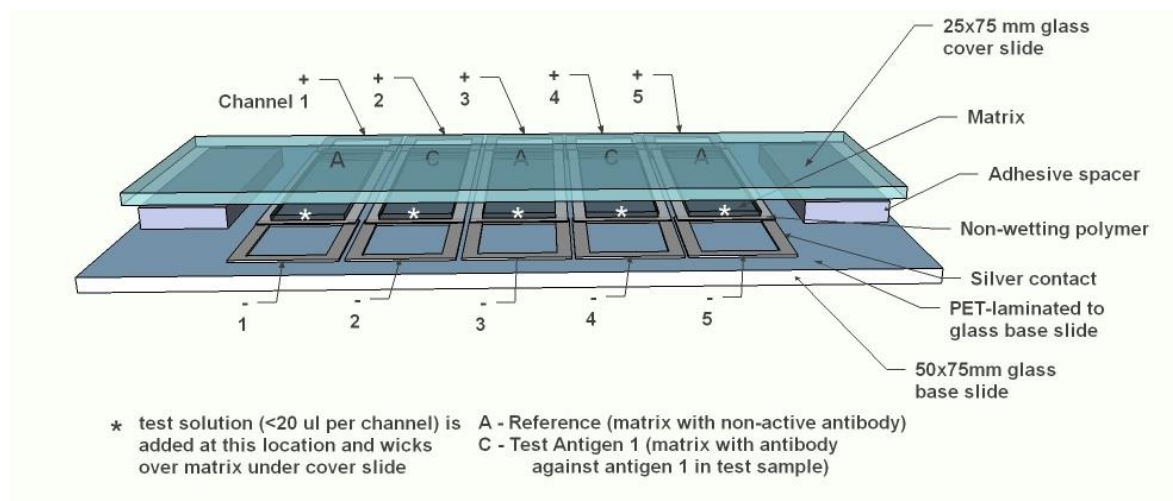
## 4.1 Objective

A long-term research laboratory goal in Dr. Young's laboratory is to formulate a stable, sensitive and fast biosensor for immunodiagnosics application. Once developed, application of printing technology would be further pursued to streamline the matrix deposition process and avoid high expenses associated with traditional sensor manufacturing processing. A disposable biosensor suitable for point-of-care application is envisioned. With low fabrication cost and capability of miniaturization, this biosensor may find potential applications in both toxin detection and immunodiagnosics. Future studies beyond the scope of this dissertation would be required to develop multipurpose functional biosensors with reliable responses. Nevertheless, future work might be done using the results from the prior sections in this thesis to explore matrix resistance versus time of antigen solution exposure curves at different antigen solution concentrations to determine if specific antibody–antigen binding can be detected.

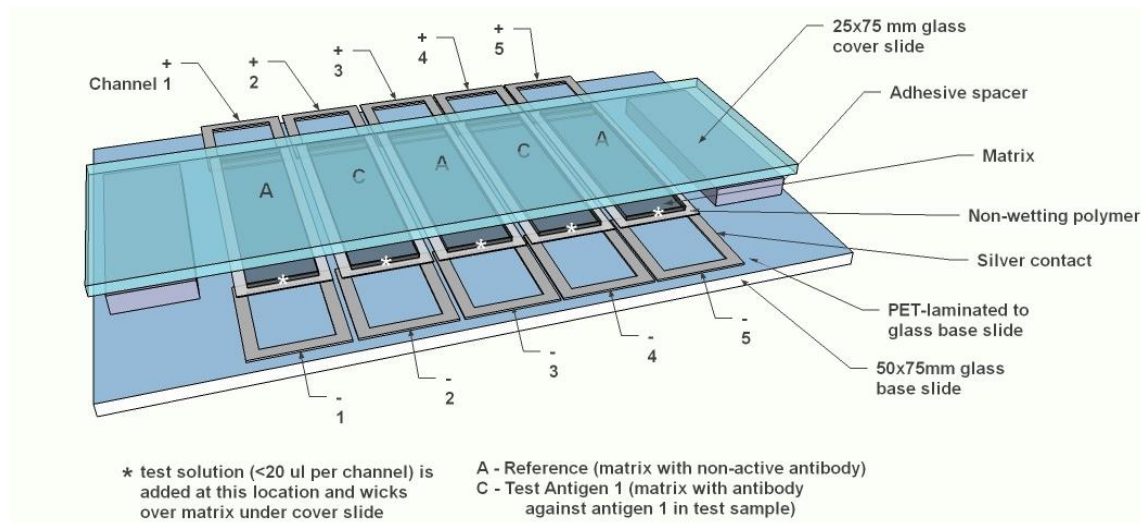
### 4.1.1 Experimental

The schematic picture of a fabricated laboratory test biosensor plate is depicted in Figure 113, Figure 114 and Figure 115. The biosensor is fabricated as follows (slightly modified from [1]), Ag silver ink will be deposited on DuPont Teijin Films™ (Melinex® ST505) polyester films using Western Michigan University's (WMU) MSP-485 screen printer (from Affiliated Manufacturers Inc.). Conductive Ag ink (Electrodag 7019) from Henkel is deposited as electrodes on Polyethylene terephthalate (PET) with 175  $\mu\text{m}$  thickness. The resulting Ag electrode prints is cured for 10 min at 107°C. Carbon ink

(Electrodag 440B) is screen-printed on the silver layer to protect the silver layer from electron migration. The patterns on the screen used for this layer are deliberately slightly larger than that of the silver pattern to make sure that the carbon layer completely covers the silver. The carbon layer is also cured for 10 min at 107°C. SonoPlot® GIX Microplotter II is used to fabricate the channels as pictured in Figure 113. 3M™ Novec™ 2702 Electronic Grade Coating fluorochemical acrylic polymer carried in a hydrofluoroether solvent as a non-wetting polymer will be used for this purpose. This polymer is characterized as a low viscosity and low surface tension solution and comes in clear aqueous solution. The deposited channels are then cure at 100 °C for 1 hr for maximum attachment. The PET sheets are then UVO treated and attached to 75×50 mm microscope slides. 0.2 ml of matrix solution with different concentration (Figure 113) is deposited in the channels using a pipette and allowed to dry. Adhesive tape is placed on the sides of the PET. Then a glass cover slip is placed on the adhesive tape leaving a small gap between the channels and the cover slip to control the amount of analyte which will later be introduced to the channels for the resistivity measurement (Figure 114). A 1"x3" glass cover slide is attached to the double stick adhesive spacer so the slide is positioned directly above the matrices (see Figure 115). This slide has eight parallel lines of non-wetting polymer that lie directly above the 8 parallel lines of non-wetting polymer on the PET below. This allows the liquid sample to migrate into the gap above each of the matrix layers. With the spacer providing a narrow air gap (4 mils or less depending on the spacer, <100,000 nm) above the matrices and the glass cover slide (see Figure 115 side view), a controlled volume of liquid sample can be applied that will precisely coat each matrix with test sample.



**Figure 113. Side view of a five-channel test plate (adapted from [1]).** The narrow gap between the bottom of the top glass slide and the top of the matrix for the five channels, through surface tension, draws the antigen containing test solution under the top glass slide when applied at the edge of the slide (see asterisk). The bottom of the top glass slide has lines of anti-wetting polymer strategically deposited to contain the applied liquid test sample over each matrix.



**Figure 114. Schematic of five-channel test plate (adapted from [1]) for testing matrix formulations.** Once connected to data acquisition system, the resistance of each matrix channel will be measured





#### **4.1.1.2 Test solution formulation**

Test solutions containing 5, 1, 0.3, 0.08, 0.02, 0.005 and 0 mg/ml of antibody I5006 were prepared in buffer solution. Different concentrations of antigens were achieved by the dilution of the stock antigen solution with buffer of the same ionic strength. All solutions had the same ion conductivity as determined by a 3 point conductivity probe to minimize any chance of misinterpreting any apparent trends or correlations that could result if test solutions were of different salt/ion concentrations. R2004 will specifically bind I5006. Therefore the test solution which contains I5006 will specifically bind to the R2004 antibody in the channels. However the channels to which the I5006 protein was added in the matrix should not exhibit any specific binding. The latter channels were used as the control channels to be compared with the other channels. Buffer solutions without the presence of protein were used to evaluate the effect of buffer alone on the results. This way, it was possible to account for the effect of the buffer on the results. A total of 7 test plates with 2 “I” channel and 3 “R” channel replicates for each test solution concentration were used in the study for a given matrix formulation.

#### **4.1.1.3 Characterization**

2-point resistance measurements (Keithley 2602 Dual-Channel System Source-Meter) using a constant current resistance measurement of 0.7 uA were used. The resistance results were plotted. Once the test solution, 30  $\mu$ L, was applied to the matrix-coated channels, the resistance values were recorded every 5 minutes and the change in the value of the resistance was correlated with the concentration of the antigen in the applied test solution. Analysis was performed to investigate the significance of the differences among the different concentrations of the antigen.

In this preliminary work, non-specific binding was shown to dominate the response and specific binding, if any, comparable to or less than inherent error in the resistance measurements.

#### **4.1.1.4 Further Work**

Future work could continue to explore antibody-antigen binding studies using this biosensor test-plate thin film matrix concept evaluated in the preliminary studies noted earlier but with CMC as the dispersant (rather than PSS) as it can be coated in more uniform coatings in a more predictable fashion compared to PSS as elucidated in the research reported in this thesis.

# OVERALL CONCLUSIONS

Certain biosensors working on the basis of conductance/impedance changes have demonstrated their potential to detect various bacteria, enzymes, and biomolecules due to their enhanced electrical properties with incorporated single-walled carbon nanotubes (SWNT). One of the most challenging aspects in the development, characterization and optimization of biosensor material formulations for immunodiagnostic applications is uniformly dispersing SWNTs within the sensing component. In this study, this challenge has been addressed through a series of experiments varying the concentration of poly(sodium 4-styrenesulfonate) (PSS) or carboxymethylcellulose (CMC), two dispersing agents, glycerol (a protein protective/stability agent), and antibody formulated relative to the concentration of a conductive SWNT (at three concentrations of 0.21%, 15% and 8.4% by weight SWNT) in a phosphate buffered aqueous solution. Various sonication protocols (duration and intensity-% amplitude) were also evaluated. Formulated matrix solutions were characterized using dynamic light scattering. Dried composite “matrix” films deposited on test plate templates were characterized using 3D optical microscopy. Resistivity was determined for each dried composite matrix film.

A total of 198 experiments (120 experiments in Phase 1 and 78 experiments in Phase 2) were designed using Response Surface Methodology (RSM) in order to model the resistance of dried thin films of matrix solution produced under various conditions. Dispersed SWNT and aggregated/bundled SWNT present after various sonication protocols were separated by centrifugation and microfiltration, and percent dispersed was determined by weight difference of dried samples. This information was augmented with

particle size distribution data using the dynamic light scattering technique. Once characterized, the SWNTs matrix solutions were deposited within test sensor channels, dried into a semi-solid/fluid film, and resistance tested using a constant microamp current. Test sensor plates used in this study were prepared using printing technology consisting of screen printing and material plotting over PET base substrates. The surface total free energy of the PET substrate used was increased from ~39 to 67 dyn/cm by UVO treatment for 5 minutes prior to use in these studies.

The purpose of the Phase 1 on this study was to identify the role of the dispersing agents (CMC and PSS) at different SWNT:dispersant ratios under various sonication protocols (intensity/amplitude and duration). It was found that run 15 matrix solution (SWNT:CMC=2, sonication amplitude=40%, sonication duration=24 hr) resulted in dried matrix films with the lowest resistance (128  $\Omega$ ) and possessed an average particle size of ~115 nm. In contrast, the largest particle size (where 81% of the particles have an average particle size of ~5000 nm), for the set of formulation experiments in which CMC was used, was observed in Run 3 (SWNT:CMC=0.5, Sonication amplitude=20%, and sonication duration= 2 hr) in which the highest dried film resistance was also obtained. From this it was concluded that a lower SWNT:CMC ratio (lower conductive:insulating components) is contributing to larger SWNT particle size that together are contributing to higher film resistivity. Higher concentration of dispersant in comparison with the SWNTs may result in the dispersant wrapping in multiple layers around SWNTs which hinders electron transport along the conductive carbon nanotubes. This may also create larger particles which results in the removal of SWNTs from the solution after filtration.

The sonication factor was found to play an important role in the dispersion and conductivity of SWNTs. The analysis of variance of the particle sizes obtained in the PSS-dispersed matrix solutions shows that the sonication amplitude and its interaction with the concentration of PSS are the only significant factors among all ( $\alpha=0.05$ ). Moreover, the sonication duration is the most significant factor in the analysis of variance of the particle sizes obtained in the CMC-dispersed matrix solutions (longer sonication, smaller particle size), although there are other interacting variables.

3D optical microscopy was performed on the dried matrix films to investigate the morphology, and roughness of the dried films as well as to measure the thickness of the coatings. Profilometry of Run 3 (CMC-dispersed coating) at 100% (no dilution) showed an average roughness of 0.274  $\mu\text{m}$  while profilometry of Run 15 at 100% (no dilution) showed an average roughness of 0.067  $\mu\text{m}$ . The same trend was noted for matrix coatings at higher dilutions (70% and 40%). Therefore, increased particle size of the matrix formulations had a strong correlation with increased roughness, and increased resistance of the CMC-dispersed coatings. Superior uniformity with low average roughness was observed for the coatings in which CMC was used as the dispersing agent in contrast to PSS.

The thickness of the dried matrix films as measured by 3D optical microscopy was found to decrease with the dilution of the matrix solution as would be expected due to reduced solids content. As an example, the thickness of matrix coatings obtained from Run 15 of the formulation in which CMC was used as the dispersant at 100% (no dilution), 70% dilution, and 40% dilution was measured to be 3.86, 2.81, and 1.63  $\mu\text{m}$ , respectively. Mass balance of each prepared solution was performed by determining the weight difference of

the solids before and after the centrifugation and filtration and the thickness of the coatings after drying were calculated based on solids content, volume applied and area. The results were compared with thickness measurements by the 3D optical microscope. The calculated values were found to be very close to the experimentally measured thickness measurements.

Based on the experimentally-determined higher quality and superior workability of the formulations and resulting dried matrix films, it was concluded that CMC rather than PSS was more consistent and should be used as the dispersant in the matrix formulations developed in the 2<sup>nd</sup> phase (chapter 3) of this dissertation. The base matrix solution formulations carried forward to the 2<sup>nd</sup> research phase achieved low resistance for each SWNT to CMC ratio studied. Matrix formulation protocols from Run 14 (SWNT:CMC = 0.5), Run 17 (SWNT:CMC = 1.25), and Run 12 (SWNT:CMC = 2) were selected and labeled as CMC-14, CMC-17, and CMC-12, respectively. The corresponding resistance results for the dried matrices were 1069, 794, and 148  $\Omega$ , respectively. 78 different solutions were prepared using the CMC-14, CMC-17, and CMC-12 base matrix formulations based on an experimental design containing various SWNT:glycerol and SWNT:antibody ratios. Their resistance responses as well as surface morphologies of the dried matrix films were investigated using profilometry. The resistance responses of these antibody/glycerol-containing dried matrix films were modeled using the RSM design. The resistance behavior of coatings prepared from CMC-12 solution were found to be statistically more predictable when compared to that of the coatings prepared from solutions CMC-14 and CMC-17. 3D optical microscopy showed that the thickness of the coatings increased with a decrease in the glycerol concentration. The uniformity of the

coatings prepared from CMC-12 matrix solutions were found to be the highest among the different matrix solutions tested and the superior uniformity resulted in lower resistance.

Among the three different ratio variations of SWNT:Glycerol studied with CMC-12, the highest 1:1 ratio of the three ratios tested was found to lead to the lowest electrical resistance. Glycerol at the ratio of SWNT:Glycerol of 1:1 can be used as a stabilizer to perhaps preserve the activity of antibody after drying. Although the addition of antibody in the system results in an increase in the resistance of the matrix coating, a concentration of antibody, as high as SWNT:Antibody ratio of 10, can be selected while maintaining relative low resistance (below 2000  $\Omega$  for the geometry used in this research), with no dilution of the formulation. These levels of antibody and glycerol together with a base matrix formulation using the CMC dispersant at the ratio of SWNT:CMC=2 and the sonication amplitude of 40% for 2 hours was found to give the best properties. It is hoped that information from this research will be helpful in the design of immunodiagnostic biosensors relying on antibody-SWNT design elements.

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**Conference Proceedings:** Photovoltaic Specialists Conference, 2005. Conference Record of the Thirty-first IEEE

**Author:** J. Sakai; E. Fujinaka; T. Nishimori; N. Ito; J. Adachi; S. Nagano; K. Murakami

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**Title:** A study of the off-contact screen printing process. I. Model of the printing process and some results derived from experiments

**Author:** J. A. Owczarek; F. L. Howland

**Publication:** Components, Hybrids, and Manufacturing Technology, IEEE Transactions on

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